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PERMEATION DISPERSAL OF TREATMENT AGENTS FOR IN SITU REMEDIATION IN LOW PERMEABILITY MEDIA: 1. FIELD STUDIES IN UNCONFINED TEST CELLS

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TABLE OF CONTENTS

		FIGURES	
LIS	ST OI	TABLES	iv
AB	BRE	VIATIONS, ACRONYMS, AND INITIALISMS	yi Yi
EX	ECU	TIVE SUMMARY	viii
AC	KNU	WLEDGMENTS	AIII
	.		1_1
1.		duction	1-2
	1.1	Purpose and Scope	1-4
	1.2	Organization	đ. i.
2.	Back	eground	2-1
	2.1	Facility Description	2-1
	2.2	Site Description	2-1
	2.3	Site Geology as Defined by CTS Characterization	2-1
	2.4	Site Geology as Defined by Pre-Treatment Characterization	2-2
3.	Tecl	mical Approach	3-1
	3.1	Task 1: Geoprobe Borings and Lysimeter Installation	3-1
	3.2	Task 2: MPIS Testing	3-3
,	3.3	Task 3: Concurrent Monitoring	3-5
	3.4	Task 4: Post-Injection Characterization and Data Collection	3-5
	3.5	Surveying	3-6
		to must the term	4_1
4.		cific Testing and Results	4-1
	4.1	Background CTS Characterization	4-1
		4.1.1 Field Activities	
		4.1.2 Pre-Treatment Soil Testing	4-2
•		4.1.3 Background water results	4_3
	4.2	Test Cell 1: Water Injection with Tracers 4.2.1 T1 Field Activities	4.3
		4.2.1 11 Field Activities	4_5
		4.2.2 T1 Soil Sample Results	4-6
		4.2.3 11 Water Duniple Research	4-7
	4.3	Test Cell 2: Hydrogen Peroxide	
		4.3.1 T2 Field Activities	
		4.3.2 T2 Soil Sample Results	ر-ب 4-11
•		4.5.5 12 Water Sample Results	 4-11
	4.4	Test Cen 3. Bionutricite with Trucors	4- 11 4-11
		4.4.1 T3 Field Activities	1-TT

CONTENTS (continued)

		4.4.2 T3 Soil Sample Results	4-13
		4.4.3 T3 Water Sample Results	4-14
	4.5	Test Cell 4: Lime Slurry	4-15
		4.5.1 T4 Field Activities	4-15
		4.5.2 T4 Soil Sample Results	4-17
		4.5.3 T4 Water Sample Results	4-17
	4.6	Test Cell 5: Potassium Permanganate	4-18
		4.6.1 T5 Field Activities	4-18
		4.6.2 T5 Soil Sample Results	
		4.6.3 T5 Water Sample Results	4-20
	4.7	Test Cell 6: Air	
		4.7.1 T6 Field Activities	4-21
		4.7.2 T6 Soil Sample Results	4-22
		4.7.3 T6 Water Sample Results	
	4.8	Test Cell 7: Micropowder Iron	
		4.8.1 T7 Field Activities	4-22
		4.8.2 T7 Soil Sample Results	4-24
		4.8.3 T7 Water Sample Results	4-24
5.	Equ	ipment Operations Observations	. 5-1
6	Sum	nmary and Conclusions	. 6-1
Ο.	Juli	mining with College State Coll	,
7.	Refe	erences	. 7-1
AT	MITA I	DIX A - GEOTECHNICAL, BACKGROUND, AND GRAIN SIZE	
		BUTION RESULTS FOR THE CTS	
ΑI	PPEN	DIX B - LITHOLOGIC LOGS FOR THE TEST CELL SOIL BORINGS	
	, 		
		DIX C - DATA TREND GRAPHS FOR VARIOUS GEOCHEMICAL	
PA	ARAN	METERS IN SOIL AND SOIL-PORE WATER	

LIST OF FIGURES

2.1	Location of Portsmouth Gaseous Diffusion Plant	2-4
2.2	Location of the CTS	2-5
2.3	Location of the CTS Overview of the test site with the DOE plant in the background	2-6
2.4	Topography of the CTS	2-7
2.5	Topography and borehole locations at the CTS Topography and borehole locations at the CTS	2-8
2.6	Photographs of the CTS subsurface silty-clay media	2-9
2.7	Stratigraphic sequence of borehole number BH07	2-10
2.8	Geologic cross sections at the CTS	2-11
3.1	lest cells within the C15	3-1
3.2	Approximate locations for testing within a typical test cell	3-8
3.3	Schematic diagram of nested lysimeters	3-9
3.4	Schematic diagram of piezometer	3-10
3.5	Profile view of the layout for the instrumented borings	3-11
3.6	Conceptual diagram of the MPIS process	3-12
3.7	Multi-port injection system	3-13
3.8	Photographs of the Hayward Baker MPIS rig and the monitoring and	
- 7 -	sampling analyses activities	3-14
3.9	Photographs of intact soil cores from the permanganate (T5) and iron	
	micropowder (T7) test cells	3-15
4.1	Test cell 1 sample locations	4-26
4.2	Test cell 1 - post-treatment soil moisture content	4-27
4.3	Test cell 1 - post-treatment soil bromide levels	
4.4	Test cell 1 - post-treatment soil INA levels	4-28
4.5	Test cell 1 - post-treatment water bromide levels	4-28
4.6	Test cell 2 sample locations	4-29
4.7	Test cell 2 - post-treatment soil peroxide levels	4-30
4.8	Test cell 2 - post-treatment soil Eh data	4-30
4.9	Test cell 2 - post-treatment water DO levels	4-31
4.10	Test cell 2 - post-treatment water peroxide levels	4-31
4.11	Test cell 3 sample locations	4-32
4.12	Test cell 3 - post-treatment soil bromide levels	4-33
4.13	Test cell 3 - post-treatment soil INA levels	4-33
4.14	Test cell 3 - post-treatment water pH	4-34
4.15	Test cell 3 - post-treatment water bromide data	4-34
4.16	Test cell 4 sample locations	4-35
4.17	Test cell 4 - post-treatment soil pH	4-36
4.18	Test cell 4 - post-treatment soil calcium levels	4-36
4.19	Test cell 4 - post-treatment water pH	4-37
4.20	Test cell 4 - post-treatment water alkalinity levels	4-37
4.21	Test cell 4 - post-treatment water conductivity levels	4-38
4.22	Test cell 5 sample locations	4-39

FIGURES (CONTINUED)

4 23	Test cell 5 - post-treatment soil manganese levels	4-40
4 24	Test cell 5 - post-treatment soil Eh levels	4-40
4.25	Test cell 5 - post-treatment water manganese levels	4-41
4.26	Test cell 6 sample locations	4-42
4.27	Test cell 7 sample locations	4-43
4.28	Test cell 7 - post-treatment water iron data	4-44

LIST OF TABLES

1.1	Field activities and analyses listed for a typical test cell for MPIS testing	1-5
2.1	Ground surface and bedrock surface elevations for CTS boreholes	2-12
2.2	Subsurface properties of the Minford Member at the CTS	2-13
3.1	ORNL SOPs for the MPIS demonstration	3-16
3.2	Activities performed in conjunction with MPIS testing	3-17
3.3	Test cell design and characteristics	3-19
3.4	Treatment agents for MPIS testing at PORTS CTS	3-20
3.5	Process operating conditions for MPIS testing at the CTS	3-21
4.1	Summary of background soil results	4-45
4.2	Water sample results from background piezometer and lysimeters	4-46
4.3	Tracer Test cell 1 (T1), soil sample results	4-48
4.4	Water sample results for the tracer test cell (T1), injection date 11/12/95	4-49
4.5	Hydrogen peroxide test cell (T2), soil sample results	4-51
4.6	Water sample results for the hydrogen peroxide test cell (T2), injection	
	date 11/15/95	4-52
4.7	Bionutrient test cell (T3), soil sample results	4-54
4.8	Bionutrient test cell (T3) heterotrophic enumerations, cells/g	4-55
4.9	Water sample results for the bionutrient test cell (T3), injection	
	date 11/18/95	4-56
4.10	Lime slurry test cell (T4), soil sample results	4-58
4.11	Water sample results for the lime slurry test cell (T4), injection date 11/13/95	4-59
4.12	Potassium permanganate test cell (T5), soil sample results	4-61
4.13	Water sample results for the potassium permanganate test cell (T5), injection	
	date11/16/94	4-62
4.14	Water sample results for air test cell (T6), injection date 11/19/94	4-64
4.15	Iron micropowder test cell (T7), soil sample results	4-65
4.16	Water sample results for iron micropowder test cell (T7), injection date	
	11/19/94	4-66
5.1	Task summary for MPIS testing	. 5-3

VIII

ABBREVIATIONS, ACRONYMS, AND INITIALISMS

bgs below ground surface CSM Colorado School of Mines

CTS Clean Test Site

d day

DNAPL dense nonaqueous-phase liquids

DO dissolved oxygen

DOE U.S. Department of Energy ESD Environmental Sciences Division

ft feet or foot g gram gal gallon

GJ Grand Junction ID inside diameter

in inch

INA ice-nucleating activity

kg kilogram L liter

LPM low permeability media

mg milligram

LMES Lockheed Martin Energy Systems, Inc.

MPIS multi-port injection system MPN most probable number

msl mean sea level mV millivolt

ORNL Oak Ridge National Laboratory
OST Office of Science and Technology

oz. ounce

PCE perchloroethylene
PE polyethylene

PORTS Portsmouth Gaseous Diffusion Plant

ppm parts per million

psi pounds per square inch PVC polyvinyl chloride

s second

SEM scanning electron microscope
SMT soil moisture and temperature
SOPs standard operating procedures

TBD to be determined TCE trichloroethylene TOC total organic carbon

U.S. United States

USEC United States Enrichment Corporation

EXECUTIVE SUMMARY

A research and demonstration project was completed to evaluate the viability of permeation dispersal as a delivery method for delivering treatment agents to enable in situ remediation of contaminated low permeability media soils. This report describes field-scale testing using unconfined test cells established at the Clean Test Site (CTS) at the DOE Portsmouth Gaseous Diffusion Plant near Piketon, Ohio. At this site a commercially available multi-port injection system (MPIS) was used to deliver contrasting agents into a silty clay deposit. A companion report describes laboratory studies completed at the Colorado School of Mines where intact soil cores collected from the CTS were injected with two chemical oxidants (hydrogen peroxide and potassium permanganate) (see Urynowicz and Siegrist, 2000).

At the CTS, seven unconfined test cells were established to enable evaluation of seven treatment agents. The treatment agents tested were selected based on their contrasting features and potential applicability to treatment of organic chemicals in the subsurface. Water amended with tracers was injected into one test cell (T1) to provide a reference for nonreactive fluid movement in the silty-clay soil. The treatment agents studied included: hydrogen peroxide (T2) and potassium permanganate (T5) as chemical oxidants capable of degradation of many toxic organics; a guar gum suspension of iron micropowder (T7) for reductive dechlorination; a lime slurry (T4) to elevate pH and cause alkaline destruction of organics as well as stabilize metals; a bionutrient/surfactant agent (T3) to enhance biodegradation; and compressed air (T6) to enhance permeability and aeration status and improve volatilization or biodegradation. Six of the test cells were 24 ft by 24 ft in surface area while one was 4 ft by 8 ft. All cells were injected with treatment agents at an average volumetric loading of 0.27 gal/ft³ to a depth of 10.4 ft below ground surface using the MPIS with four rack-mounted injectors on 2-ft spacings. Extensive monitoring during injection and sampling and analysis before and after injection were completed to evaluate the effectiveness of MPIS delivery and the impacts of the various agents on the ambient subsurface conditions.

Field test results revealed that treatment agents can be rapidly delivered into the shallow subsurface using MPIS technology. The treatment agents injected appear to rapidly advect away from the injector bore in existing pathways such as root channels and fractures. For those agents that are persistent and react slowly in the subsurface, further dispersal can occur by slower advection in fine matrix pores as well as by diffusive transport processes. The initial dispersal of the treatment agents was similar to that of the conservative tracers. With time after the injections were completed, there were expanding effects in the subsurface within the lime, potassium permanganate and iron cells. Evaluation of injection rates, equipment and labor costs indicate that MPIS technology may be applied for \$2 to \$3 per cubic yard of soil not including the cost of reagents.

ACKNOWLEDGMENTS

This report on the demonstration of permeation and dispersal of reactive fluids for in situ treatment in low permeability media at the U.S. Department of Energy (DOE) Portsmouth Gaseous Diffusion Plant (PORTS) is based on the combined efforts of scientific and engineering staff at Oak Ridge National Laboratory (ORNL) and Hayward Baker Environmental in collaboration with the Colorado School of Mines (CSM) and others. Project sponsorship was provided through the Subsurface Contaminant Focus Area of the DOE Office of Science & Technology along with the DOE Office of Environmental Restoration at PORTS.

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1. Introduction

Chlorocarbons like trichloroethylene (TCE) are common contaminants of concern at U. S. Department of Energy (DOE) facilities and industrial sites across the United States (U.S.) and abroad (Huling and Weaver 1991; U.S.EPA 1992; MacDonald and Kavanaugh 1994). These contaminants of concern are present in source areas and in soil and ground water plumes as dissolved or sorbed phase constituents as well as dense nonaqueous-phase liquids (DNAPLs). These DNAPL compounds can be released to the environment through a variety of means including leaks in storage tanks and transfer lines, spills during transportation, and land treatment of wastes. When DNAPL compounds are present in low permeability media (LPM) like silt and clay layers or deposits, there are major challenges with assessment of their behavior and implementation of effective in situ remediation technologies.

In situ remediation technology development has largely overlooked treatment of DNAPLs in LPM. Poor accessibility to the contaminants and the difficulty in delivery of treatment agents have rendered conventional bioremediation, vapor extraction, and pump-and-treat ineffective for this type of media. As a result, effective in situ treatment methods for DNAPL compounds in fine-grained deposits was recently one of the top-ranked environmental restoration needs across the DOE Complex. Similarly, within the petroleum industry, nearly 40% of the underground storage tanks in the world are located on clay soils and remediation of contaminants from leaking underground tanks in these settings has been a major challenge.

As a result of the need for solutions and the gap in the current knowledge and technology base, a project was initiated by the DOE Office of Science and Technology (OST) and the DOE Portsmouth Gaseous Diffusion Plant (PORTS) near Piketon, Ohio in collaboration with the American Petroleum Institute (API) in 1993 (API 1995, DOE 1996). In this project, in situ remediation technologies are being evaluated for both enhanced mass removal and in place destruction of DNAPL compounds in LPM, specifically chlorinated solvents [e.g., TCE, perchloroethylene (PCE)] in the vadose and saturated zones of LPM. The overall project has included a series of related tasks including: (1) preparation of 16 DNAPL focus papers and reports, (2) a field pilot test of hydraulic fracturing for dewatering, (3) a field test of enhanced air flushing for NAPL removal, (4) a field test of hydraulic fractures for hydraulic and pneumatic control and hot fluid injection, (5) a field comparison of multiple point injection and permeation dispersal of different reactants, (6) a field-scale demonstration of soil fracturing for thermally enhanced mass recovery and reactive barrier degradation, and (7) numerical and experimental analyses of the mobility of residual NAPLs versus varying degrees of remediation. The field testing activities have occurred at both clean and contaminated sites in the U.S. and Canada.

This report describes a field demonstration that was conducted at the PORTS Clean Test Site (CTS) to evaluate the feasibility of permeation and dispersal of reagents into LPM. Various reagents and tracers were injected at seven test cells primarily to evaluate the feasibility of

delivery, but also to evaluate the effects of the injected reagents on LPM. The various reagents and tracers were injected at the PORTS CTS using a multi-port injection system (MPIS) developed and provided by Hayward Baker Environmental, Inc. The work reported here was performed by personnel from Oak Ridge National Laboratory (ORNL) in collaboration with Lockheed Martin Energy Systems [(LMES), now Bechtel Jacobs Company, LLC], Hayward Baker Environmental, Inc., and others.

Supplementary laboratory experiments at the Colorado School of Mines focused on the effects of chemical oxidants on LPM using intact cores collected from the CTS. The results of the laboratory experiments are presented in the companion report, *Permeation Dispersal of Treatment Agents for In Situ Remediation in Low Permeability Media: 2. Laboratory Studies with Intact Cores* (Urynowicz and Siegrist 1999). Other facets of the project have been focused on the reaction mechanisms of reagents used in this project and alternative reagent delivery methods and the results have been reported elsewhere (e.g., API 1995; DOE 1996; Pfiffner et al. 1997; Case 1997; Gates and Siegrist 1995; Gierke et al. 1995; Murdoch et al. 1997a; Murdoch et al. 1997b; Siegrist et al. 1993; Siegrist et al. 1995c; Siegrist et al. 1995c; Siegrist et al. 1996; Siegrist et al. 1995; Siegrist et al. 1995; Strong-Gunderson and Palumbo 1995; Walden 1993; West et al. 1995).

1.1 Purpose and Scope

The purpose of this project was to test an innovative approach for in situ treatment of volatile organic compounds, which are common contaminants at DOE facilities. The PORTS CTS was chosen as an appropriate location for determining the feasibility of performing multi-point injection permeation and dispersal testing because it is representative of the low permeability soils contaminated at many DOE sites. Additionally, the CTS enabled evaluation of the feasibility of the process but did not present the complications of contaminated site operations.

The various reagents and tracers injected included water, oxidants, a bionutrient, lime, air, and zero-valence iron. Water alone was injected to provide a baseline test cell. The oxidants (hydrogen peroxide and potassium permanganate) were injected because they can catalyze and chemically oxidize chlorinated organics. The bionutrient and surfactant were injected to determine the feasibility of using the mixture to enhance the natural breakdown of chlorinated solvents by indigenous bacteria. The lime injection was tested because it could be used to adjust soil pH for in situ stabilization of metals. Air was injected to test the MPIS for pneumatic fracturing of LPM and zero-valence iron (iron micropowder) was injected because it has been demonstrated to act as a reducing agent for chlorinated solvents.

The field work for this project included four tasks: (1) pre-treatment characterization of the lithology and soil conditions within seven test cells at the CTS, (2) MPIS injection and testing using various reagents and tracers at seven test cells, (3) concurrent monitoring during MPIS testing, and (4) post-treatment sampling and monitoring of the seven test cells. Several post-treatment sampling events were performed.

Samples collected during this project included soil samples for logging lithology, soil fracture characteristics and geochemical properties, and soil-pore water samples for water quality parameters and geochemical properties. Field and laboratory analysis was performed on soil samples and soil-pore water samples as indicated on Table 1.1.

The following aspects of the permeation and dispersal were evaluated during this project:

- Feasibility of using a MPIS for injection of treatment agents into the subsurface. The agents were various solutions, slurries, or emulsions of compounds that may reduce toxicity through treatment or containment, and
- Relative effectiveness of the treatment agents for in situ remediation in silty clay soils.

The general objectives of the testing and the procedures followed to achieve them are listed below.

Objective 1: To characterize fracture size and continuity in the untreated soil and determine changes in the soil after reagent injection. Pre-treatment soil samples were collected from soil borings to describe fracture morphology and determine baseline geochemical properties. Tracers were injected with the treatment agents, where compatible, in order to facilitate detection of the injected fluids and the fractures that transmitted them. Soil-pore water samples were collected from suction lysimeters to evaluate the dispersal of the injected fluids. Soil moisture probes were emplaced to monitor the increase in soil moisture due to injections. Soil samples from post-treatment borings were inspected to determine changes in fracture size and density and geochemical properties. At one location, a test pit was excavated to permit visual examination of the soil to determine if the effects of the injection technique were visible in the subsurface.

Objective 2: To determine matrix effects of the various fluids released with respect to changes in soil-pore water and soil. Suction lysimeters provided soil-pore water samples that were monitored for changes in temperature, pH, dissolved oxygen (DO), conductivity, total organic carbon (TOC), alkalinity, Fe, Mn, Cl⁻, No₃ and SO₄². Soil moisture probes were monitored to determine the relative dispersal of the injected fluids. Soil samples were collected for evaluation of matrix effects and detection of injected fluids.

Objective 3: To determine dispersal of reactive particles in LPM. An emulsion or slurry of reactive particles (e.g., iron micropowder with guar gum) was injected into the subsurface using the MPIS. The area of influence was evaluated by collecting post-treatment core samples. These cores were macroscopically inspected and chemically analyzed to determine the presence, concentration, and distribution of the reactive particles and/or tracers. Soil-pore water samples collected from suction lysimeters also provided information on dispersal of reagents.

Objective 4: To determine dispersal of oxidants in LPM. Solutions of hydrogen peroxide and potassium permanganate were injected. The area of influence was determined by collecting core samples and by collecting soil-pore water samples. These cores and water samples were analyzed to determine the effects of the reactive fluids.

Objective 5: To determine the operation and maintenance characteristics of the MPIS equipment. Observations of injection volumes, flow rates, back pressure, operational problems, etc., were documented during field tests to evaluate multi-point injection performance.

1.2 Organization of the Report

The remainder of this report is organized as follows:

- Section 2 describes the CTS.
- Section 3 describes the technical approach for this project.
- Section 4 discusses the testing performed in each test cell and the soil and water sampling results.
- Section 5 discusses observations made related to equipment operation.
- Section 6 is the project summary.
- Section 7 discusses application of the technique to a contaminated site.

Table 1.1. Field activities and analyses for a typical test cell for MPIS testing

Pre-treatment activities	Testing during injection	Post-treatment No. 1 activities	Post-treatment No. 2 activities
Soil Analyses: Five soil borings per test cell:% moisture, grain size color, mineralogy, SEM, X-ray, pH, Eh, TOC, Fe oxides, cations, anions, bacteria	None	Five soil borings per test cell: % moisture, grain size, color, mineralogy, SEM, X-ray, pH, Eh, TOC, Fe oxides, cations, anions, bacteria, visual inspection	Two soil borings per test cell: % moisture, grain size, color, mineralogy, SEM, X-ray, pH, Eh, TOC, Fe oxides, cations, anions, bacteria
Soil-Pore Water Analyses:	Water injection with tracers: monitored penetration rate, flow rate, pressure, and reagent concentration		
Three lysimeters per test cell: temp., pH, DO, conductivity, TOC, alkalinity, Fe, Mn, Cl ⁻ , NO ₃ ⁻ , SO ₄ ²⁻ , Br	Lysimeters: temp., pH, DO, conductivity, TOC, alkalinity, Fe, Mn, Cl, NO ₃ , SO ₄ ² , Br	Lysimeters: temp., pH, DO, conductivity, TOC, alkalinity, Fe, Mn, Cl, NO ₃ , SO ₄ ² , Br	Lysimeters: temp., pH, DO, conductivity, TOC, alkalinity, Fe, Mn, Cl, NO ₃ , SO ₄ ² , Br
One piezometer per test cell: temp., pH, DO, conductivity, TOC, alkalinity, Fe, Mn, Cl, NO ₃ , SO ₄ ² , Br	Piezometer: monitored water levels	Piezometer: temp., pH, DO, conductivity, TOC, alkalinity, Fe, Mn, Cl, NO ₃ , SO ₄ ² , Br	Piezometer: temp., pH, DO, conductivity, TOC, alkalinity, Fe, Mn, Cl ⁻ , NO ₃ ⁻ , SO ₄ ² -, Br

Note: Not all samples were analyzed for all constituents listed. For example, only samples from one soil boring per test cell were analyzed for % moisture.

SEM = scanning electron microscope TOC = total organic carbon

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2. Background

2.1 Facility Description

PORTS is a federal facility owned by DOE and operated under a contract with United States Enrichment Corporation (USEC). LMES (now Bechtel Jacobs Company, LLC) performs DOE-required environmental restoration, waste management, and site management functions. The 3,714-acre federal reservation lies in Pike County, Ohio, between the cities of Chillicothe and Portsmouth, approximately 70 miles south of Columbus, Ohio (Fig. 2.1).

Operating since 1954, PORTS enriches uranium for commercial nuclear reactors. The enrichment process uses molecular diffusion techniques to separate the ²³⁵U isotope from the ²³⁸U isotope. The plant has an extensive support complex of machine shops, laboratories, utilities, and decontamination facilities. As a result of plant operations, PORTS generates a wide variety of wastes, including low-level radioactive wastes, spent solvents, polychlorinated biphenyl-contaminated oils, electroplating wastes, paint wastes, metal sludges, acids, and caustics.

2.2 Site Description

The CTS is located at the south end of PORTS, north of the intersection of Hewes Street and Perimeter Road (Figs. 2.2 and 2.3). The site lies outside the plant security fence and is in an uncontaminated area. The topography of the CTS is relatively level, with a drainage area along the western boundary and a rise of higher ground along the eastern boundary of the site (Fig. 2.4). The high point of the area is located at the northeast corner, at an approximate elevation of 667 ft above mean sea level (msl). A storm-water retention pond is located approximately 75 ft west of the northwest corner of the site, but is downgradient with respect to both surface and groundwater and does not appear to affect CTS conditions.

2.3 Site Geology as Defined by CTS Characterization

During the spring of 1994, the ORNL Grand Junction, Colorado (ORNL-GJ) characterized the geology of the CTS by installing nine boreholes to bedrock (BH01 through BH09) at a depth of approximately 30 ft below ground surface (bgs) and collecting soil samples with a hollow-stem-auger drilling rig and a GeoProbeTM rig (Fig. 2.5). The ORNL Environmental Sciences Division performed a ground-penetrating radar survey to delineate the bedrock surface. The geologic data collected from this site characterization were used to construct a bedrock surface map.

Drilling was accomplished using an all-terrain, CME 55 drill rig. The rig used hollow-stem augers with a 3-in.-outside-diameter by 5-ft-long continuous sampler that ran a few centimeters ahead of the lead auger to obtain undisturbed soil samples for lithologic logging. An average of three soil samples per hole were collected and analyzed for moisture content, liquid limit, plastic limit, and grain size. In the summer, an additional six borings (BH10 through BH15) were drilled south of the CTS to characterize the geology in that area prior to the installation of horizontal wells. A detailed lithologic log was prepared for each hole; the continuous samples obtained from the hole were labeled and archived for future use. Lithologic logs and the results of the geotechnical analyses are provided in Appendix A. Table 2.1 summarizes the surface and bedrock elevation data for all of the borings. Table 2.2 summarizes the subsurface properties at the CTS and shows the properties determined during this study.

The soil above bedrock at the CTS is composed of unconsolidated Quaternary fluvial and lacustrine deposits of the Teays Formation. Figures 2.6 and 2.7 are examples of the soil cores and stratigraphic sequence. These deposits are characterized by 15 to 22 ft of low permeability clays and silts known as the Minford Member overlying 2 to 6 ft of moderately permeable sandy gravels, gravelly sands, and silty sands known as the Gallia Member. The bedrock underlying the Quaternary deposits is composed of Mississippian-age Sunbury Shale and Berea Sandstone and Shale. The Sunbury is a very low permeability shale unit underlying the saturated alluvium at the site. Figure 2.8 shows cross sections of the geology at the CTS.

The Gallia forms the principal water-bearing unit in the alluvium at the site. Water-level measurements taken during the drilling project indicate that groundwater in the Gallia, and perhaps the lower portion of the Minford, is confined by the overlying clays and silty clays. When the confining layers were penetrated by augers, the groundwater level rose an average of 13.5 ft above the depth at which saturated sediments were first encountered during drilling (14 ft to 24.5 ft bgs).

2.4 Site Geology as Defined by Pre-Treatment Characterization

Thirty-two soil borings were completed within the CTS prior to the start of injection testing. The borings were drilled to a depth of 12 ft and were sampled continuously with a Geoprobe megabore sampler. Lithologic logs were prepared describing the soil characteristics (Appendix B). The most notable characteristics included:

- the presence of high-angle fractures in the silty clay,
- very fine laminar bedding in some sediments with multi-colored banding,
- a zone of thin-bedded gypsum (less than 1 cm thick) apparent across the site at depths between 6.5 to 8 ft bgs,

- increasing clay content with depth, and
- increasing resistance to penetration with depth.

Based on these characteristics, the upper part of the Minford appears to be primarily composed of shallow lake bed (lacustrine) sediments. There were periods during deposition when the lake dried up, thus forming the thin gypsum beds and desiccation mud cracks that were observed in the finely laminated sediments. This interpretation is consistent with the general description of the Minford given above.

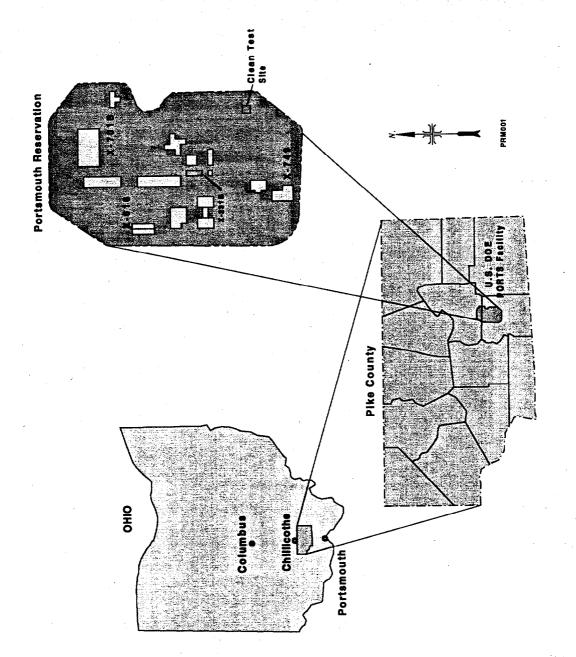
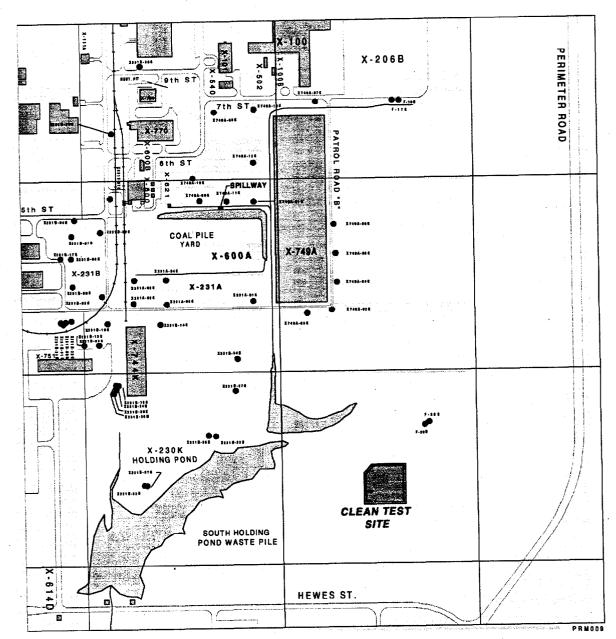


Fig. 2.1. Location of the Portsmouth Gaseous Diffusion Plant.



Portsmouth Facility

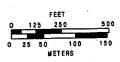
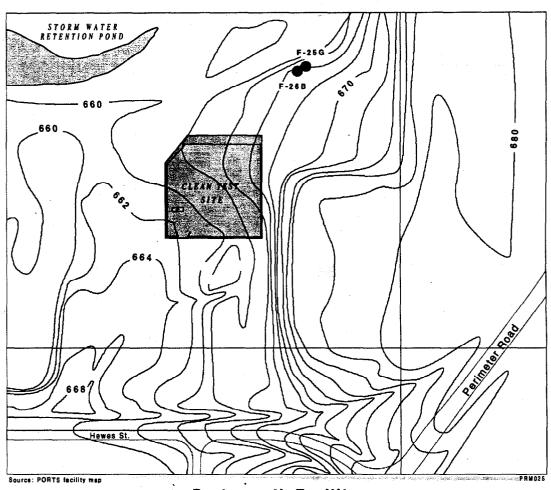




Fig. 2.2. Location of the CTS.



Overview of the test site with the DOE plant in the background.



Portsmouth Facility

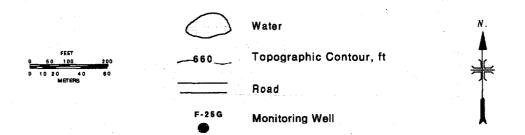


Fig. 2.4. Topography of the CTS.

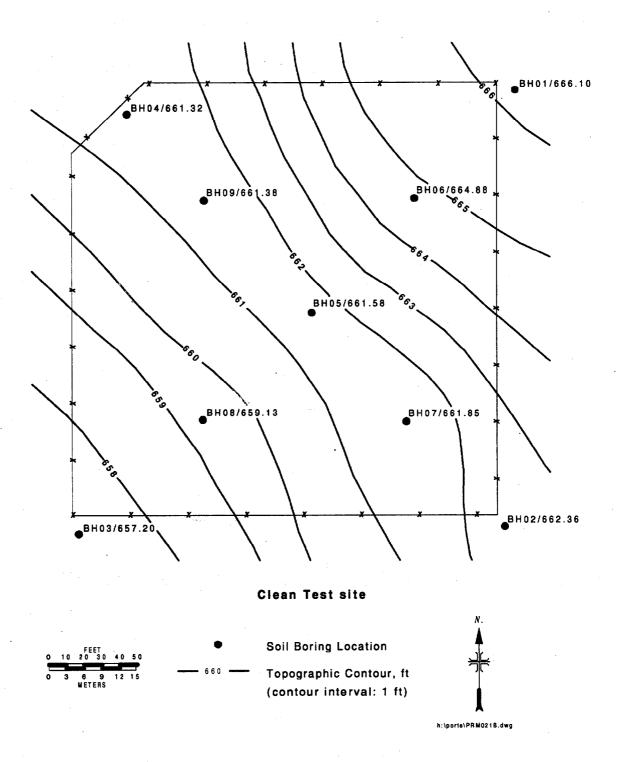


Fig. 2.5. Topography and borehole locations at the CTS.



- a. Soil core sample from approx. 2-ft depth.
- **b.** Soil core sample from approx. 10-ft depth.

Fig. 2.6. Photographs of the CTS subsurface silty-clay media.

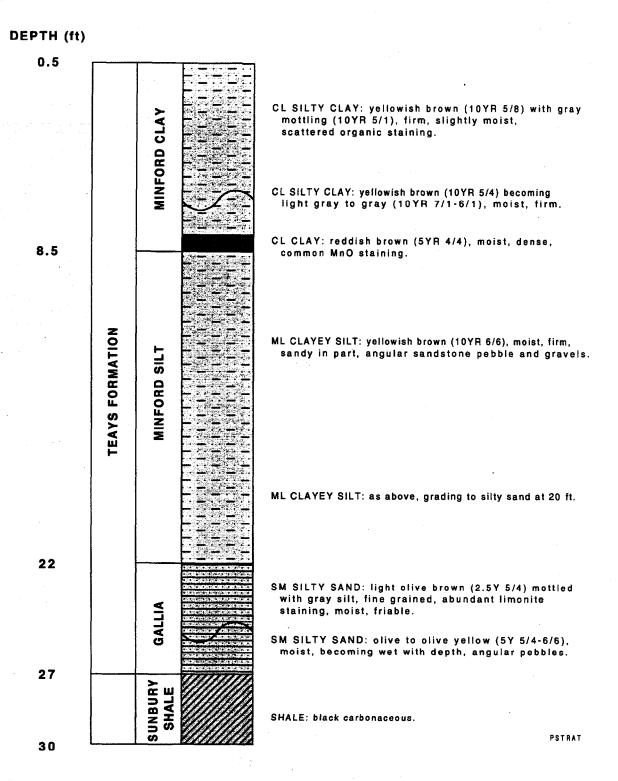


Fig. 2.7. Stratigraphic sequence of borehole number BH07.

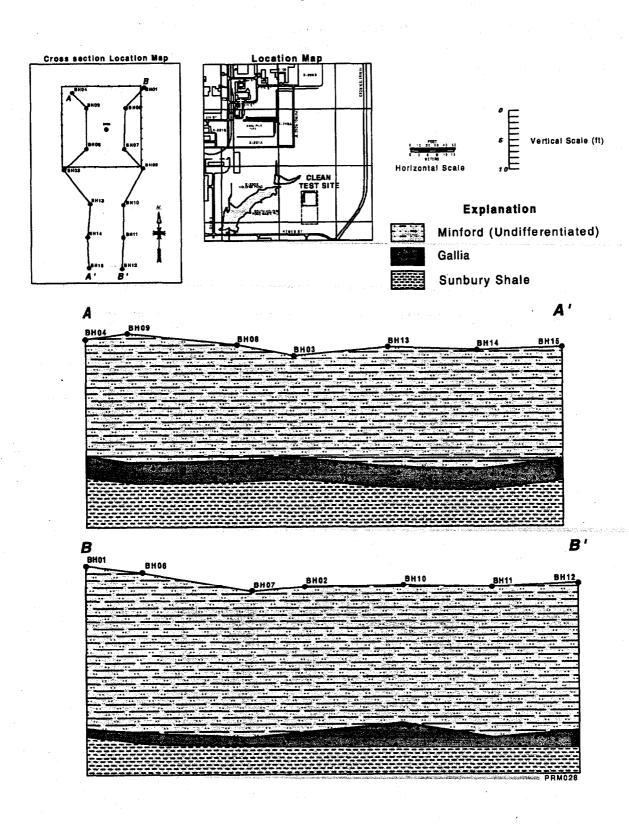


Fig. 2.8. Geologic cross sections at the CTS.

Table 2.1. Ground surface and bedrock surface elevations for CTS boreholes

Borehole number ^a	Ground elevation, ft msl	Depth to bedrock, ft bgs	Bedrock elevation, ft msl
1	666.10	29.5	636.60
2	662.36	28.5	633.86
3	657.20	21.5	635.70
4	661.32	24.5	636.86
5	661.58	26.0	635.58
6	664.88	29.7	635.18
7	661.85	27.0	634.85
8	659.13	24.0	635.13
9	661.38	26.5	634.88
10	662.70	28.3	634.40
11	662.73	28.8	633.93
12	663.07	29.1	633.97
13	658.80	24.5	634.30
14	658.12	24.5	633.62
15	658.88	24.0	634.88

^a See Figs. 2.5 and 2.8 for borehole locations.

msl = mean sea level

bgs = below ground surface

Table 2.2. Subsurface properties of the Minford Member at the CTS

Soil type and genesis	Silty-clay deposits of fluvio-lacustrine origin. Typically 15-ft-thick upper clay unit (CH) transitioning to a lower 10-ft-thick silt unit (CL).
Soil particle-size distribution:	en er
Sand size (0.050 to 2.000 mm)	~0.5 dry weight %
Silt size (0.002 to 0.050 mm)	~8.5 dry weight %
Clay size (< 0.002 mm)	~10 dry weight %
Soil mineralogy	In the Minford clay unit, the sand fraction consists of mainly quartz with minor goethite. The silt fraction consists of quartz and minor feldspars, but no goethite. The clay fraction is a mixture of illite (~33%), quartz (~29%), kaolinite (~26%), and smectite (~12%).
Soil physical properties:	
Bulk density	1.8 g/cm ³
Water content	20 weight %
Liquid limit	~60%
Plastic index	~35%
Soil-pore system:	o diagnostic productivamento de la merio describiro de la compansión de describiro de la compansión de la comp Compansión de la compansión
Total fractional porosity a	0.40 v/v
Water-filled saturation	90% pores
Pore water saturation	10% pores
Soil Chemistry:	en e
pH (in water)	4 to 8
Eh	140 to 400 mV
Organic carbon	500 to 7000 ppm
Iron oxides	
Free	23,000 mg/kg
Amorphous	13.50 mg/kg
Cation exchange capacity	17.5 meq/100 g
Exchangeable ions:	
Mg	<10 to 45 mg/kg
Ca	<1000 to 5000 mg/kg
Soil microbiology:	
Total bacteria	100 to 1000 organisms/g

a estimated

Note: The information shown is based on a compilation from various sources including analyses of Minford soils from the X-231B site (ORNL 1994) and the ORNL reconnaissance boring and soil analyses completed during April and May 1994.

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3. Technical Approach

Work plans prepared for the project included a technical plan, a sampling and analysis plan, a quality assurance project plan, and a health and safety plan (ORNL 1994). All plans were reviewed, revised, and finalized prior to mobilization to the CTS. To ensure that representative data were collected for this project, ORNL standard operating procedures (SOPs) were used for conducting field activities (ORNL 1993). Table 3.1 lists the SOPs that were pertinent to this project. Table 3.2 summarizes the testing performed at each test cell. A readiness review for the pre-treatment site characterization was held at the PORTS facility on October 18, 1994. The readiness review for the injection testing was held on November 3, 1994, and the project was approved for start-up.

3.1 Task 1: Geoprobe Borings and Lysimeter Installation

The first task of this project was to characterize the soils to a depth of 12 ft in each of seven test cells (Fig. 3.1). Each test cell was 24×24 ft in surface area except for test cell 7, which was 4×8 ft. The size of test cell 7 was decreased to reduce the volume of reagent required, thereby reducing the cost for the iron micropowder. Test cell design and characteristics are summarized in Table 3.3.

For the pre-treatment characterization, lithologic samples were collected from five bore-holes in each test cell (labeled A through E) (Fig. 3.2), except for test cell 7, which was characterized with one pre-treatment borehole. The lithologic logs and fracture descriptions were documented on borehole lithologic forms and used to describe pre-treatment soil conditions (Appendix B). Other baseline determinations included water content, grain size, color, mineralogy, scanning electron microscope (SEM) morphology, pH, TOC, iron oxides, cations, and anions. Note that all analyses were not performed on all soil samples because the parameters were not expected to vary significantly over the entire test area and to control analytical costs.

Personnel from ORNL-GJ, used a rig with Geoprobe™ tools to collect the lithologic samples. On-site technical direction was provided by an ORNL-GJ geologist. Continuous 4-ft-long cores were collected with a megabore sampler. The on-site geologist was responsible for visual classification of the soils encountered and for completing an accurate, depth-based soil boring log. Microscopic examination of selected samples was also performed by the geologist; however, detailed fracture morphology was evaluated through laboratory analyses at ORNL.

Task 1 included completion of 32 Geoprobe borings, five borings in each of test cells 1 through 6, one boring in test cell 7, and one in a background area. The background location was approximately 50 ft south of the southeast corner of test cell 6 (Fig. 3.1). The boring locations were staked and surveyed shortly after completion. All pre-treatment borings were backfilled with granular bentonite or completed as piezometers or lysimeters.

Task 1 also included the installation of 24 porous ceramic cup lysimeters for sampling soil-pore water. Three lysimeters were placed in each of the seven test cells and three at a background location. The three lysimeters at each location were nested in a single borehole at depths of 4, 8, and 14 ft bgs (Fig. 3.3). Pre-treatment borehole C was consistently used for the installation of these lysimeters except in test cell 7, where borehole A was used. Lysimeters were also installed in the background soil boring. The lysimeters were 1 bar type with 15-psi bubbling pressure. The average pore diameter was 2.9 μ m, suitable for use in soils with 0 to 1 bar of soil suction.

Installed according to manufacturers specifications, each lysimeter body was surrounded by silica slurry pack mixed from 200 mesh, 99.88% pure crystalline silica flour and distilled water at a ratio of 150 mL of water to 450 g of silica flour. A bentonite seal was placed above each lysimeter to isolate it within the borehole. Before installation, lysimeters were pre-wetted with distilled water which was evacuated after installation. A vacuum of 22 in. of Hg was applied to each lysimeter 12 hours before anticipated sample collection. Samples were evacuated by applying positive pressure to the lysimeter and forcing the water in the lysimeter body to the surface through the sampling tube.

A total of five 2-in.-inside-diameter (ID) piezometers were also installed, one each in four of the test cells and one at the background location (Table 3.2 and Fig. 3.4). Borehole D was used for piezometer installations. Constructed with 1-ft-long screens placed at approximately 14 ft bgs, the piezometers were used for monitoring water levels and collecting groundwater samples. The lysimeters and piezometers were installed using solid-stem augers and standard well construction practices. All lysimeters and piezometers were sampled twice prior to the initiation of MPIS testing in order to establish baseline conditions.

Soil moisture and temperature (SMT) probes were installed approximately 2 ft east of each set of lysimeters. A total of 24 SMT probes were installed at depths corresponding to the lysimeter depths: 4, 8, and 14 ft bgs. The probes were seated in native soil, with wire leads routed to the ground surface inside of 1.5-in. polyethylene (PE) casing. The PE casing was backfilled with sand. An 18-in.-long piece of 4-in.-diameter polyvinyl chloride (PVC) casing with a slip-on cap was placed over the wire leads at the ground surface as a protective cover. Figure 3.5 provides a profile view of the layout for the instrumented borings.

3.2 Task 2: MPIS Testing

Task 2 consisted of a series of multi-point injections in six of the test cells with injection done on 2-ft centers (Fig. 3.2). Test cell number 6 was injected with air and had only eight injector penetrations. The MPIS developed and provided by Hayward Baker Environmental, Inc., consisted of a tractor-mounted unit with a set of four removable injection augers mounted 2 ft apart on a bar across the front of the tractor. These augers have injection ports in a spiral pattern near the tip. All four of the 2-in-diameter augers were simultaneously pushed and turned into the soil to a depth of over 10 ft, stopping at 15-in. intervals for fluid injection (Fig. 3.6). Approximately 2 gal of fluid was injected from each auger at each interval. An operator sitting on the tractor controlled the rate of penetration and injection (Fig. 3.7). The following describes the procedure used.

- The test cell was gridded on 2-ft centers for ease of controlling the injection spacings.
- The MPIS was positioned in the northeast corner of the test cell at setup number 1.
- The augers (4) were drilled into the soil to a depth of approximately 20 in., and 2 gal of reactive fluid were injected from each auger tip simultaneously.
- The augers were then drilled to the next position, approximately 35 in. bgs, and 2 gal were injected from each auger tip.
- This process continued at 15-in. intervals (eight injector positions) until a total depth of approximately 125 in. was attained and a total of 64 gal of fluid was injected at that setup (16 gal per injector).
- Four flow meters mounted in front of the MPIS operator were used to monitor the volume of injected fluid from each injector and the operator controlled flow to each injector separately, thus ensuring that the appropriate amount of reagent was injected from each injector at each position.
- Depth calibration marks on the mast indicated the depth of the injector points. A geologist
 monitored the injection process to ensure that the injections were performed uniformly
 across each test cell.

Few, if any, soil cuttings were generated during penetration. Upon reaching a depth of 10.4 ft (125 in.), the operator reversed the hydraulics and turned the augers up out of the soil. No fluid was injected during the upstroke. The total depth of the injections was based on the capabilities of the equipment used. Larger equipment with injection capability to 40 ft bgs is available.

To assess the performance of the multi-point injection equipment by means of operational checks and practice borings, process shakedown was performed in an area north of the CTS prior to initiation of the planned injection testing.

The reagents injected into each test cell, shown in Table 3.4, were delivered to the MPIS by a hose connecting the system to a pump that pulled liquid from a battery of mixing tanks stationed nearby. An air compressor provided pressurized air for the air injection test. The oxidants were injected because hydrogen peroxide and potassium permanganate can catalyze and chemically oxidize chlorinated organics to produce carbon dioxide and water. Zerovalence iron (iron micropowder) was injected because it has been demonstrated to act as a reducing agent for chlorinated solvents. Water alone was injected to provide a baseline test cell. The lime injection was tested because it could be used to adjust soil pH for in situ stabilization of metals. The bionutrient and surfactant were injected to determine the feasibility of using the mixture to enhance the natural breakdown of chlorinated solvents by indigenous bacteria. Air was injected to test the MPIS for pneumatic fracturing of LPM.

Tracers, including potassium bromide and Snowmax®, were mixed into two of the solutions to provide a means of differentiating the injected fluid from the existing soil-pore water and to permit detection of soil fractures (Table 3.4). Bromide is a conservative tracer (i.e., travels at the same velocity and direction of the water it is dissolved in) used to track the extent of injected fluid penetration into the soil. Snowmax® is a commercially available, ice-nucleating active bacteria product that is processed (the bacteria are killed and mixed with an inert ingredient) and sold as a nucleator for artificial snow. ORNL researchers have developed an innovative and unconventional use of Snowmax® and other bacteria as environmental tracers (Strong-Gunderson 1995). Snowmax® concentration is expressed in ice-nucleating activity (INA) particles per mg of soil.

The injections were performed on a pattern starting with four simultaneous injections 2 ft apart; the tractor-mounted MPIS was then moved 2 ft backwards and another injection performed until the entire test cell had been penetrated on 2-ft centers. Table 3.5 lists the process operating conditions monitored during the MPIS testing.

Because the field work for this project was performed at an uncontaminated site, equipment cleaning procedures were minimal and were performed as necessary to maintain proper operation of the equipment and to prevent mixing of different reagents. Injectors, hoses, tanks, pumps, and mixing equipment were cleaned on site using either a fresh water rinse or a portable steam cleaner as appropriate. Fluids generated by flushing the MPIS were discharged to the ground surface away from the CTS along with other water being pumped from the site.

3.3 Task 3: Concurrent Monitoring

Monitoring was performed during the MPIS injections made in each cell (Table 3.5 and Fig. 3.8). The volume of reagent injected was monitored on four separate flow gauges mounted on the MPIS unit and on a single flow meter at the pump discharge. The separate flow gauges on the MPIS unit recorded the amount of fluid injected by each of the four injectors and the flow gauge on the pump discharge recorded the total amount of fluid injected. Other observations included penetration rate, injection rate, injection pressure, surface blowout of fluid, and temperature and moisture changes in the test cell soils. Samples of the injected fluid were collected at the connection of the delivery hose to the MPIS unit periodically throughout each test cell injection to provide information on the uniformity of the batching and mixing operations.

3.4 Task 4: Post-Injection Characterization and Data Collection

The first phase of Task 4 for each test cell included drilling and sampling five additional soil borings in test cells 1 through 5 (boreholes F through J) and two additional soil borings (boreholes B and C) in test cell 7 (Fig. 3.1). These borings were drilled within 24 h after the injections in each cell to evaluate the effective distribution of treatment agents by the system and any rapid changes occurring in the subsurface (Fig. 3.9). Soil samples were examined both macroscopically and microscopically for fracture characteristics and the presence of tracers. Morphology was evaluated in the laboratory by SEM. If bromide was injected as a tracer, soil samples were extracted and analyzed for bromide on site with an ion-selective electrode. Samples for INA analysis were sent to ORNL for analysis. Selected soil samples were also analyzed for pH, TOC, Eh, Fe oxides, and Mn.

A soil inspection trench was excavated in the shakedown area to observe the effects of lime injection on the subsurface. Because the trench did not reveal characteristics that were not observable in the soil cores, no additional soil inspection trenches were excavated.

Soil-pore water samples were collected the day after each injection and daily thereafter for several days. Water samples were collected from the lysimeters and piezometers to determine the effects of the injections on the soil-pore water chemistry. Water samples were analyzed on site for temperature, pH, DO, conductivity, alkalinity, Br, Fe, Mn, Cl, NO₃, and SO₄. Analyses for metals and TOC were performed at ORNL.

The first phase of post-treatment characterization was completed within 3 days after the last injection was performed (November 21, 1994). The second phase of post-treatment characterization was performed about 2 weeks later and consisted of collecting another set of water samples from the lysimeters and piezometers on December 5, 1994. The third phase of

post-treatment characterization was performed about 1 week after the second phase and consisted of drilling two additional soil borings in test cells 1 through 5 (boreholes K and L) and in test cell 7 (boreholes D and E) and collecting another set of water samples from the lysimeters and piezometers on December 15, 1994. A fourth phase of water sampling only was performed during the week of February 15, 1995, and a final round of groundwater samples were collected the week of May 6, 1996.

3.5 Surveying

The locations of all test cells, borings, lysimeters, piezometers, SMTs, and elevation bench marks were surveyed. A nearby monitoring well was used as a benchmark. The required horizontal accuracy was ± 0.5 ft, and the required vertical accuracy was ± 0.01 ft. To ensure the required accuracies, the survey was looped and closed. Surveying was performed with a theodolite equipped with electronic distance measuring and a level. Lysimeters and piezometers were identified by numbers written on the side of the protective casing. The lysimeters were numbered T1L1 and T2L1, etc., in order to identify the test cell (T1) and the lysimeter (L1). Similarly, the piezometers were marked T1P1, etc., identifying the test cell (T1) and the piezometer (P1). The soil borings were numbered in accordance with a prearranged scheme in each test cell as follows: T1-A, to indicate test cell 1, soil boring A, T1-B to indicate test cell 1 and similarly in the other test cells. Elevations were checked prior to injection testing and then several times after testing to determine the amount of soil heaving caused by the injection and to determine the rate of settling.

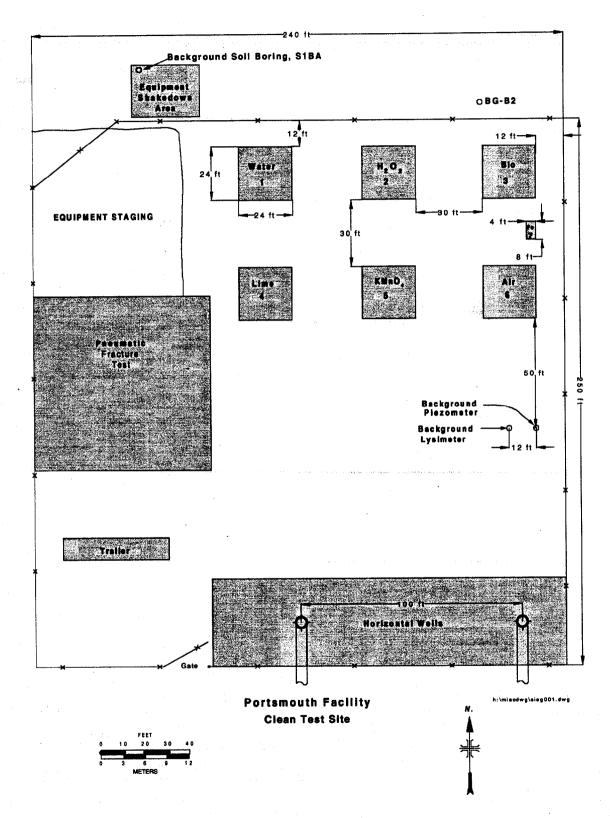
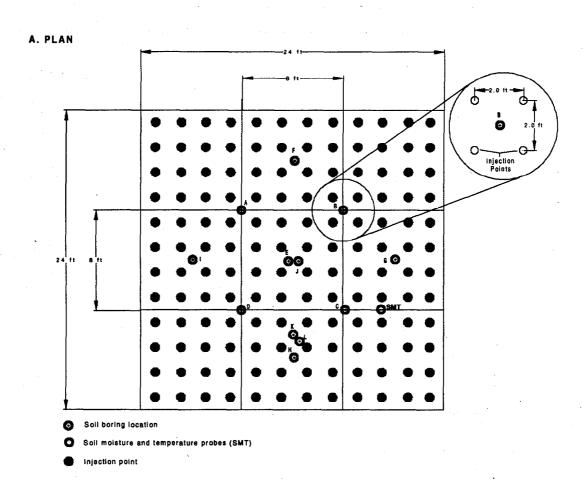


Fig. 3.1. Test cells within the CTS.



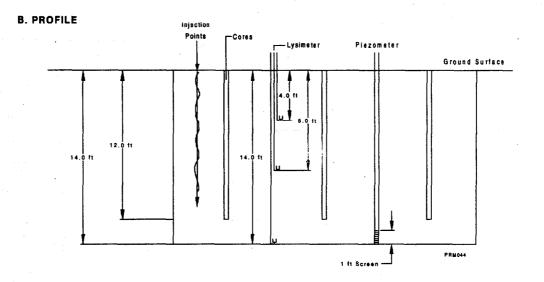
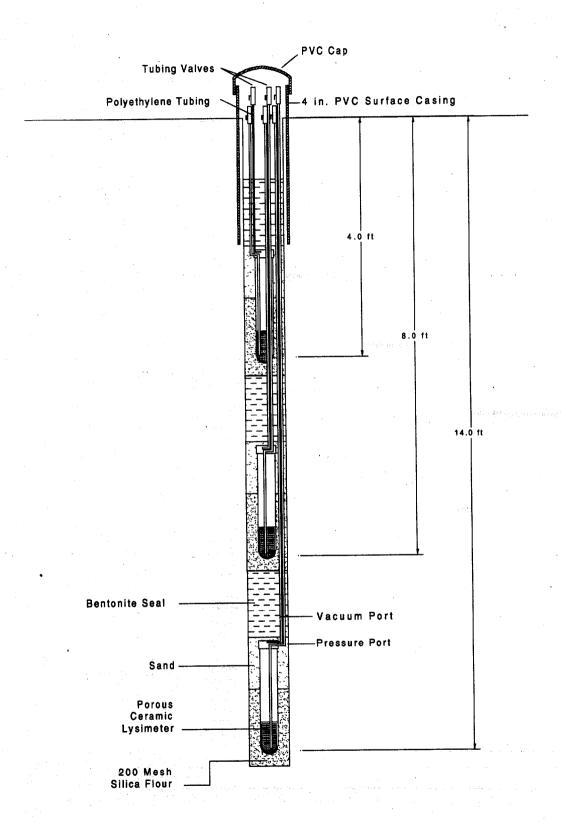


Fig. 3.2. Approximate locations for testing within a typical test cell.



NESTLYSM

Fig. 3.3. Schematic diagram of nested lysimeters.

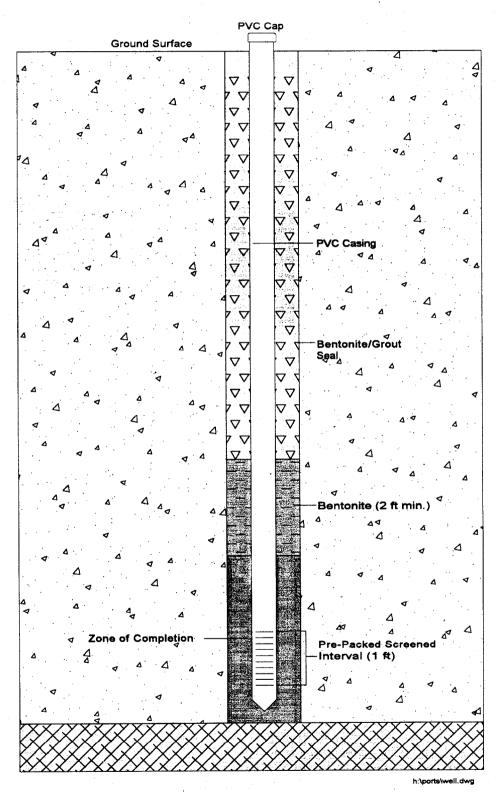
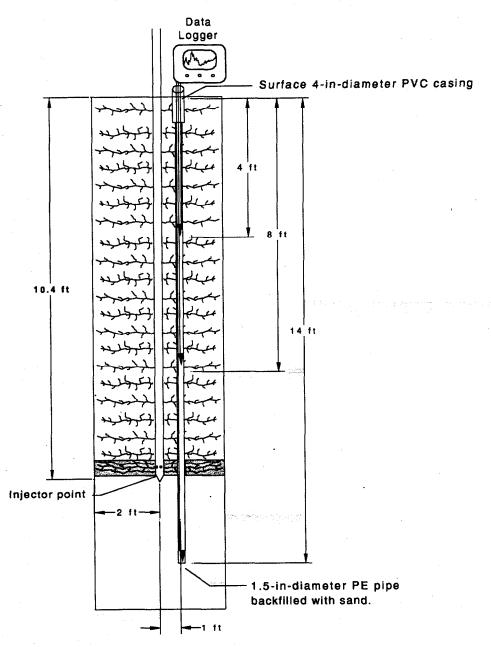


Fig. 3.4. Schematic diagram of piezometer.



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Reacting Solution

Active Injection

Soil Moisture and Temperature Probe

Fig. 3.5. Profile view of the layout for the instrumented borings.

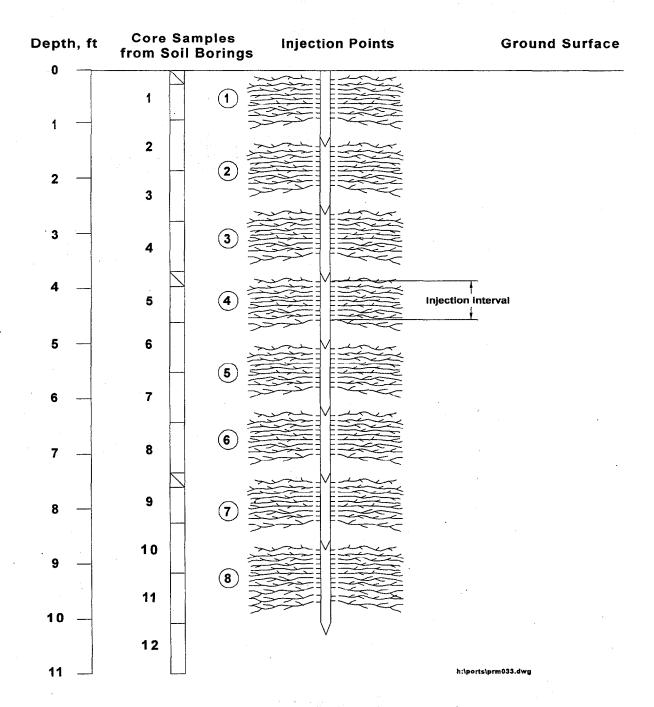
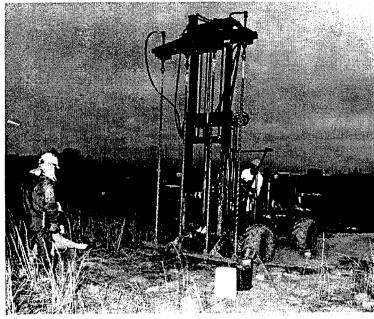


Fig. 3.6. Conceptual diagram of the MPIS process.

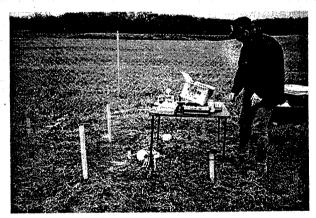
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Closeup view of auger and injection tip 18 holes in spiral pattern

Fig. 3.7. Multi-port injection system.



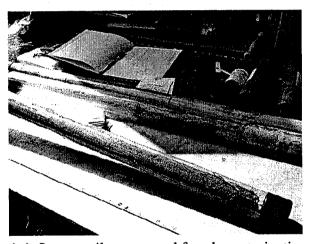
(a.) MPIS equipment performing an injection of bionutrient solution at test cell 3.



(b.) ORNL staff monitoring at test cell 7.

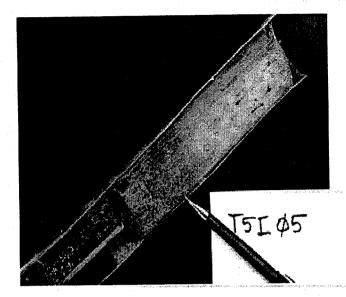


(d.) Onsite laboratory for soil core analyses.

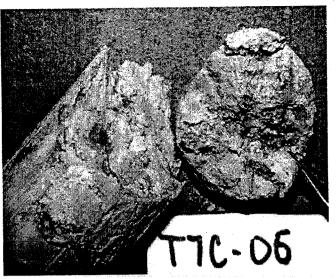


(c.) Intact soil cores used for characterization.

Fig. 3.8. Photographs of the Hayward Baker MPIS rig and the monitoring and sampling analyses activities.



(a.) Permanganate impacted core at 5-ft. depth (the soil redox is dramatically elevated through core beyond the obvious purple staining.



(b.) Iron micropowder deposition within silty clay soil at 6-ft depth..

Fig. 3.9. Photographs of intact soil cores from the permanganate (T5) and iron micropowder (T7) test cells.

Table 3.1. ORNL SOPs for the MPIS demonstration

ORNL SOP number	Title			
AD-050	Quality Assurance			
TE-061	Measurement of pH of Water Samples			
TE-062	Measurement of Electrical Conductivity of Water Samples			
TE-063	Measurement of Dissolved Oxygen in Water Samples			
TE-071	Sample Documentation			
TE-072	Sample Packaging, Preservation, and Shipping			
TE-073	Equipment Decontamination			
TE-086	Soil Sampling for Field Screening Using the Geoprobe and U2CRT			
TE-094	Water-Level Measurement			
TE-100	Drilling Log Preparation and Well Construction Documentation			
TE-105	U2CRT Operation			
TE-106	Solid-Stem Augering Using the U2CRT			
TE-120	Physical Surveying			
TE-130	Peristaltic Pump Operation			
TE-150	Combustible Gas Indicator Operation			

3 - 17

Table 3.2. Activities performed in conjunction with MPIS testing

Test cell, injection treatment	Pre-treatment activities	Injection testing	Post-injection 1 activities ^e	Post-injection 2 activities ^a
No. 1, Water	5 soil borings 3 lysimeters 1 piezometer 3 soil moisture probes 5 elevation markers	Inject water with tracers Monitor parameters	5 soil borings Sample lysimeters and piezometer Monitor parameters Surveyed elevation markers	2 soil borings Sample lysimeters and piezometer Monitor parameters Surveyed elevation markers
No. 2, H ₂ 0 ₂	5 soil borings 3 lysimeters 1 piezometer 3 soil moisture probes 5 elevation markers	Inject reagent Monitor parameters	5 soil borings Sample lysimeters and piezometer Monitor parameters Surveyed elevation markers	2 soil borings Sample lysimeters and piezometer Monitor parameters Surveyed elevation markers
No. 3, Bionutrient	5 soil borings 3 lysimeters 1 piezometer 3 soil moisture probes 5 elevation markers	Inject reagent with tracers Monitor parameters	5 soil borings Sample lysimeters and piezometer Monitor parameters Surveyed elevation markers	2 soil borings Sample lysimeters and piezometer Monitor parameters Surveyed elevation markers
No. 4, Lime	5 soil borings 3 lysimeters 3 soil moisture probes 5 elevation markers	Inject reagent Monitor parameters	5 soil borings Sample lysimeters Monitor parameters Surveyed elevation markers	2 soil borings Sample lysimeters Monitor parameters Surveyed elevation markers
No. 5, KMn0 ₄	5 soil borings 3 lysimeters 1 piezometer 3 soil moisture probes 5 elevation markers	Inject reagent with tracers Monitor parameters	5 soil borings Sample lysimeters and piezometer Monitor parameters Surveyed elevation markers	2 soil borings Sample lysimeters and piezometer Monitor parameters Surveyed elevation markers

3-18

Table 3.2. (continued)

Test cell, injection treatment	Pre-treatment activities	Injection testing	Post-injection 1 activities	Post-injection 2 activities*
No. 6, Air	5 soil borings 3 lysimeters 3 soil moisture probes 5 elevation markers	Inject air Monitor parameters	Sample lysimeters Monitor parameters Surveyed elevation markers	Sample lysimeters Monitor parameters Surveyed elevation markers
No. 7, Fe	1 soil boring 3 lysimeters 3 soil moisture probes 1 elevation marker	Inject reagent Monitor parameters	2 soil borings Sample lysimeters Monitor parameters Surveyed elevation marker	2 soil borings Sample lysimeters Monitor parameters Surveyed elevation marker
Background location	1 soil boring 1 piezometer 3 lysimeters 3 soil moisture probes	Monitor parameters	Sample lysimeters and piezometer Monitor parameters	Sample lysimeters and piezometers Monitor parameters
Total	32 soil borings 24 lysimeter samples 5 piezometer samples 24 soil moisture probes	7 injections Multiple parameter data set	27 soil borings Multiple lysimeter samples Multiple groundwater samples Multiple parameter data set	12 soil borings 24 lysimeter samples 5 groundwater samples Multiple parameter data set

^a The parameters monitored are shown in Table 1.1.

Table 3.3. Test cell design and characteristics

Test cell geometry:	24 × 24 ft
Ground surface area	576 ft ²
Soil depth	10.4 ft
Soil volume	5,990 ft ³
	44,800 gal
	169,640 L
Soil weight (wet)	305,310 kg
Soil bulk density (field moist)	1.8 g/cm ³
Soil water content:	
Field moist basis	20 weight %
Total water weight	70,450 kg
Soil-pore system:	
Total fractional porosity ^a	0.40 v/v
Total pore volume	78,280 L
Water-filled pore volume/cell	70,450 L
Water-filled porosity	36% cell
Pore water saturation	90% pores
Air-filled pore volume/cell	7,830 L
Air-filled porosity	4% cell
Pore air saturation	10% pores

[«] estimated.

Note: See Table 2.1 also.

Table 3.4. Treatment agents for MPIS testing at PORTS CTS

Test cell	Treatment type	Medium	Treatment/tracer	Concentration
1	Control	Water	Water/bromide	NA
2	Oxidant	Water	H ₂ O ₂ /none	10%
3	Bionutrient	Water	Surfactant, nutrients/	10%
			bromide and Snowmax®	•.
4	Stabilizer	Water	Lime/none	20%
5	Oxidant	Water	KMnO ₄ /none	5%
6	Fracturing	Air	None	NA
7	Reductant	Water	10- μ Fe filings, guar gum/none	20%

NA = not applicable

Table 3.5. Process operating conditions for MPIS testing at the CTS

	The second of th
Injector operation:	down and up ange
Motion	down and up once 2.75 to 7.9 min
Injection time	~12.6 ft ²
Area of affected region	~12.6 ft 10.4 ft
Depth of affected region	130 ft ³
Volume of affected region	
Injections per cell, cells 1 through 5	144
Unit area of surface per injector	~4 ft²/point
Unit weight of soil per injector	~2445 kg/point
Water injection test:	0.00
Volumetric water addition	0.15 to 0.25 gal/ft ³
Total volume per injector	16 gal
Total volume per cell	2400 gal
Rate of injection addition	8 to 23 gal/min
Increase in soil water content	~2.0 weight %
Solute tracers	
Br concentration	205 mg/L
Br mass loading (avg. cell)	9 mg/kg soil
H ₂ O ₂ injection test:	
H ₂ O ₂ solution strength	10% by weight
H ₂ O ₂ solution addition/injection	16 gal
H ₂ O ₂ mass loading (avg. cell)	~2.5 g/kg soil
Nutrient/surfactant test:	
Volume water addition/injection	16 gal
Surfactant concentration	10% volume
Lime Injection:	n de en
Total volume per injector	16 gal
Injection rate	8 to 23 gal/min
Lime slurry concentration	20 weight %
Lime mass loading (avg. cell)	~6.9 g/kg soil
Permanganate test:	er der er vikt i det for det geliefen i de det til der til en skriver i det skriver i det skriver i de det ett Det skriver
KMnO ₄ solution strength	5% weight %
KMnO ₄ solution addition/injection	16 gal
KMnO ₄ mass loading (avg. cell)	~1.3 g/kg soil
Air test:	other than to the decision of the consequence of the enterior fraction. More than 100 to 100 to 100 to 100 to 1 The consequence of the consequence
Air injection rate	60 scfm
Air pressure	60 to 200 psig
Air temperature	ambient °C
	an a survey and a survey and a survey and and an analysis of the survey
Iron test: Volumetric water addition/injection	16 gal
	$5 \mu \text{m}$
Fe(0) particle size	20 weight %
Fe(0) solution concentration	25.5 kg
Fe(0) mass/injection	10.4 mg/kg soil
Fe(0) mass loading (avg. cell)	10.4 mg/kg 500

4. Specific Testing and Results

Field activities were documented by daily entries into the project logbook and task-specific field logbooks: a geologist's logbook, health and safety logbook, water sample logbook, soil sample logbook, survey logbook, and laboratory logbook. The project logbook provides a summary record of all field activities. Information was also recorded on forms designed specifically for this project. Prior to injection, pre-treatment elevations were surveyed using control stakes placed in each test cell. These elevations are included in the ground surface uplift graphs for each test cell that show the relative change in elevation after injection. The following is a description of the results obtained at each test cell at CTS.

4.1 Background CTS Characterization

4.1.1 Field Activities

Characterization of the geology of the CTS is described in Sect. 2.3. The site was further characterized by collecting background data specifically for this project, including drilling three additional soil borings, two in the background area (Fig. 3.1) and one in the shakedown area. Lithologic logs of these borings were prepared, and soil samples were shipped to ORNL for analysis. A piezometer was installed at boring B1-A, and three lysimeters were nested in boring B1-B. Three SMT probes were installed approximately 2 ft east of the lysimeters. Water samples collected from the piezometer and lysimeters on November 1, 1994, were shipped to ORNL for analysis. Other water samples were collected periodically from November 12, 1994 to May 9, 1996. These samples were analyzed on-site in the field laboratory, with some ancillary testing performed at ORNL.

4.1.2 Pre-Treatment Soil Testing

Measurements of pre-treatment soil conditions at the CTS were made both within individual test cells (borehole E in test cells 1 through 6 and borehole A in test cell 7) and at one background location (near the equipment shakedown area). Values from these eight locations were averaged to provide a basis for experimental design and for comparison with post-treatment soil conditions. Parameters measured included soil moisture content, bulk density, temperature, pH, Eh, TOC, grain size, bromide, manganese, and calcium. A summary of the pre-treatment soil conditions is presented in Table 4.1.

The average depth-specific soil moisture content ranged from 15 to 26% with moisture content generally increasing with depth. The moist bulk density ranged from 1.57 to 2.25 g/cc with the an overall bulk density average of approximately 1.84 g/cc. Pre-treatment soil temperatures averaged 56° F at 4 ft bgs, 58° F at 8 ft bgs, and 56° F at 14 ft bgs. Soil pH increased with depth and ranged from 4.3 to 7.4. Soil Eh, measured only at the background location near the shakedown area, was 400 mV near the ground surface and decreased to 140 mV at 12 ft bgs. Average TOC ranged from 6397 ppm near the ground surface and decreased to a minimum value of 395 ppm at 9 ft bgs.

During horizontal well installation at the CTS, selected samples were submitted for grain size distribution analysis. Of these samples, four samples were collected near the multi-point injection permeation and dispersal test area at depths of 4.5 to 5 ft bgs and 13.5 to 14 ft bgs (Appendix A). In these samples 85 to 98% of the sample passed through a #200 mesh sieve confirming the soils as silty clays. Visual inspection of the soil cores identified the soils as silty clay with colors ranging from light yellowish brown to reddish brown with gray and green mottling.

Manganese and calcium were analyzed in soil samples from pre-treatment borings in test cells 1, 2, and 4. Most values for manganese were less than 10 ppm, except for the top one foot of soil where manganese concentrations were 20 to 45 ppm. Calcium concentrations were generally less than 1000 ppm to depths of 6 or 7 ft bgs and then increased dramatically to generally more than 5000 ppm. These data indicated that calcium has been leached from the upper soil profile and probably redeposited at depths greater than 6 ft. This is consistent with the acidic pH of the upper soil, where after breakdown of CaCO₃, calcium would be leached from the soil. The soil would then be enriched with calcium where soil pH was more neutral, thus supporting the formation of gypsum crystals observed in the 6.5 to 8 ft bgs zone.

Discussion of the bromide data and additional detail related to cation analyses are presented with the post-treatment results within the following specific test cell sections.

4.1.3 Background Water Testing

Parameters routinely measured in water samples from the lysimeters and piezometers included water level, pH, conductivity, temperature, DO, TOC, NO, Br, alkalinity, Fe, Mn, Cl, and SO₄. Results from the background piezometer (B1P1) and lysimeters (B1L1, B1L2, and B1L3) are shown in Table 4.2. Water pH varied from 6.1 to 8.9 during the period of November 1, 1994 to February 15, 1995. The pH values were consistently lowest in the piezometer water samples, possibly due to contact of the water with air in the casing. DO values varied from 2 mg/L to 11.2 mg/L, with no consistent trends in the data with depth or

time. Conductivity measurements remained about the same for all sampling points over the duration of testing; however, it is interesting to note that the lowest conductivity values were measured in the shallowest sample B1L1 and the highest values were measured in the 8 ft sample, B1L2. Alkalinity varied from 94 to 319 mg/L. Alkalinity values measured in the shallow lysimeter were somewhat erratic, while values measured in the 8 ft and 14-ft lysimeter and the piezometer were more consistent. Bromide concentrations were less than 1 mg/L in all samples tested. Similarly, nitrate concentrations were less than 0.5 mg/L in all background samples tested. Manganese values were 1 mg/L or less and averaged approximately 0.25 mg/L. Chloride ranged from 2.5 to 23 mg/L. Sulfate concentrations ranged from 425 to 1200 mg/L.

4.2 Test Cell 1: Water Injection with Tracers

The first injection was in Test Cell 1 where water with tracers were injected. This was done to familiarize field personnel with the injection procedure and demonstrate that liquid injection was possible at this site. Thus, the test served as a practice session, while the tracers provided information on the behavior of water injected in the subsurface.

4.2.1 T1 Field Activities

Five pre-treatment soil borings, T1-A through T1-E, were drilled and sampled on October 23, 1994. Lithologic logs were prepared and soil samples shipped to ORNL for analysis. One piezometer was installed at boring T1-D, and three lysimeters were nested in boring T1-C (Fig. 4.1). Three SMT probes were installed approximately 2 ft east of the lysimeters. Water samples were collected from the piezometer and the lysimeters on November 1, 1994, and were shipped to ORNL for analysis. Preparation for injection testing at test cell 1 began on November 12, 1994. A set of water samples was collected from the lysimeters and piezometer, and elevation benchmarks were surveyed prior to the start of injection. Water with bromide and Snomax® tracers was injected on 2-ft centers to a depth of 10.4 ft starting at 2:00 p.m. The test cell injection was completed at 5:45 p.m.; thus, 225 min were required to perform 144 injections (36 MPIS set-up locations with four injector locations per setup) (Fig. 3.1). The average time for each injection setup was 6.2 min excluding 35 min to refill and mix three 500-gal tanks.

The objective was to inject 2 gal per injector at each 15-in. depth interval. With four injectors per set-up location and eight positions for each injector, the target total for injection at each setup was 64 gal (4 injectors \times 8 positions \times 2 gal each), and the target for 36 set-up locations was 2,304 gal (36 setups \times 64 gal each) for the entire test cell. According to the flow meters,

approximately 2400 gal of solution were delivered; the estimated amount of solution lost to seepage at the ground surface was 400 gal. Thus, the amount of solution injected into the subsurface was estimated to be 2000 gal.

Six batches of powdered KBr and Snomax® solution were mixed and added to each of six 500-gal batches of water. Two samples of each of the six batches were collected during injection to evaluate the uniformity of the injected solutions. The samples were analyzed for bromide in the on-site laboratory. Bromide concentrations varied from 190 to 235 mg/L with a mean of 205 mg/L.

Test Cell No.	Total solution injected	Target solution strength	Average actual solution strength	Duration of injection testing	Average time per setup	Total surface seepage
T1	2400 gal	200 mg/L Br	205 mg/L Br	225 min	6.25 min	400 gal

Soil resistance and temperature readings were taken before, during, and after the injection. Resistance measurements showed that the soil was near saturation at 14 ft bgs prior to injection, but was not saturated at 8 and 4 ft bgs. However, approximately 8 min after injection started, the backfilled soil around the subsurface probes was saturated with injected solution, and solution began to run out the top of the protective casing onto the ground surface. Soil temperature showed an increase of 3°F at 4 ft bgs and no change at 8 and 14 ft bgs.

The first phase of post-treatment characterization began with the completion of five additional soil borings (T1-F through T1-J) to a depth of 12 ft each. The borings were continuously sampled with 48-in.-long megabore samplers, and the resulting 1.5-in.-diameter soil cores were extensively subsampled. Each core was divided into 1-ft segments, and the following samples were taken from each segment: 60 g of soil for water content, TOC, and pH; 10 g of soil for archive; 1 kg of soil for iron oxides, x-ray analysis, SEM, and conductivity; 1 kg of soil to the field lab for pH, Br⁻, Cl⁻, Fe, Mn, SO₄², H₂O₃ and KMnO₄; and ~200 g for INA and Br⁻. A few samples were wax-coated for x-ray analysis. Not all analyses were performed on all samples. Lithologic descriptions were also prepared (Appendix B).

Post-treatment water samples were collected from the lysimeters and piezometer eight times during the 9 days following the injection and again approximately 1 and 3 months after injection. Similarly, elevation benchmarks (S1, S2, S3, S4 damaged, and S5) were surveyed six times during the nine days following the injection and again about one month after injection. During the fourth week after injection, December 12 to 14, 1994, two additional soil borings (T1-K and T1-L) were cored and sampled. The subsampling performed was not as extensive as that performed during the first phase of post-treatment soil sampling and was

limited to performing Eh measurements as the cores were divided into 1-ft segments, preparing a lithologic log, and shipping the cores to ORNL for additional analysis.

4.2.2 T1 Soil Sample Results

Pre-treatment soil sample results include measurements of soil pH, TOC, % moisture, temperature, and bulk density. Soil sample results are summarized in Table 4.3 and data trend graphs are presented in Appendix C. Values for pre-treatment soil pH were measured on the core from borehole E and ranged from a low of 4.3 near the ground surface to a high of 7.8 at 10 ft bgs. There was a general increase of pH with increasing depth. Because the injection of water with tracers was not expected to have much effect on soil pH, no post-injection pH measurements were performed.

TOC was evaluated on pre-treatment core samples from borehole E. The values ranged from a high of over 6000 ppm near the surface to about 350 ppm at depth. The values were highest near the ground surface due to humic material in the top soil. TOC values decreased steadily to a depth of 7 ft bgs and then were variable but less than 1000 ppm down to 12 ft bgs. Because no change in TOC was expected due to the injection of water with tracers, post-treatment TOC was not measured.

Moisture content was measured in samples from one pre-treatment boring, borehole E. Samples ranged from 15% to 27% moisture (dry wt.%). These values are consistent with other background soil moisture measurements for the CTS. Moisture content was measured at 1 ft intervals on five post-treatment borings (Fig. 4.2). Comparison of depth-specific moisture values showed that moisture content of the near-surface interval (0 to 4 ft bgs) had increased from ~15% to as much as 24%. The intermediate-depth interval (4 to 8 ft bgs) was unchanged, and the deepest interval (8 to 12 ft bgs) showed a slight increase in soil moisture. Thus, the injection of 2000 gal of water into the test cell had little effect on soil moisture content. This is consistent with predictions made which suggested an average increase of a 2.0 wt.% based on the volume of water added into the volume of media in the cell (Table 3.5).

Soil surface elevation data show that the surface of the test cell was raised about 0.1 ft by the injection and remained near that elevation through the last measurement period (three months after injection). The volume change in a 24 ft × 24-ft area that swells 0.1 ft is about 57 ft³ or about 430 gal. Thus, 1570 gal of water are unaccounted for by swell measurements alone.

Core samples from the pre-treatment borings were carefully logged, and visible pores and fractures in the soil were noted (Appendix A). There were numerous root pores and vesicles seen from the surface down to about 5 ft deep. Open-bed partings were noted at 7.5 to 8.5 ft bgs, with infillings of authigenic gypsum crystals. Clay content and soil stiffness increased

with depth. A few high-angle fractures were noted at 5 to 6 ft deep, but most of the fractures were between 9.5 and 11.5 ft deep. Some of these fractures had gray-clay infillings. Post-treatment core samples were also carefully logged. There were no differences seen between the number of fractures and the appearance of fractures before and after injection.

Bromide concentrations were determined in eight pre-treatment soil samples. Concentrations ranged from 0.7 to 1.5 ppm and averaged 0.88 ppm. Bromide concentrations were determined in post-treatment soil samples on 1-ft intervals for five borings drilled 12 ft bgs (Fig. 4.3). For the depair interval of 0 to 2 ft bgs, all measurements were above the average background bromide concentration of 0.88 ppm, mostly between 3 and 8 ppm. The interval of 2 to 5 ft bgs showed bromide concentrations ranging from background to about 6 ppm, with an average of about 3 ppm. The interval of 5 to 7 ft bgs showed concentrations ranging from background to 3.5 ppm, averaging about 1.5 ppm. The interval of 7 to 9 ft bgs showed concentrations ranging from background up to 6 ppm and averaging about 3.5 ppm. The deepest interval, 9 to 12 ft bgs, had bromide concentrations ranging from background to 4 ppm, averaging 1.0 ppm. These data indicate that different zones in the soil were preferentially enriched with bromide tracer as a result of the injection. The estimated increase in Br concentration to be caused by the concentration and volume of solution injected into the cell was approximately 9 ppm (Table 3.5). As observed, the entire cell down to 10 ft bgs appears to have been somewhat enriched in bromide. The greatest increase in bromide was in the near surface, 0 to 2 ft, where the soils were the least compact and had the highest density of root vesicles and pores. The zone between 7 and 9 ft, where the open-bed partings were noted in pre-treatment borings, was the second most enriched zone. The least-enriched zone was below 10 ft, which correlates to the total depth of injection of about 10.4 ft.

Snomax® tracer was evaluated by performing INA analysis on samples from five post-treatment borings (Fig. 4.4). The INA values were generally much higher in the top 4 to 5 ft of soil and ranged from 18 to 3500 particles/mg of soil. Values dropped dramatically below 5 ft and were generally less than 10 particles/mg of soil. Some borings showed an increase in INA between 7 and 9 ft bgs, again probably related to the visible, open-bed partings. Values in other borings dropped to near 0 from 7 to 12 ft bgs.

Eh was not measured on the pre-treatment soil cores, but post-treatment Eh values ranged from 275 to 500 mV in two boreholes, T1-L and -K. These values are similar to those measured in the pre-treatment background boring B1A.

4.2.3 T1 Water Sample Results

Post-treatment water sample data are presented in Table 4.4 and data trend graphs are presented in Appendix C. Water sample results indicate that the pH of the water in the

lysimeters and piezometer varied from 6.5 to 8.7 with no apparent trends with depth of the water sample or time elapsed after the injection.

DO values in water samples from the three lysimeters were highest prior to the injection and decreased markedly in T1L2, the 8-ft-deep lysimeter, after the injection. DO values appeared to stabilize after about 3 days (November 17, 1994) and remained relatively constant until November 21, 1994. Then, when samples were collected on December 15 and February 16, DO values were again near pre-treatment values. Piezometer DO values were more consistent throughout the analysis period.

Electrical conductivity remained about the same before and after injection except in T1L2, the 8-ft-deep lysimeter, where conductivity doubled after injection (November 14, 1994) and remained at a higher value. Data for the December 5, 1994, sampling event show low conductivity for all samples; it appears that there may have been a problem with the conductivity meter at that time.

Background bromide concentrations were around 1 mg/L. Bromide was detected at 80 to 100 mg/L in T1L1 and T1L2 after the injection and then slowly decreased over the next several days (Fig. 4.5). These concentrations are about one-half the strength of the injected reagent, 205 mg/L, indicating dilution by the resident soil pore water. Bromide concentrations were minimal in the deep lysimeter and piezometer, both at 14 ft bgs, indicating that the bromide tracer was not migrating rapidly downward after injection.

However, samples taken in December and February from the deep lysimeter and the piezometer showed increasing Br concentrations, indicating that some downward infiltration of injected solution was occurring.

Other analytes tested included alkalinity, Fe, Mn, Cl⁻, NO₃⁻, and SO₄² but since there were no noticeable changes in concentration in the first few samples, analysis for these parameters was discontinued, and no evaluation of the limited data sets was deemed necessary.

4.3 Test Cell 2: Hydrogen Peroxide

4.3.1 T2 Field Activities

Five pre-treatment soil borings, T2-A through T2-E, were drilled and sampled on October 22, 1994. Lithologic logs were prepared, and soil samples were shipped to ORNL for analysis. One piezometer was installed at boring T2-D, and three lysimeters were nested in boring T2-C (Fig. 4.6). Three SMT probes were installed approximately 2 ft east of the lysimeters. Water samples collected from the piezometer and lysimeters on November 1,

1994, were shipped to ORNL for analysis. Preparation for injection testing at test cell 2 began the morning of November 15, 1994. A set of water samples was collected from the lysimeters and piezometer, and elevation benchmarks were surveyed prior to the start of injection. A 10% by wt. hydrogen peroxide solution was injected on 2-ft centers to a depth of 10.4 ft starting at 9:45 a.m. The first two positions were injected at a rate of 2.5 gal per injector per position; however, there was a large amount of surface seepage, so the injection rate was reduced to 2.0 gal per injection. The test cell injection was completed at 1:10 p.m.; thus, 205 min were required, and the average time for each injection setup was 5.7 min.

The objective was to inject 2 gal per injector at each 15-in depth interval or a total of 2,304 gal (36 setups × 64 gal each) for the entire test cell. According to the flow meter at the pump, 2,352 gal were injected, and according to flow meters on the MPIS unit, 2,418 gal were injected. The estimated amount of solution lost to seepage at the ground surface was 500 gal. Thus, the amount of solution injected into the subsurface was estimated to be 1800 gal.

The solution was delivered to the site in one batch in a 3000-gal tank with no tracer added. Six samples were collected during injection to evaluate the uniformity of the injected solution. The samples were screened for H_2O_2 in the on-site laboratory and analyzed for H_2O_2 at ORNL.

Test Cell No.	Total solution injected	Target solution strength	Average actual solution strength	Duration of injection testing	Average time per setup	Total surface seepage
T2	2300 gal	10% H ₂ O ₂	10% H ₂ O ₂	205 min	5.7 min	500 gal

Soil resistance and temperature were measured before, during, and after the injection testing. Resistance measurements showed that the soil was near saturation at 8 to 14 ft bgs prior to injection testing, but was not saturated at 4 ft bgs. However, soon after injection started, the backfilled soil around the subsurface probes was saturated with injected solution, and solution began to run out the top of the protective casing onto the ground surface. Soil temperature was monitored throughout the test and showed an increase of 8°F at 4 ft bgs, 2°F at 8 ft bgs, and no change at 14 ft bgs. Temperature was also monitored in one of the reacting injector holes and varied from a pre-treatment value of 56°F up to 80°F less than 1 hour after injection. Temperature decreased to 70°F about 3 hours after injection was completed even though gas bubbles indicated that subsurface reactions were still in progress.

Hydrogen peroxide appeared to react with the soil immediately upon injection, and gas and mud bubbled from all injector locations for several hours after injection was completed. At

the peak of reactivity, there was brown, foaming mud up to 6 in. deep over the entire test cell. The reaction of peroxide in the soil produced gases, which rose to the surface, pushing fluid and mud out of the injection holes generating the foam as part of the reaction. In addition, there was a blowout during the injection from an injector hole in test cell 1 about 40 ft away from the position being injected in test cell 2. There were several areas within and near test cell 2 where the ground surface was raised to about 8 in. above the pre-treatment position, apparently due to gas buildup beneath the sod. Most of these raised areas subsided rapidly after blowouts relieved the pressure.

The first phase of post-treatment characterization began with the completion of five additional soil borings (T2-F through T2-J) to a depth of 12 ft each. The borings were continuously sampled with 48-in.-long megabore samplers, and the resulting 1.5-in.-diameter soil cores were extensively subsampled. Each core was divided into 1-ft segments, which were then subsampled. Eh was measured on the core as it was subsampled; lithologic descriptions were also prepared.

Post-treatment water samples were collected from the lysimeters and piezometer seven times during the 7 days following the injection. Similarly, elevation benchmarks (S1 - S5) were surveyed five times during the 7 days following the injection.

Another set of water samples was collected from the lysimeters and piezometer on December 5, 1994, approximately 3 weeks after the injection. During the fourth week after injection, December 12 to 14, 1994, two additional soil borings (T2-K and T2-L) were cored and sampled. The on-site subsampling performed was not as extensive as that performed during the first phase of post-treatment soil sampling and was limited to measuring Eh as the cores were divided into 1-ft segments, preparing a lithologic log, and shipping the cores to ORNL for additional analysis. An additional set of water samples was collected from the lysimeters and piezometer in December 1994, February 1995 and May 1996.

4.3.2 T2 Soil Sample Results

Pre-treatment soil samples collected from test cell 2 were analyzed for pH, Eh, TOC, % moisture, manganese, and calcium. Samples were collected from borehole E. Analysis results were consistent with the background values discussed in Sect. 4.1. Post-treatment soil samples collected from the first five soil borings, T2-F through T2-J, were analyzed for pH, peroxide, nitrate, Eh, TOC, and % moisture. The two additional borings, T2-K and T2-L, were analyzed for Eh only. Soil sample results for test cell 2 are summarized in Table 4.5 and data trend graphs are presented in Appendix C.

Results from pH measurements were similar for all boreholes, with values ranging from 4.2 to 7.5 and pH increasing with depth. These values are similar to the pre-treatment sample pH values; thus, the peroxide injection did not effect a significant pH change.

Peroxide was detected in four of the five post-treatment borings in the 2- to 3-ft interval (Fig. 4.7). Values ranged from approximately 4 to 100 mg/L. No other peroxide was detected in the samples, excluding the 6 to 7 ft interval in borehole F.

Nitrate analysis was performed on soil extracts for which soil samples were extracted with water and the resulting water samples analyzed. Values ranged from 0 to 3.75 ppm; however, the results were sporadic. Values from borehole H were higher than those from other boreholes, however, these values are not considered to be significantly higher due to a variation in the analysis procedure. No significant trends are seen in the nitrate data.

Soil Eh measurements for all boreholes showed a general decreasing trend with depth. An average maximum of about 450 mV was measured 2 to 3 ft bgs and an average minimum of approximately 250 mV was observed at 7 to 12 ft bgs (Fig. 4.8). The higher Eh values in the 2 to 3-ft interval correspond with the depth of peroxide detection in the cell.

Post-treatment TOC measurements ranged from approximately 400 to 2900 ppm. Compared to pre-treatment measurements, there is a dramatic decrease in TOC at the 0 to 2-ft interval. Indeed, TOC content at the 1-ft interval dropped from a pre-treatment level of 6100 ppm to a post-treatment concentration of 2900 ppm. This effect correlates with the distribution of peroxide observed above and demonstrates the oxidizing potential of peroxide. Little change was observed at depths greater than 2 ft.

Post-treatment soil moisture readings ranged from approximately 12 to 28%. Comparison to pre-treatment, depth-specific moisture values shows that moisture content in the near-surface interval (0 to 4 ft) increased by approximately 3%. The intermediate depth interval (4 to 8 ft) was unchanged, and the deepest interval (8 to 12 ft) showed a slight increase in soil moisture. These results are similar to those seen for test cell 1 (see Sect. 4.2.2).

Soil surface elevation data show that the surface of the test cell was raised approximately 0.3 ft by the injection and remained near that elevation through the last measurement period (3 months after injection). Core samples from both the pre-treatment and post-treatment borings were carefully logged, and visible pores and fractures were noted. No differences were seen between the number and appearance of fractures before and after injection.

4.3.3 T2 Water Sample Results

Post-treatment results are presented in Table 4.6 and data trend graphs are presented in Appendix C. Results indicate that the pH of the water in the lysimeters and piezometer varied from 6.6 to 9.0. The only trend noted was in the shallowest lysimeter, T2L1, at 4 ft bgs, where pH decreased steadily from the high of 9.0 to about 7.4 within 6 days after injection of peroxide. DO values were between 4 and 8 mg/L the day after injection and increased dramatically in T2L1 the second day after injection (Fig. 4.9). A significant increase (to 14 mg/L) was also observed in T2L2. DO in T2L3 remained about the same for 5 days after injection (4 to 9 mg/L) and then increased to a maximum of 11.2 mg/L approximately 2 weeks later. DO values in the piezometer were steady throughout the period measured probably due to contact of the water with air inside the casing. Conductivity varied considerably with the depth of the water sample, with the shallowest lysimeter having the lowest conductivity at about 800 to 1900 μ mhos and the deepest lysimeter and the piezometer having values around 4500 μ mhos. These values are not significantly different from the pre-treatment values, excluding T2L2, which had an increase of approximately 1700 μ mhos after injection.

Peroxide was detected in water samples from T2L1 and T2L2 the day after injection, but undetected 5 days later (Fig. 4.10). Nitrate values varied between undetected and 2 ppm for all water samples for 7 days after injection except in T2L2, where nitrate increased dramatically to 12 ppm the day after injection and then decreased to background by the fifth day after injection. It is not known if the increase in T2L2 is actual or due to a variation in the analysis procedure.

Alkalinity ranged from approximately 14 to 860 mg/L, with a general trend of increase with depth. These values are not significantly different from background values, excluding T2L2, which showed a dramatic increase the fifth day after injection. This increase corresponds to the conductivity increase observed for T2L2.

Other analytes tested included Fe, Mn, Cl⁻, and SO₄², but since there were no noticeable changes in concentration in the first few samples, analysis for these parameters was discontinued, and no evaluation of the limited data sets was deemed necessary.

4.4 Test Cell 3: Bionutrient with Tracers

4.4.1 T3 Field Activities

Five pre-treatment soil borings, T3-A through T3-E, were drilled and sampled on October 20, 1994. Lithologic logs were prepared, and soil samples were shipped to ORNL for

analysis. One piezometer was installed at boring T3-D, and three lysimeters were nested in boring T3-C (Fig. 4.11). Three SMT probes were installed approximately 2 ft east of the lysimeters. Water samples collected from the piezometer and lysimeters on November 1, 1994, were shipped to ORNL for analysis. Preparation for injection testing at test cell 3 began on the morning of November 18, 1994. A set of water samples was collected from the lysimeters and piezometer, and elevation benchmarks were surveyed prior to the start of injection. A proprietary bionutrient/surfactant solution with bromide and Snomax® as tracers was injected on 2-ft centers to a depth of 10.4 ft starting at 1:35 p.m. The test cell injection was completed at 5:45 p.m.; thus, 250 min were required to perform 144 injections (36 MPIS set-up locations with four injector locations per setup). The average time for each injection setup was 6.9 min; however, this includes stopping to mix the injection solution twice during the process (approximately 45 min).

The injection solution was mixed as follows: five batches of 500 gal each (2500 gal total), with each batch containing 50 gal of bionutrient/surfactant, 450 gal of water, 421 g of KBr, and 215 g of Snomax®. The target solution strength was 10% bionutrient/surfactant, 150 mg/L Br⁻, and 114 mg/L Snomax®.

The objective was to inject 2 gal per injector at each 15-in. depth interval. With four injectors per set-up location and eight positions for each injector, the target total for injection at each setup was 64 gal (4 injectors × 8 positions × 2 gal each); the target for 36 set-up locations was 2,304 gal (36 setups × 64 gal each) for the entire test cell. According to the flow meter at the pump, 2,200 gal of solution were injected, and according to flow meters on the MPIS unit, 2,160 gal were injected. The estimated amount of solution lost to the ground surface was 400 gal. Thus, the amount of solution injected into the subsurface was estimated to be 1800 gal. One sample of each of the six batches was collected during injection to evaluate the uniformity of the injected solutions.

Test Cell No.	Total solution injected	Target solution strength	Average actual solution strength	Duration of injection testing	Average time per setup	Total surface seepage
Т3	2200 gal	10% bionutrient surfactant 150 mg/L Br- 114 mg/L Snowmax®	10% bionutrient surfactant 150 mg/L Br- 114 mg/L Snowmax®	250 min	6.9 min	400 gal

Soil resistance and temperature were measured before, during, and after the injection testing. Resistance measurements showed that the soil was near saturation at 14 ft bgs prior to injection testing, but was not saturated at 8 and 4 ft bgs. However, within 30 min after injection started, the backfilled soil around the subsurface probes was saturated with injected solution, and solution began to run out the top of the protective casing onto the ground

surface. Soil temperature was monitored throughout the test, showing an increase of 4° F at 4 ft bgs, 1° F at 8 ft bgs, and no change at 14 ft bgs.

The bionutrient/surfactant appeared to react mildly at the surface, and a faint, musky odor was detectable. This odor persisted for several days after the test. At the peak of reactivity, brown foam up to 2 in. deep appeared at each injector location, but did not persist for more than an hour.

The first phase of post-treatment characterization began with the completion of five additional soil borings (T3-F through T3-J) to a depth of 12 ft each. The borings were continuously sampled with 48-in.-long megabore samplers, and the resulting 1.5-in.-diameter soil cores were extensively subsampled. Each core was divided into 1-ft segments and then subsampled as described in Sect. 4.2. Eh was measured on the core as it was subsampled; lithologic descriptions were also prepared.

Post-treatment water samples were collected from the lysimeters and piezometer three times during the 3 days following the injection. Similarly, elevation benchmarks (S1 - S5) were surveyed three times during the 3 days following the injection.

Another set of water samples was collected from the lysimeters and piezometer on December 5, 1994, approximately 2 weeks after the injection. During the third week after injection, December 12 to 14, 1994, two additional soil borings (T3-K and T3-L) were cored and sampled. The on-site subsampling performed was not as extensive as that performed during the first phase of post-treatment soil sampling and was limited to measuring Eh as the cores were divided into 1-ft segments, preparing lithologic logs, and shipping the cores to ORNL for additional analysis. An additional set of water samples was collected from the lysimeters and piezometer in December 1994, February 1995 and May 1996.

4.4.2 T3 Soil Sample Results

Post-treatment soil sample results are summarized in Table 4.7 and data trend graphs are presented in Appendix C. Results from pH measurements were similar for all boreholes, with values ranging from 4.7 to 7.5 and pH increasing with depth. These values are similar to the pre-treatment sample pH values; thus, the bionutrient injection did not effect a significant pH change.

Microbial biomass was evaluated using most probable number (MPN) techniques for aerobic and anaerobic heterotrophic populations. Both enumerations were based on turbidity being exhibited over a dilution range in 1% PTYEG medium (Balkwill) in screw-capped test tubes (Pfiffner 1994). Aerobic enumerations were set-up in a three tube MPN series dilution scheme. Anaerobic heterotrophic enumerations utilized the same media with the additions

of cysteine hydrochloride as the reducing agent to reach anaerobic conditions, rezaurin as Eh indicator, and nitrogen/CO₂ as the headspace gas mixture (Pfiffner 1994). Aerobic enumerations were set up in a single-series dilution scheme. The results of the microbial activity are presented in Table 4.8.

The analysis of tracers injected with the bionutrients yielded different results. Bromide was detected in the soil profile at test cell 3 from a depth of 0 to 11 ft bgs (Fig. 4.12), ranging from 1 to approximately 7 ppm, with the maximum concentration at 7 ft bgs. Snomax®, measured as INA, was detected in the soil at depths from 1 to 9 ft bgs and at 12 ft bgs (Fig. 4.13). However, maximum concentrations were detected in the top 1 ft of soil. Thus, it appears that the injection resulted in adequate distribution of the solution within the soil profile.

Soil Eh increased to a maximum of about 430 mV at 2 to 4 ft bgs and then generally decreased to around 200 mV at 12 ft bgs. This trend is similar to that observed for background borings.

Post-treatment soil moisture readings ranged from approximately 13 to 26 %. Comparison to pre-treatment, depth-specific moisture values shows that moisture content in the near-surface interval (0 to 4 ft) increased by approximately 3%. The intermediate depth interval (4 to 8 ft) and the deepest interval (8 to 12 ft) were unchanged.

Soil surface elevation data show that the surface of the test cell was raised approximately 0.5 ft by the injection and remained near that elevation through the last measurement period (3 months after injection). Core samples from both the pre-treatment and post-treatment borings were carefully logged, and visible pores and fractures were noted. No differences were seen between the number and appearance of fractures before and after injection.

4.4.3 T3Water Sample Results

Post-treatment water sample results are presented in Table 4.9 and data trend graphs are presented in Appendix C. The pH of the water in the lysimeters steadily increased following injection of the bionutrient solution (Fig. 4.14). Maximum pH increases in T3L1 and T3L2 were observed approximately 2 weeks after injection, followed by a drop during the next 2 weeks. The pH of T3L3 was initially lower than in the upper lysimeters; however, the pH continued to rise through the last sampling period (4 weeks after injection). This indicates a slow percolation of the reagent through the soil, as also shown by the bromide data (Fig. 4.15). A dramatic increase in bromide concentration is observed for T3L1 and T3L2 immediately following the injection period. Approximately 2 weeks after the injection, bromide was first observed in T3L3, and concentrations in the upper lysimeters began to decrease.

DO values were slightly higher after the injection, but were inconsistent and not significantly different from background values. Conductivity remained about the same before and after injection except in T3L2, the 8-ft-deep lysimeter, where the conductivity increased dramatically after testing (November 18, 1994). Nitrate concentrations ranged from 0.1 to 1.6 ppm. Concentrations in T3L1 and T3L2 were above pre-treatment background levels, but no increases were observed in T3L3 and T3P1. The bionutrient solution probably contained a nitrate source that resulted in increases in the shallower intervals.

Alkalinity values for test cell 3 ranged from approximately 200 to 530 mg/L, with a general trend of increase with depth. These values and trend are similar to those observed in the background cell.

Other analytes tested included Fe, Mn, Cl⁻, and SO₄², but since there were no noticeable changes in concentration in the first few samples, analysis for these parameters was discontinued, and no evaluation of the limited data sets was deemed necessary.

4.5 Test Cell 4: Lime Slurry

4.5.1 T4 Field Activities

Five pre-treatment soil borings, T4-A through T4-E, were drilled and sampled on October 23 and 24, 1994. Lithologic logs were prepared, and soil samples were shipped to ORNL for analysis. No piezometer was installed in this test cell; however, three lysimeters were nested in boring T4-C (Fig. 4.11). Three SMT probes were installed approximately 2 ft east of the lysimeters. Water samples were collected from the lysimeters on November 1, 1994, and were shipped to ORNL for analysis. Preparation for injection testing at test cell 4 began on November 13, 1994. A set of water samples was collected from the lysimeters, and elevation benchmarks were surveyed prior to the start of injection. A lime solution (calcium carbonate) was then injected on 2-ft centers to a depth of 10.4 ft starting at 11:20 a.m. The test cell injection was completed at 4:05 p.m.; thus, 285 min were required to perform 144 injections (36 MPIS set-up locations with four injector locations per setup). The average time for each injection setup was 7.9 min; however, this includes 130 min of lime-slurry mixing time.

The objective was to inject 2 gal per injector at each 15-in. depth interval. With four injectors per set-up location and eight positions for each injector, the target total for injection at each setup was 64 gal (4 injectors × 8 positions × 2 gal each), and the target for 36 set-up locations was 2,304 gal (36 set-ups × 64 gal each) for the entire test cell. According to flow meters on the MPIS unit, 2,304 gal were injected. The estimated amount of solution lost to the ground surface was 500 gal. Thus, the amount of solution injected into the subsurface was estimated to be 1800 gal.

The solution was mixed as five batches of 500 gal of water with 400 lbs of dry lime each with no tracer added. This mix was approximately 20% lime by weight. One sample was collected from each batch to evaluate the uniformity of the injected solution. The average injected solution was 10.7% lime by weight.

Test Cell No.	Total solution injected	Target solution strength	Average actual solution strength	Duration of injection testing	Average time per setup	Total surface seepage
T4	2300 gal	20% lime	10.7% lime	285 min	7.9 min	500 gal

Soil resistance and temperature were measured before, during, and after the injection testing. Resistance measurements showed that the soil was near saturation at 8 and 14 ft bgs prior to injection testing, but was not saturated at 4 ft bgs. However, soon after injection started, the backfilled soil around the subsurface probes was saturated with injected solution, and solution began to run out the top of the protective casing onto the ground surface. Soil temperature was monitored throughout the test and showed an increase of 2° F at 4 ft bgs and no change at 8 and 14 ft bgs.

The first phase of post-treatment characterization began with the completion of five additional soil borings (T4-F through T4-J) to a depth of 12 ft each. The borings were continuously sampled with 48-in.-long megabore samplers, and the resulting 1.5-in.-diameter soil cores were extensively subsampled. Each core was divided into 1-ft segments and then subsampled as described above. Eh was measured on the core as it was subsampled; lithologic descriptions were also prepared.

Post-treatment water samples were collected from the lysimeters eight times during the days following the injection. Similarly, elevation benchmarks (S1 - S5) were surveyed six times during the eight days following the injection.

Another set of water samples was collected from the lysimeters on December 5, 1994, approximately three weeks after the injection. During the fourth week after injection, December 12 to 14, 1994, two additional soil borings (T4-K and T4-L) were cored and sampled. The subsampling performed was not as extensive as that performed during the first phase of post-treatment soil sampling and was limited to dividing cores into 1-ft segments, preparing a lithologic log, and shipping the cores to ORNL for analysis. An additional set of water samples was collected from the lysimeters in December 1994, February 1995 and May 1996.

4.5.2 T4 Soil Sample Results

Post-treatment soil results for test cell 4 are summarized in Table 4.10 and data trend graphs are presented in Appendix C. Post-treatment pH values ranged from 4.5 to 12.6 (Fig. 4.17). Approximately one-half of these values are greater than the maximum value measured during pre-treatment sampling. The greatest increases over pre-treatment values occurred between 0 to 2 ft and between 5 to 11 ft. Values for the 3- to 5-ft interval were near background levels. Generally, it appears that the MPIS was effective in distributing the lime slurry throughout the soil profile. Indeed, post-treatment soil moisture readings were higher than pre-treatment measurement for all depths, excluding the 7- to 8-ft interval.

Post-treatment calcium concentrations are shown in Fig. 4.18. Although depth-specific results are sporadic between boreholes, concentrations are generally higher than the pretreatment samples. This is particularly true for the 0- to 6-ft intervals, where dramatic increases were observed. The scattered nature of the results indicates flow of the slurry through existing preferential flow channels (e.g., fractures).

Soil surface elevation data show that the surface of the test cell was raised approximately 0.15 ft by the injection and remained near that elevation for approximately one month. An additional 0.10-ft rise was observed during the last measurement period (three months after injection). Core samples from both pre-treatment and post-treatment borings were carefully logged, and visible pores and fractures were noted. Post-treatment logs indicated abundant lime in root pores and microfractures in the 0- to 2-ft interval. Some lime was seen in both vertical and horizontal fracture and bedding planes and in remnant root vesicles in the 4- to 8-ft interval.

4.5.3 T4 Water Sample Results

Post-treatment water sample results are presented in Table 4.11 and data trend graphs are presented in Appendix C. The lime injection resulted in an immediate increase in the pH of the water in the two shallower lysimeters, T4L1 and T4L2 (Fig. 4.19). Increases from 7.8 to approximately 12.4 were observed, and pH remained fairly constant for 5 weeks, at which time values began decreasing. No increases in the pH of water samples from the deep lysimeter (T4L3) were observed. Alkalinity and conductivity values correlate well with the pH measurements, indicating that the short-term influence of the lime injection primarily affected the 0 to 8-ft interval. Alkalinity values in T4L1 and T4L2 increased by 700 to 1000 mg/L following the injection and remained constant for approximately 4 weeks; however, a slight increase (approximately 200 mg/L) was also observed in T4L3 (Fig. 4.20). A similar pattern was observed for conductivity measurements; however, the deeper lysimeter showed dramatic increases above background concentrations (Fig. 4.21). In fact, values for all three lysimeters more than doubled following the injection and remained elevated for approximately

4 weeks. These data indicate that the calcium ions and the various carbonate species from the lime injection significantly affected the soil-pore water in test cell 4.

Post-treatment DO values ranged from approximately 2 to 12 mg/L. No significant changes from background values were observed. Other analytes tested included Fe, Mn, Cl⁻, and $SO_4^{\ 2}$, but since there were no noticeable changes in concentration in the first few samples, analysis for these parameters was discontinued, and no evaluation of the limited data sets was deemed necessary.

4.6 Test Cell 5: Potassium Permanganate

4.6.1 T5 Field Activities

Five pre-treatment soil borings, T5-A through T5-E, were drilled and sampled on October 21 and 22, 1994. Lithologic logs were prepared, and soil samples were shipped to ORNL for analysis. One piezometer was installed at boring T5-D, and three lysimeters were nested in boring T5-C (Fig. 4.22). Three SMT probes were installed approximately 2 ft east of the lysimeters. Water samples collected from the piezometer and lysimeters on November 1, 1994, were shipped to ORNL for analysis. Preparation for injection testing at test cell 5 began the morning of November 16, 1994. A set of water samples was collected from the lysimeters and piezometer, and elevation benchmarks were surveyed prior to the start of injection. A solution of 500 gal of water and 100 kg of KMnO₄ was mixed for injection (5% KMnO₄ by weight). The KMnO₄ solution was injected on 2-ft centers to a depth of 10.4 ft starting at 1:40 p.m. The test cell injection was completed at 5:40 p.m.; thus, 240 min were required, with an average time for each injection setup of 6.7 min. This time included stopping twice to mix more injection solution (approximately 45 min).

The objective was to inject 2 gal per injector at each 15-in. depth interval or a total of 2,304 gal for the entire test cell. According to the flow meter at the pump, 2,223 gal were injected, and according to flow meters on the MPIS unit, 2,328 gal were injected. The estimated amount of solution lost to blowout at the ground surface was 400 gal. Thus, the amount of solution injected into the subsurface was estimated to be 1900 gal. One sample of each of five batches was collected during injection to evaluate the uniformity of the injected solution. The average injected solution was 4.2 % KMnO₄ by weight.

Test Cell No.	Total solution	Target solution	Average actual	Duration of injection	Average time per	Total surface
	injected	strength	solution strength	testing	setup	seepage
T5	2300 gal	5% KMnO ₄	4.2% KMnO ₄	240 min	6.7 min	400 gal

Soil resistance and temperature were measured before, during, and after the injection testing. Resistance measurements showed that the soil was near saturation at 8 and 14 ft bgs prior to injection testing, but was not saturated at 4 ft bgs. However, about 40 min after injection started, the backfilled soil around the subsurface probes was saturated with injected solution, and solution began to run out the top of the protective casing onto the ground surface. Soil temperature was monitored throughout the test and showed an increase of 2° F at 4 ft bgs, 1° F at 8 ft bgs, and no change at 14 ft bgs. The KMnO₄ reacted mildly upon injection, with only small amounts of gas produced; however, the fact that the soil-pore water samples were still dark purple 4 weeks later indicated that the reagent was persistent in the soil.

The first phase of post-treatment characterization began with the completion of five additional soil borings (T5-F through T5-J) to a depth of 12 ft each. The borings were continuously sampled with 48-in.-long megabore samplers, and the resulting 1.5-in.-diameter soil cores were extensively subsampled as described in Sect. 4.2. Each core was divided into 1-ft segments and then subsampled. Eh was measured on the core as it was subsampled; lithologic descriptions were also prepared.

Post-treatment water samples were collected from the lysimeters and piezometer 5 times during the 5 days following the injection. Similarly, elevation benchmarks (S1, S2, S3 damaged, S4, and S5) were surveyed 4 times during the 5 days following the injection.

Another set of water samples was collected from the lysimeters and the piezometer on December 5, 1994, approximately 3 weeks after the injection. During the fourth week after injection, December 12 to 14, 1994, two additional soil borings (T5-K and T5-L) were cored and sampled. The on-site subsampling performed was not as extensive as that performed during the first phase of post-treatment soil sampling and was limited to preparing a lithologic log and segmenting the core for shipment to ORNL for analysis. An additional set of water samples was collected from the lysimeters and piezometer in December 1994, February 1995 and May 1996.

4.6.2 T5 Soil Sample Results

Post-treatment soil sample results are summarized in Table 4.12 and data trend graphs are presented in Appendix C. Soil sample results show a pH ranging from 4.3 to 7.8, with a trend

of increasing pH values with depth. These values and trend are similar to the pre-treatment sample results; thus, no significant changes in pH were observed. Purple staining was noted in many of the soil extracts indicating that the KMnO₄ had penetrated the soil. Core samples from the post-treatment borings showed purple staining in the upper 8-in. of soil from surface infiltration. The deepest soil sample with purple-colored soil extract was the 8 to 9-ft sample, and this condition was noted in three of the five borings. Purple-stained zones were observed on the soil cores at depths of 5 and 9 ft bgs.

Compared to pre-treatment conditions, post-treatment soil moisture content increased slightly in the 0- to 4-ft interval, decreased slightly in the 5- to 8-ft interval, and remained approximately the same in the 9- to 12-ft interval. Therefore, it appears that the bulk of the solution was injected in the top 4 to 5 ft of the soil. This is also indicated by the post-treatment manganese concentrations measured (Fig. 4.23). Manganese concentrations at depths of 0 to 5 ft bgs ranged from approximately 20 to 250 mg/L above background levels; however, no increases were observed in the 6- to 12-ft interval. Post-treatment Eh measurements ranged from 240 to 880 mV (Fig. 4.24). Dramatic increases in Eh were observed throughout the soil profile, where readings were approximately double pre-treatment measurements with the exception of the 0- to 2-ft interval. Thus, this treatment appeared to have the greatest effect on soil Eh.

Soil surface elevation data show that the surface of the test cell was raised approximately 0.1 ft by the injection and remained near that elevation through the last measurement period (three months after injection). Core samples from both pre-treatment and post-treatment borings were carefully logged and visible pores and fractures were noted. No differences were seen between the number and appearance of fractures before and after injection, excluding the purple staining noted above.

4.6.3 T5 Water Sample Results

Post-treatment water sample results are presented in Table 4.13 and data trend graphs are presented in Appendix C. Manganese results ranged from background (<1 mg/L) to approximately 18,000 mg/L (Fig. 4.25). The maximum concentration was detected the day after injection in the intermediate-depth lysimeter, T5L2 (8 ft bgs). Concentrations in T5L2 slowly dropped to near background levels over a 5-week period. Concentrations in the shallow lysimeter, T5L1, followed a similar trend, however, the maximum concentration was lower (4,500 mg/L). The initial field description of water collected from the lysimeters was "very purple" and "purple" respectively. A maximum concentration of 50 mg/L was detected in the deepest lysimeter, T5L3, the day after injection. The maximum concentration in the piezometer was 8 ppm. These results indicate that injection was effective in distributing the solution to the top 8 ft of the test-cell soil.

Post-treatment pH results ranged from approximately 6.0, to 8.1. Initial average values for individual lysimeters were similar to pre-treatment levels, excluding T5L1, where an average 0.5 pH unit decrease was observed. The lowest value observed in T5L1 occurred 5 days after injection. DO values were difficult to determine because the purple color of the water interfered with the colorimetric analysis. Due to intermittent data collection and questions regarding the validity of the measurements, no conclusions can be drawn from the DO data.

Electrical conductivity results were scattered; however, post-treatment results for T5L1 and T5L2 were generally higher than pre-treatment readings. No significant changes were observed in T5L3 or the piezometer. Post-treatment alkalinity values ranged from approximately 100 to 630 mg/L. No pre-treatment alkalinity measurements were taken in test cell 5; thus, values were compared to samples taken from the background cell. Values for all depths at test cell 5 are approximately twice those from the background cell; however, this difference is likely a result of natural geochemical variation rather than an effect caused by the solution injection. Other analytes tested included Fe, Mn, Cl⁻, and SO₄²⁻, but since there were no noticeable changes in concentration in the first few samples, analysis for these parameters was discontinued, and no evaluation of the limited data sets was deemed necessary.

4.7 Test Cell 6: Air

4.7.1 T6 Field Activities

Five pre-treatment soil borings, T6-A through T6-E, were drilled and sampled on October 21, 1994. Lithologic logs were prepared, and soil samples were shipped to ORNL for analysis. Three lysimeters were nested in boring T6-C (Fig. 4.26). Three SMT probes were installed approximately 2 ft east of the lysimeters. Water samples collected from the lysimeters on November 1, 1994, were shipped to ORNL for analysis. Preparation for injection testing at test cell 6 began the morning of November 19, 1994. A set of water samples was collected from the lysimeters, and elevation benchmarks were surveyed prior to the start of injection. Air was injected with two injectors at four locations within the test cell to a depth of 10.4 ft starting at 4:10 p.m. The test cell injection was completed at 4:30 p.m.; thus, 20 min were required, with an average time for each injection setup of 5 min.

The objective was to inject air at each setup until air pressure reached at least 100 psi or until surface blowout was achieved. Three additional locations south of the test cell were penetrated with the injectors. However, no air was injected into these three holes since a gaseous-phase tracer study was planned at a later date to evaluate the extent of air-induced fracturing from the air injections performed within the test cell. All injector holes, including the three locations south of the test cell, were completed by placing 1/4-in-diameter tubing in the holes and backfilling with sand and bentonite.

The first phase of post-treatment characterization began with the collection of water samples from the lysimeters three times during the three days following the injection. Additional sets of water samples were collected from the lysimeters on December 5, 1994, approximately 3 weeks after the injection; on December 15, 1994, approximately 4 weeks after injection; on February 15, 1995, approximately 13 weeks after injection; and on May 7, 1996, approximately one and a half years after injection.

4.7.2 T6 Soil Sample Results

No post-treatment soil samples were collected from test cell 6. Soil surface elevation data show no significant increases in elevation following the air injection.

4.7.3 T6 Water Sample Results

Post-treatment water sample results are presented in Table 4.14. Results for test cell 6 are limited due to poor sample recovery. However, little change was expected from the air injection. Because the upper lysimeter was consistently dry, only one sample was collected from T6L1. Results for pH, DO, and conductivity measurements are within the background range. Other analytes tested included Fe, Mn, Cl⁻, NO₃⁻, and SO₄²⁻, but since there were no noticeable changes in concentration in the first few samples, analysis for these parameters was discontinued, and no evaluation of the limited data sets was deemed necessary.

4.8 Test Cell 7: Micropowder Iron

4.8.1 T7 Field Activities

One pre-treatment soil boring, T7-A, was drilled and sampled on October 24, 1994. A lithologic log was prepared, and soil samples were shipped to ORNL for analysis. No piezometer was installed in this test cell; however, three lysimeters were nested in boring T7-A (Fig. 4.27). Three SMT probes were installed approximately 2 ft east of the lysimeters. Water samples collected from the lysimeters on November 1, 1994, were shipped to ORNL for analysis. Preparation for injection testing at Test Cell 7 began the morning of November 19, 1994, when a set of water samples was collected from the lysimeters. A solution of iron micropowder (approximately 20% by weight), water, and guar gum was injected on 2-ft centers to a depth of 10.4 ft at the 4 ft by 8-ft test cell. The injection started at 12:40 p.m. and was completed at 1:02 p.m.; thus, 22 min were required to perform 16 injections (four MPIS set-up locations with four injector locations per setup). The average time for each injection setup was 5.5 min.

The objective was to inject 2 gal per injector at each 15-in. depth interval. With four injectors per set-up location and four positions for each injector, the target total for injection at each setup was 64 gal (4 injectors × 8 positions × 2 gal each), and the target for four set-up locations was 256 gal (4 setups × 64 gal each) for the entire test cell. According to the flow meter at the pump, 308 gal were injected. Because the flow meters on the MPIS unit were not working, no readings were obtained from them. The estimated amount of solution lost to the ground surface was 20 gal. Thus, the amount of solution injected into the subsurface was estimated to be 290 gal. The solution was mixed as a single batch of 500 gal of water with 400 lbs of dry iron micropowder (5 um nominal particle size) and 25 lbs of guar gum. The guar gum was added to keep the iron in suspension during injection. However, the tank required continuous stirring to prevent settling of the iron micropowder. The mixture was approximately 20% iron by weight.

Test Cell No.	Total solution injected	Target solution strength	Average actual solution strength	Duration of injection testing	Average time per setup	Total surface seepage
Т7	308 gal	20% iron	not	22 min	5.5 min	20 gal

Soil resistance and temperature were measured before, during, and after the injection testing. Resistance measurements showed that the soil was near saturation at 14 ft bgs prior to injection testing, but was not saturated at 4 and 8 ft bgs. However, soon after injection started, the backfilled soil around the subsurface probes was saturated with injected solution, and solution began to run out the top of the protective casing onto the ground surface. Soil temperature was monitored before and after the test and showed an increase of 2° F at 4 ft bgs, 1° F at 8 ft bgs, and no change at 14 ft bgs.

The first phase of post-treatment characterization began with the completion of two additional soil borings (T7-B through T7-C) to a depth of 12 ft each. The borings were continuously sampled with 48-in.-long megabore samplers, and the resulting 1.5-in.-diameter soil cores were extensively subsampled. Each core was divided into 1-ft segments and then subsampled as described in Sect. 4.2. Eh was measured on the core as it was subsampled; lithologic descriptions were also prepared.

Post-treatment water samples were collected from the lysimeters twice during the 2 days following the injection. Similarly, the elevation benchmark was surveyed two times during the 2 days following the injection. Another set of water samples was collected from the lysimeters on December 5, 1994, approximately 2 weeks after the injection. During the third week after injection, December 12 to 14, 1994, two additional soil borings (T7-D and T7-E) were cored and sampled. The on-site subsampling performed was not as extensive as that

performed during the first phase of post-treatment soil sampling and was limited to dividing cores into 1-ft segments, preparing a lithologic log, and shipping the cores to ORNL for analysis. An additional set of water samples was collected from the lysimeters in December 1994, February 1995 and May 1996.

4.8.2 T7 Soil Sample Results

Post-treatment soil samples are summarized in Table 4.15 and data trend graphs are presented in Appendix C. Samples were analyzed for pH, Eh, and iron. The pH ranged from 5.2 to 7.1, with a trend of increasing values with depth. These values are within the background range of samples collected from the site. Post-treatment Eh values ranged from 135 to 420 mV, increasing to a maximum at 2 to 3 ft bgs and then steadily decreasing with depth. These values and trend are consistent with sample data from the background cell.

Soil iron results showed concentrations ranging from less than 1 ppm to over 30 ppm. No pre-treatment samples were analyzed for iron; thus, a direct comparison cannot be made. Also, analytical results cannot be compared to a regional background range of values due to differences in the analysis methods (water extraction vs acid digestion). However, based on a relative comparison of the post-treatment data collected from test cell 7, substantial increases in iron were observed at the 1 to 3-ft interval and the 4 to 5-ft interval of borehole C. The particle size of the micropowder iron was approximately 5 μ m and was suspended with the guar gum.

Soil surface elevation data show that the surface of the test cell was raised approximately 0.1 ft by the injection, and remained near that elevation through the last measurement period (3 months after injection). Core samples from both pre-treatment and post-treatment borings were carefully logged, and visible pores and fractures were noted. Guar gum was observed in fractures and voids at 1 to 3 ft bgs and at 6 to 7 ft bgs.

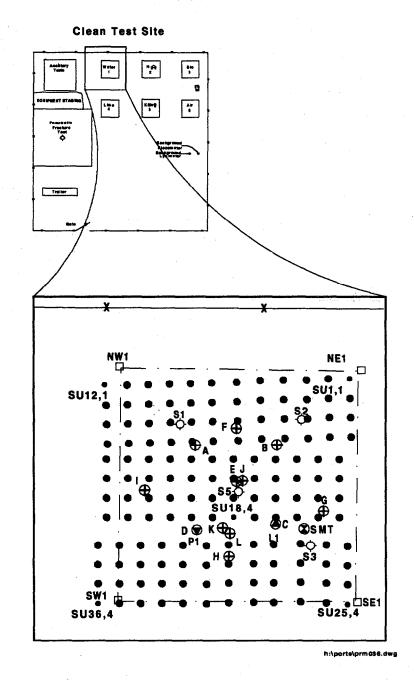
4.8.3 T7 Water Sample Results

Post-treatment water sample results are presented in Table 4.16 and data trend graphs are presented in Appendix C. Post-treatment iron concentrations in test cell 7 ranged from 0.1 to over 30 ppm (Fig. 4.28). Two samples from the test cell (December 15 samples from T7L1 and T7L2) were recorded as "over range" by the analysis method. The upper limit of detection for the analysis was approximately 30 ppm. These points are graphed as 30 ppm on Fig. 4.28. Background concentrations of iron in water, based on analyses from other test cells, ranged from 0.1 to 1.2 ppm. Therefore, iron levels above this range are considered significant. No effects were observed in lysimeter samples collected immediately after the injection (days 1 through 3); however, dramatic increases were observed in T7L1 and T7L2

approximately 2 weeks later. This evidence suggests that some of the injected iron was reduced to soluble Fe⁺² between sampling periods. Iron levels in these lysimeters began to drop after approximately 4 weeks. Concentrations in the deep lysimeter, T7L3, steadily increased to a maximum of 1.8 ppm at the last sampling period. This was possibly the result of percolation from the upper soil zone.

Post-treatment pH values from lysimeter samples ranged from 6.2 to 8.8. There is a trend of decreasing pH in the upper lysimeters, T7L1 and T7L2, following the injection period. This correlates with the reduction of iron discussed above. Hydrogen ions, produced as a result of iron reduction, would effect a decrease in pH. Measurements in T7L3 were sporadic; however, there is no clear trend with respect to pH.

DO values are scattered, and no significant changes are indicated when compared to background measurements. It appears that DO increased and then decreased after the injection. However, this is attributed to variation in the field measurements observed at all test cells. Conductivity values ranged from approximately 600 to 4400 μmhos. Conductivity from the deepest lysimeter, T7L3, appears to have increased over background levels; however, no significant changes were observed in T7L1 and T7L2. Other analytes tested included alkalinity, Mn, Cl⁻, NO₃⁻, and SO₄⁻, but since there were no noticeable changes in concentration in the first few samples, analysis for these parameters was discontinued, and no evaluation of the limited data sets was deemed necessary.



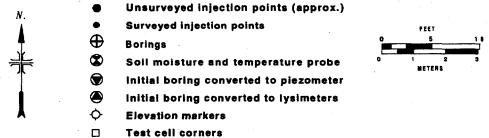


Fig. 4.1. Test cell 1 sample locations.

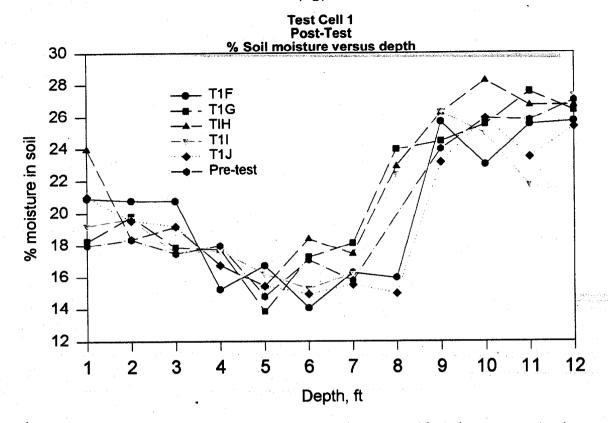


Fig. 4.2. Test cell 1 - post-treatment soil moisture content.

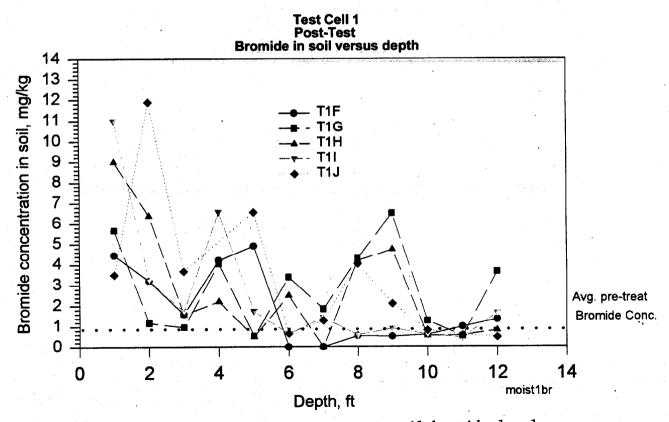


Fig. 4.3. Test cell 1 - post-treatment soil bromide levels.

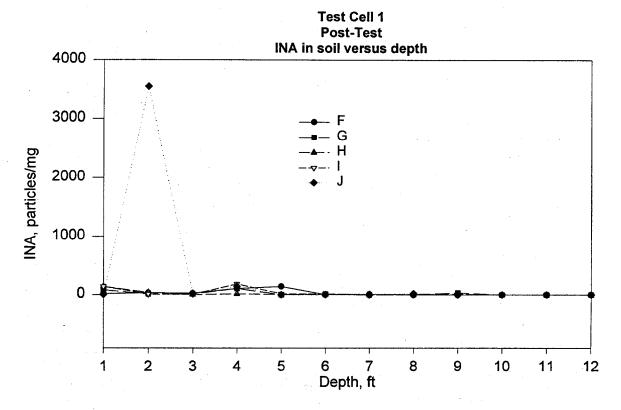


Fig. 4.4. Test cell 1 - post-treatment soil INA levels.

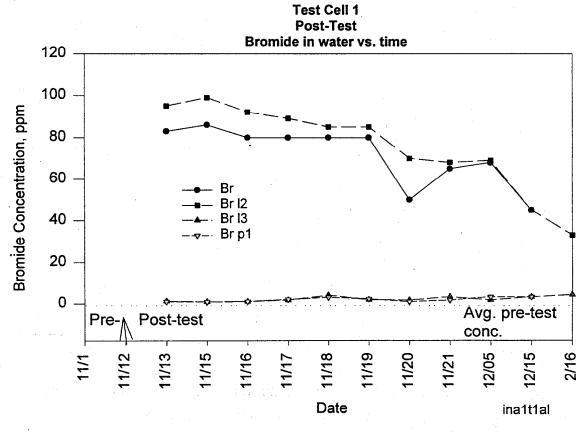
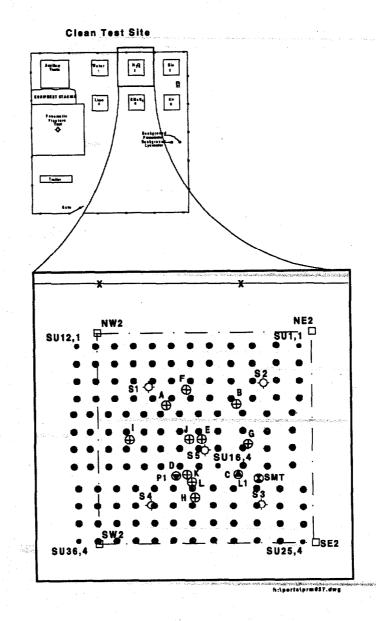


Fig. 4.5. Test cell 1 - post-treatment water bromide levels.



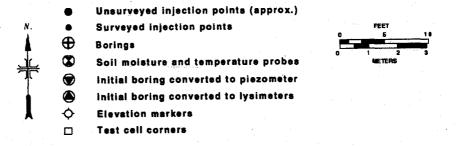


Fig. 4.6. Test cell 2 sample locations.

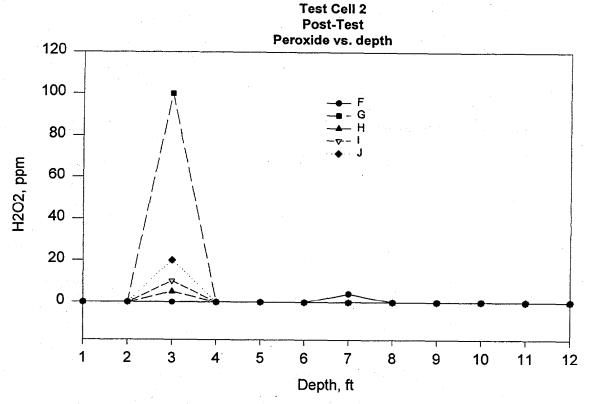


Fig. 4.7. Test cell 2 - post-treatment soil peroxide levels.

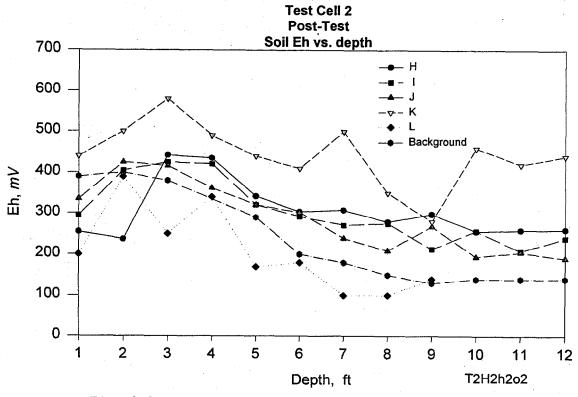
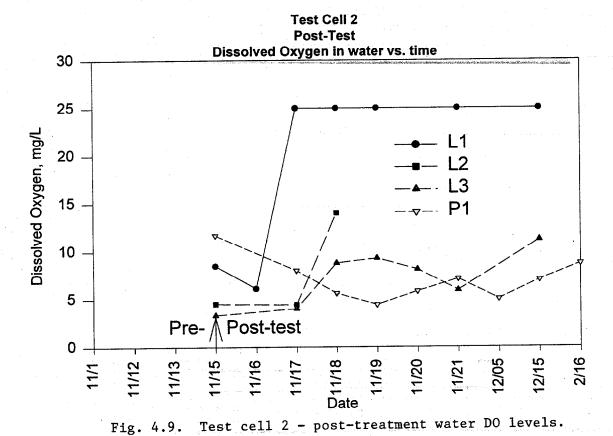


Fig. 4.8. Test cell 2 - post-treatment soil Eh data.



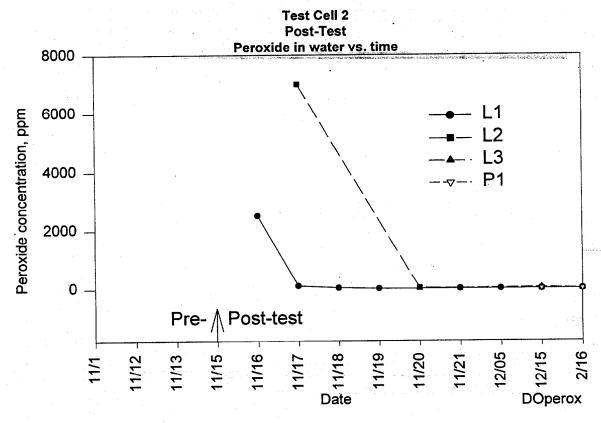
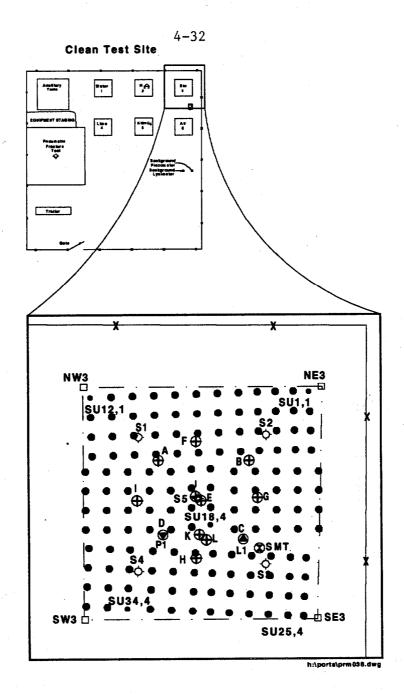


Fig. 4.10. Test cell 2 - post-treatment water peroxide levels.



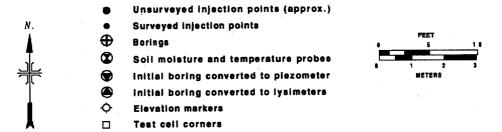


Fig. 4.11. Test cell 3 sample locations.

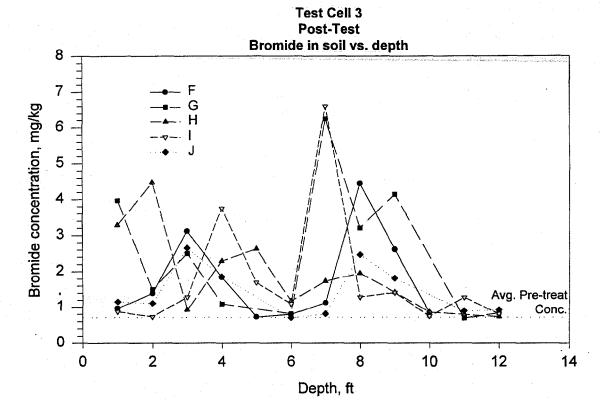


Fig. 4.12. Test cell 3 - post-treatment soil bromide levels.

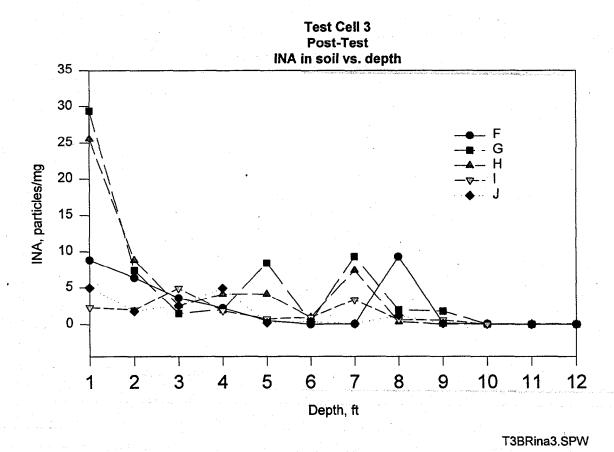


Fig. 4.13. Test cell 3 - post-treatment soil INA levels.

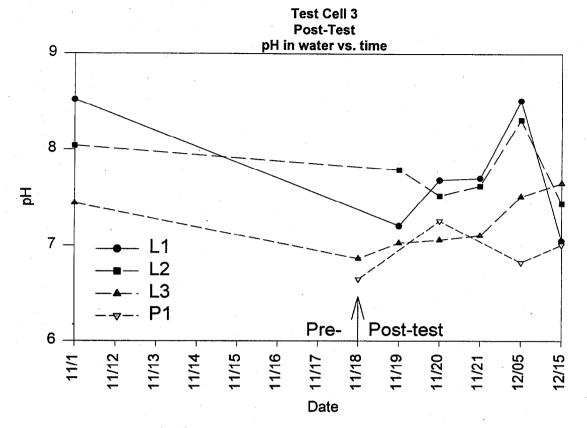


Fig. 4.14. Test cell 3 - post-treatment water pH.

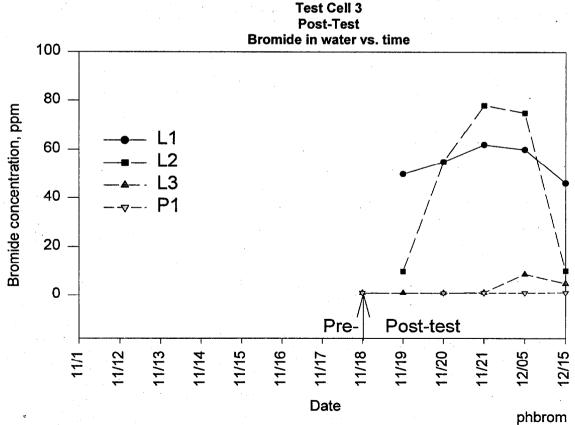
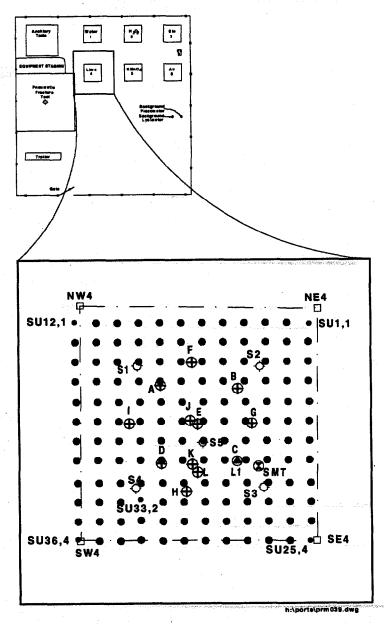


Fig. 4.15. Test cell 3 - post-treatment water bromide data.

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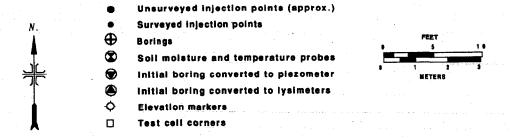


Fig. 4.16. Test cell 4 sample locations.

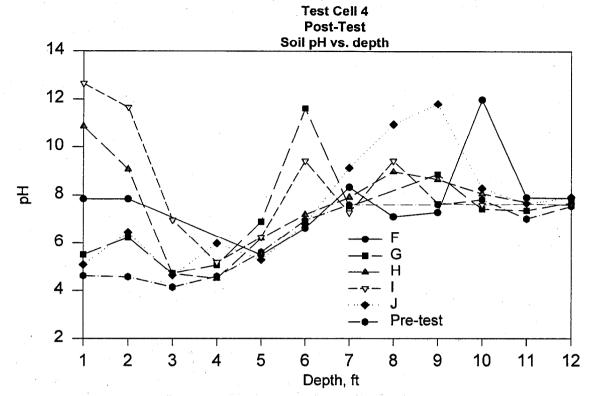


Fig. 4.17. Test cell 4 - post-treatment soil pH.

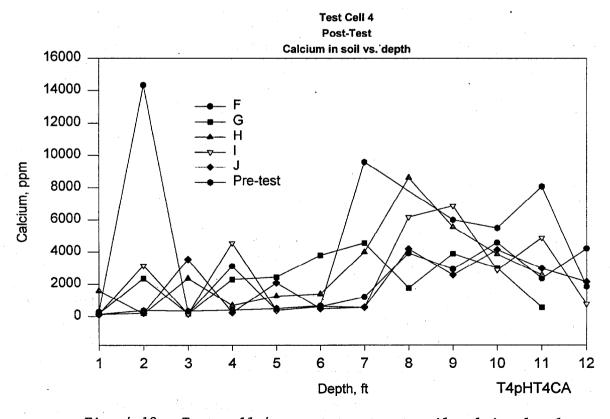


Fig. 4.18. Test cell 4 - post-treatment soil calcium levels.



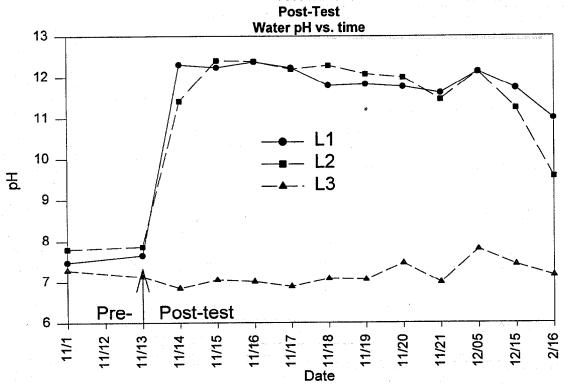


Fig. 4.19. Test cell 4 - post-treatment water pH.

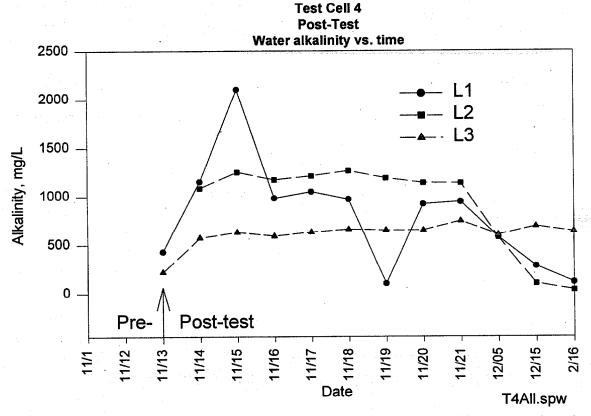


Fig. 4.20. Test cell 4 - post-treatment water alkalinity levels.

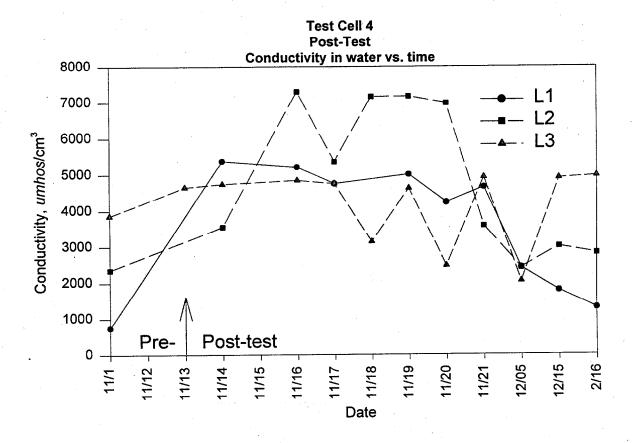
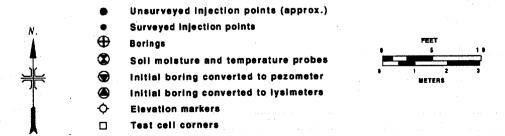


Fig. 4.21. Test cell 4 - post-treatment water conductivity levels.

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Fig. 4.22. Test cell 5 sample locations.

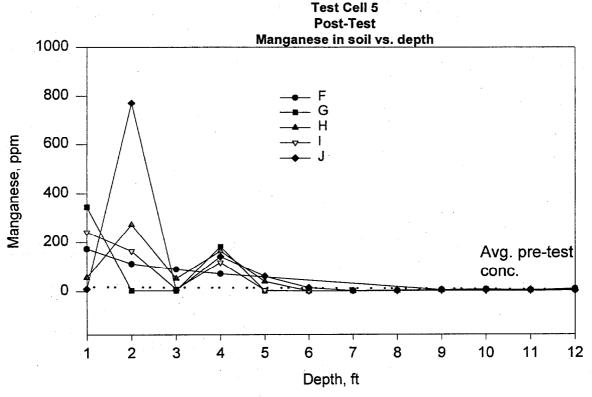
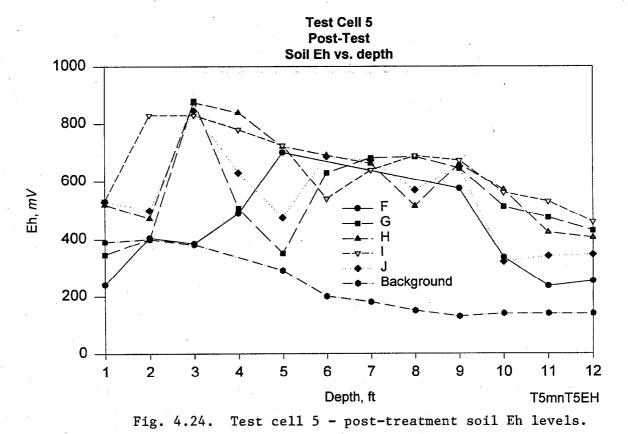


Fig. 4.23. Test cell 5 - post-treatment soil manganese levels.



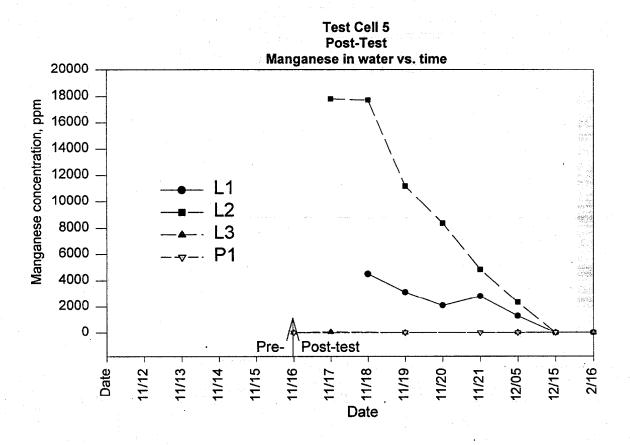
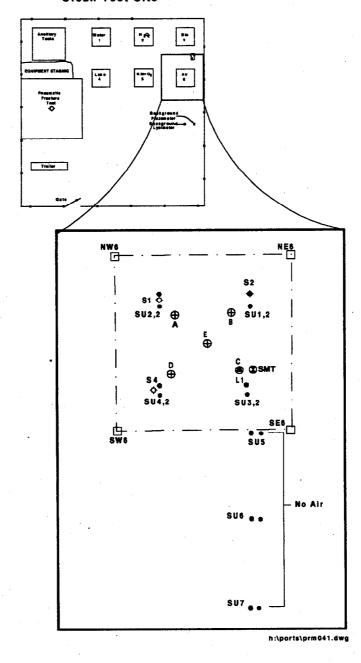


Fig. 4.25. Test cell 5 - post-treatment water manganese levels.

T5MANG

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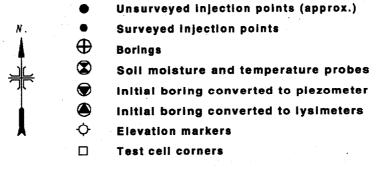
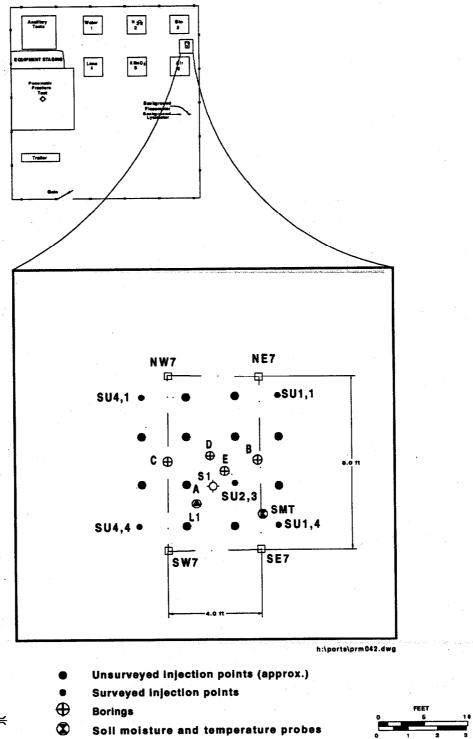


Fig. 4.26. Test cell 6 sample locations.

Clean Test Site



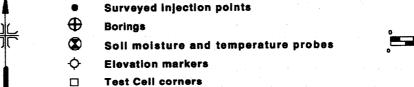


Fig. 4.27. Test cell 7 sample locations.

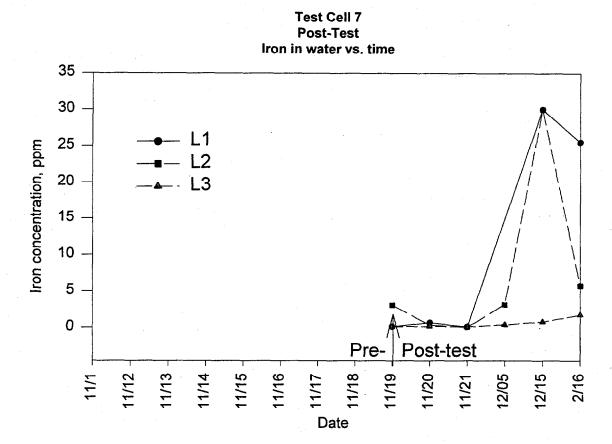


Fig. 4.28. Test cell 7 - post-treatment water iron data.

T7IRON

Table 4.1. Summary of background soil results

Depth,	Soil moisture, ^a %	Moist bulk density, g/cm ³	pH ^a	Eh, ^b mV	TOC, ^a ppm	
1	17	1.93	4.9	390	6397	
2	18	1.66	4.5	400	1630	
3	19	1.79	4.3	380	1448	
4	15	1.83	4.7	. 	1082	
5	15	1.99	5.7	290	1021	
6	16	1.96	6.5	200	995	
7	18	. .	6.6	180	615	
8	21	2.07	7.0	150	558	
9	24	1.90	7.2	130	395	
10	26	1.81	7.6	140	421	
11	24	1.87	7.4	140	543	
12	24	1.95	7.4	140	526	

^a Values averaged from borehole E in test cells 1 through 6, from borehole A in test cell 7 and the background borehole in the shakedown area.

b Values from borehole 1 in the shakedown area (S1BA)

^{-- =} No measurement taken

Table 4.2. Water sample results from background piezometer and lysimeters

								•				
Sample location	Date sampled	Temperature,	pН	DO, mg/L	Electrical conductivity, µmhos	Alkalinity, mg/L	Br, mg/L	Fe, mg/L	Mn, mg/L	Cl ⁻ , mg/L	NO ₃ -, mg/L	SO ₄ ² ·, mg/L
B1L1	11/01/94	12.4	8.7		960							
2.0.	11/12/94	17.4	8.3		1112	132		0	0.25	23.3	0.1	550
	11/13/94	14.3	7.9		1250	319	1.0	0	0.25			525
	11/14/94	12.5	8.0	7.3	1390	157		0.15	0.25	15.0		
	11/15/94	13.1	7.8	3.8	1430					16.8		
	11/18/94	15.6	7.9	4.8	1440	216	0.9	0		9.5	∶0.4	475
	11/21/94	13.7	7.7	3.7	1520	.153	0.9	0	0	11.0	0	650
	12/05/94		7.9									
	02/16/95	3.9	8.0	2.5	1076	94	0.6	0.15	0	_	0	
B1L2	11/01/94 11/12/94	12.9	8.9	.*								
	11/13/94	15.4	7.9	3.6	2112	204	0.9	0.05	0.25			1075
	11/14/94	13.8	8.1	3.3	2190	229		0.10	0.25	10.5		
	11/15/94	14.5	7.2	4.7	2210					9.0		
	11/18/94	15.1	8.1	8.1	1430	252 .	0.9	0	0.30	11.2	0.1	1100
	11/21/94	13.7	7.4	2.2	2330	248	0.8	0.25	0.17	7.8	0.2	1200
	12/15/94	10.6	8.3	4.5	1130	99	8.0	0	0.18		0.1	
	02/16/95	4.9	7.7	3.4	2910	246	0.3	0	0.20			
B1L3	11/01/94	12.5	7.9		1530							
27.23	11/12/94	20.6	7.5	2.7	1652	212		O ·	0.25	7.8	0	800
	11/13/94	15.6	7.4	2.8	1700	186	0.9	0.05	0.25			750
	11/14/94	14.2	7.4	4.9	1760 .	240		0.10	0.25	6.3		
	11/15/94	14.1	6.6	6.2	1800					6.3		
	11/18/94	15.3	8.1	4.2	1790	277	0.9	0		3.2 5.2	0.1	
	11/21/94	14.9	7.1	2.8	1700	238	0.7	0	0.17	5.2	0	750
	12/05/94	•	8.2			213	8.0	0.02	0.10			850
	12/15/94	12.3	8.3	11.2	2530	260	0.8	0.025	0.90			
	02/16/95	6.3	6.6	2.3	1730	218	0	0.025	0		0.1	

Table 4.2. (continued)

Sample location	Date sampled	Temperature, °C	pН	DO, mg/L	Electrical conductivity, μmhos	Alkalinity, mg/L	Br, mg/L	Fe, mg/L	Mn, mg/L	Cl ⁻ , mg/L	NO ₃ , mg/L	SO ₄ ² -, mg/L
B1P1	11/01/94	12.3	7.2		1480							
duplicate	11/01/94	12.3	7.2		1480							
•	11/12/94	17.2	6.6	1.9	1547	188		0.125	0.25	5.5	0.1	750
	11/13/94	16.3	7.1	4.2	1520	169	0.9	0.075	0.25			650
	11/14/94	13.9	6.6	2.6	1600	174		0.125	0.25	9.5		
duplicate	11/14/94			4.4		182		1.7		5.0		
•	11/15/94	14.1	6.6	2.0	1590		0.9			5.0		
	11/18/94	15.3	6.8	2.9	1580	194	0.9	0.05	1.00	2.5	0.1	425
•	11/21/94	15.0	6.1	4.9	1520	198	0.8	0.18		6.5	0	700
	12/05/94	15.3	6.9	3.1	1240	190	0.7	0.08	0.10			725
	12/15/94	15.0	7.0	4.3	1550	198	0.8	0.23	0.36			
	02/16/95	7.3	7.5	NA	1650	192	0.1	NA	0		0 .	

See Fig. 3.1 for boring locations.

Table 4.3. Tracer test cell (T1) soil sample results

Depth, ft	рН	Eh, mV	TOC, ppm	% Soi	l moisture	Bromide, ppm	INA, particles/mg
₹** •	Pre ^a	Post ^b	Prea	Pre ^a	Post ^b	Post ^b	Post ^b
1	4.3	360 to 380	6258	18.0	18.3 to 24.0	4 to 11	4 to 178
2	4.4	350	1295	18.4	18.4 to 20.8	1 to 12	12 to 3543
3	4.6	390 to 460	2130	17.5	17.6 to 20.8	1 to 4	4 to 26
4	4.6	390 to 430	1471	18.0	15.7 to 17.8	2 to 7	18 to 181
5	4.8	395 to 490	1226	14.8	13.9 to 18.7	1 to 7	2 to 138
6	6.0	370 to 390	1216	17.1	14.1 to 18.4	1 to 3	3 to 16
7	7.0	280 to 350	685	15.8	15.5 to 18.2	1 to 2	1 to 9
8		305 to 325			15.0 to 24.0	1 to 4	0 to 20
9	7.1	320 to 340	354	24.0	23.2 to 26.3	1 to 7	0 to 34
10	7.8	295 to 305	333	25.9	23.0 to 28.3	1 to 2	0 to 1
11	7.6	305 to 330	653	25.8	21.7 to 27.5	1	0
12	7.4	275 to 330	555	27.0	25.4 to 27.3	1 to 4	0

Pre-treatment results
 Ranges of values are from five post-treatment soil borings.
 No sample taken

Table 4.4. Water sample results for the tracer test cell (T1), injection date 11/12/95

Sample location	Date sampled	Temperature, °C	рH	DO, mg/L	Electrical conductivity, μmhos	Alkalinity, mg/L	Br, mg/L	Fe, mg/L	Mn, mg/L	Cl ⁻ , mg/L	NO ₃ -, mg/L	SO ₄ ² , mg/L
T1L1	11/01/94	12.8	8.4		1270	137.1-		;			2,	
	11/12/94		6.6	13.9				0.075	0.25	22.50	1.0	600
	11/13/94	11.6	6.6	8.2	1311	148	83 86	0	0.25	OR		425
	11/15/94		7.1	6.9		220		0.15		OR		475
	11/16/94	12.7	6.9	6.1	1410	244	80					
	11/17/94	13.4	6.8	5.2	1700		80	0.3				
	11/18/94	14.3	6.7	5.0	1400	. 15.75 15.75	80					
	11/19/94	12.9	6.9	6.4	1270		80					
	11/20/94	11.7	7.0	5.2	1230	10 mg/s	50					
	11/21/94	13.9	7.3	5.6	1130	341111	65 68					
	12/05/94	12.9	8.7	12.7	1054		45					
	12/15/94	8.2	7.1 6.5	12.0	1260 1775	1971, 11 1979	43 34					
	02/16/95 05/07/96	4.0 12.7	6.1	0.3	530		34					
	05/08/96	16.5	6.6	0.3	1100	14 3 11 1		0.00		•		
	05/09/96	16.0	5.9	0.8	520	- 400 a		0.07	·			···
T1L2	11/01/94	11.7	7.7		2110					15.50	1.0	2100
	11/12/94	16.3	6.7	0.0	2250	115	0.5	0.05	0.05	15.50	1.0	2100
	11/13/94	13.1	6.6 7.3	8.9 1.7	2250	115	95 99	0.25	0.25	OR		950
	11/15/94	141	7.3	1./	4190	196	99	0.25		OR		13,007
	11/16/94	14.1 13.7	7.1 7.1	1.8 4.3	4180 4220	223 260	92 89	0.05				
	11/17/94 11/18/94	15.4	7.1 7.0	3.1	4310	200	85	0.03				
	11/19/94	14.4	7.0 7.1	3.8	4230		. 05					
	11/20/94	13.5	6.9	3.2	4270		85 70 68					
	11/21/94	15.3	7.3	2.4	4320		68					
	12/05/94	13.9	7.9	9.3	3170		69					
	12/15/94	9.8	7.5	5.7	5100		45					
	02/16/95	5.5	6.9	10.0	5650		33					
	05/07/96	12,2	6.9	0.7	5370							
	05/08/96	13.8	7.0	1.9	5310			0.04				
	05/09/96	14.6	7.1	1.1	5150		`	0.18				
T1L3	11/01/94	12.1	7.7		3650							
+	11/12/94	15.6	7.4	4.5	4960	540		1.08	0.75	9.75	0.2	
	11/13/94	13.0	6.9	6.5	5060	448	1.2	4.7	0.50	7.00		3050
	11/15/94		7.2	2.7		570	0.95	0.88	1.20	9.80		3050
	11/16/94	13.6	7.1	2.2	5230	592	1.0					3150
	11/17/94		7.3	3.0	5120	657	1.6	1.2				
	11/18/94	15.5	7.0	2.8	5210	+ <u>{</u>	4.0					
	11/19/94	12.6	7.2	2.0	5200		1.8					
	11/20/94	13.8	7.0	2.3	5100		1.6					
	11/21/94	15.0	7.3	1.6	5320		3.2					
	12/05/94	14.1	7.4	• •	4090		1.6					
	12/15/94	11.6	7.5	2.8	5420		3.2 4.2					
	02/16/95	5.8	7.0	6.6	5380		4.2					
	05/07/96	11.8	7.2	2.3	5600			0.00				
	05/08/96	14.0	7.3	1.9	5550			0.03				
	05/09/96	16.8	7.1	1.4	9240			0.07				

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4-5(

Table 4.4. (continued)

Sample location	Date sampled	Temperature, °C	pН	DO, mg/L	Electrical conductivity, μmhos	Alkalinity, mg/L	Br, mg/L	Fe, mg/L	Mn, mg/L	Cl ⁻ , mg/L	NO ₃ -, mg/L	SO ₄ ² , mg/L
T1P1	11/01/94	11.4	7.2		5060							100
	11/12/94	16.0	6.8	5.4	5150	560		0	0	10.50	0.6	3250
	11/13/94	14.3	7.0	6.4	5340		1.0	0.125	0.50	7.25		3450
	11/15/94		6.8			601	0.9	0.1		10.30		3250
	11/16/94	13.9	6.8	5.3	5250	564	1.0					
	11/17/94	14.6	6.8	4.3	5140	602	2.0	0.02				
	11/18/94	14.9	6.8	4.6	5200		3.0					
	11/19/94	13.5	6.8	5.6	5260		2.2					
	11/20/94	13.4	6.9	2.3	4980		0.9					
	11/21/94	15.0	7.0	5.4	5170		1.6					
	12/05/94	14.5	7.1	4.6	4050		3.2 3.2					
	12/15/94	12.0	7.0		5300		3.2					1
	02/16/95	5.8	7.1		5060							1
	05/07/96											4
	05/08/96	16.5	7.2	5.8	4850			0.02				Y.
···	05/09/96	19.0	7.0	3.9	8470			0.11				
B1P1	5/09/96	19.2	6.9	11.8	1700			0.42				
B1L1	5/09/96	20.0	8.1	2.4	450	· · · · · · · · · · · · · · · · · ·		0.05				
B1L2	5/09/96	14.8	7.7	0.7	3550			0.06				
B1L3	5/09/96	19.2	8.2	2.5	1020			0.05				

OR = over range

Table 4.5. Hydrogen peroxide test cell (T2), soil sample results

Depth, ft		pН	Eh, mV		TOC, ppm	% S	oil moisture	Nitrate, ppm	Peroxide, mg/L
	Pre ^a	Post ^b	Post ^b	Pre ^a	Post ^b	Pre	Post ^b	Post ^b	Post ^b
1	4.6	4.7 to 5.4	200 to 440	6107	2495 to 2896	17.6	19.5 to 23.1	0 to 3.75	0
2	4.3	4.2 to 4.7	237 to 500	1576	1747 to 1525	19.5	19.4 to 21.2	0 to 1.25	0
3	4.1	4.5 to 4.6	250 to 580	1083	1437 to 1770	17.8	16.9 to 20.5	0 to 3.75	5 to 100
4	4.6	4.8 to 4.9	340 to 490	993	1048 to 1163	15.7	15.2 to 15.4	0 to 1.20	0
5	5.9	6.1 to 6.9	170 to 440	1353	958 to 1006	16.0	12.1 to 14.7	0 to 1.25	0
6	6.9	6.5 to 7.0	180 to 410	890	976 to 1329	17.2	15.6 to 17.9	0 to 1.25	0
7	7.0	6.2 to 7.3	100 to 500	431	653 to 782	17.3	18.1 to 19.6		0 to 4
8		6.7 to 7.2	100 to 350	: 	401 to 2079		15.9 to 22.0	0 to 1.25	0
9	7.8	7.2 to 7.5	140 to 298	288	427 to 545	24.6	22.1 to 26.5	0 to 2.50	0
10	7.7	5.5 to 7.5	194 to 460	436	543 to 630	25.0	23.0 to 27.6		0
11	7.8	7.1 to 7.5	205 to 420	425	608 to 771	25.8	22.1 to 27.6	0 to 2.50	0
12	7.7	7.2 to 7.4	190 to 440	640	648 to 784	21.8	20.7 to 27.0	0 to 1.25	0

^a Pre-treatment results
^b Ranges of values are from five post-treatment soil borings.
-- No sample taken

Table 4.6. Water sample results for the hydrogen peroxide test cell (T2), injection date 11/15/95

Sample location	Date sampled	Temperature, °C	pН	DO, mg/L	Electrical conductivity, μmhos	Alkalinity, mg/L	H ₂ O ₂ , mg/L	Fe, mg/L	Mn, mg/L	Cl , mg/L	NO ₃ -, mg/L	SO ₄ ² -, mg/L
T2L1	11/01/94 11/15/94	12.7	9.0		790	• • • •		0.10		22.0		
	11/15/94	14.0	8.5	8.5	1198	102		0.18	0	22.8	0	40
	11/16/94 11/17/94	13.9 16.3	8.1	6.1	1326	146	2500			16.5	1.3	550
	11/17/94		8.2	25.0	1380	400	100				1.6	
	11/18/94	16.2	7.7	25.0	792	108	30				1.8	
	11/19/94	14.5	7.3	25.0	1610	95	1				2.3	
	11/20/94		- 4	05.0	1200	114	0				2.1	
	11/21/94	15.4	7.4	25.0	1380	114	0.				2.1	
	12/05/94	13.7	8.7	25.5	1420	204	0					
	12/15/94	10.0	7.4	25.0	1750	180	0				1.2	
	02/16/95	4.5 13.9	7.0	OR	1920	194	0				1.3	
	05/07/96		6.6	11.0	1820 1920			0.00				
	05/08/96	14.0 16.5	6.7 6.5	11.0 8.6	2130			0.00				
	05/09/96			8.0				0.07				
T2L2	11/01/94	14.4	8.2	4.5	2010							
	11/14/94	14.6	7.1	4.5		265				0.0	0	2250
	11/15/94		7.3	4.4	2700	265	7000			8.0	0	2250
	11/16/94	11.5	6.9	4.4	3680	1.4	7000			6.0	12.0	2200
	11/17/94		7.6	14.0	3580	14					8.5	
	11/18/94	17.1	7.4		3550	221 375	30				4.0 2.4	
	11/19/94	14.6	7.2		3710 3880	254	30 1				1.8	
	11/20/94	16.4	7.4		3000	234	1				1.0	
	11/21/94 12/05/94	14.5	8.9		3700	614	3					
	12/03/94	11.5	7.6		4930	860	ő					
	02/16/95	4.3	7.0	OR	5460	402	ŏ				0.6	
	05/07/96	4.3	7.1	OK	J-100	702	U				0.0	
	05/08/96			13.0		•						
	05/09/96			13.0				0.05				
Γ2L3	11/01/94	12.6	7.5		4250							
1213	11/14/94	14.4	7.0	3.3	4580							
	11/15/94		7.4	2.2		685		0.1	0.25	4.0	0	2750
	11/16/94	12.1	7.0	4.0	4570	551		0		4.0	Ŏ	2600
	11/17/94	15.5	7.0	8.8	4500	576		-		**		
	11/18/94	15.5	7.4	9.3	4500	613					0	
	11/19/94	13.8	7.5	8.1	4320	625	0				0.1	
- 1	11/20/94	14.8	7.0	5.9	4520	595					0	
	11/21/94	15.2	6.6		2390	604						
	12/05/94	14.5	8.3	- 11.2	3890	732	30					
	12/15/94	12.3	7.1		4610	615	0					
	02/16/95	4.8	6.8		4840	620	0				0.1	
	05/07/96	13.4	6.9	1.0	4390							
	05/08/96	14.7	7.0	4.9	4490		•	0.15				,
	05/09/96	15.4	6.9	2.2	4540			0.85	·			

4

4-53

Table 4.6. (continued)

Sample location	Date sampled	Temperature, °C	рН	DO, mg/L	Electrical conductivity, μ mhos	Alkalinity, mg/L	H ₂ O ₂ , mg/L	Fe, mg/L	Mn, mg/L	Cl ⁻ , mg/L	NO ₃ , mg/L	SO ₄ ² , mg/L
T2P1	11/01/94	11.6	7.4	11.7	4410							
	11/14/94	14.4	6.6	11.7	4850	##O /FOF		0.1	0.05	4.6		0500
	11/15/94		6.9			570/587		0.1	0.25	4.0	0.8	3500
	11/16/94	12.7	6.8	8.0	4720	549		0		3.8	0.4	2750
	11/17/94	16.0	6.9	5.6	4700	662			•		1.2	
	11/18/94	15.6	6.8	4.4	4380	561					0.6	
	11/19/94	14.3	7.0	5.8	4530	560					0.6	
	11/20/94	14.4	7.2	7.1	4370	556					0.8	
duplicate	11/20/94	14.4	7.0	7.1	4370	· 556						
-	11/21/94	15.0	6.7	5.0	2500	604					0	
	12/05/94	14.7	7.0	7.0	3700	569	0					
	12/15/94	11.6	7.0	8.7	4071	597	0					
	02/16/95	7.3	7.4	-4	4610	560	0				0.5	
	05/07/96			4.8								
	05/08/96	14.9	7.3	5.3	4180			0.6				
	05/09/96	18.7	7.1	5.6	7410			0.13				1.4

 \overline{OR} = over range

Table 4.7. Bionutrient test cell (T3), soil sample results

Depth, ft		pН	Eh, mV	TOC, ppm		Soil sture	Bromide, ppm	INA, particles/mg
	Pre ^a	Postb	Post ^b	Pre ^a	Pre ^a	Post ^b	Post ^b	Post ^b
1	4.9	5.2 to 6.2	335 to 360	7713	17.9	19.4 to 22.3	1 to 4	2 to 29
2	4.3	4.9 to 5.6	360 to 402	1851	18.2	19.5 to 20.8	1 to 4	2 to 9
3	4.2	4.9 to 5.6	350 to 400	1261	20.9	19.3 to 21.2	1 to 3	1 to 5
4	4.4	4.7 to 5.6	300 to 430	954	17.6	15.8 to 17.5	1 to 4	2 to 5
5	5.5	5.5 to 6.5	300 to 375	947	18.0	13.6 to 19.2	0 to 3	0 to 8
6	6.5	6.5 to 6.8	250 to 295	1062	15.0	12.7 to 20.7	0 to 1	0 to 1
7	6.2	6.7 to 6.9	180 to 275	836	12.7	17.8 to 19.4	1 to 7	0 to 9
8	7.0	6.4 to 6.8	160 to 270	304	21.8	21.2 to 23.7	1 to 4	0 to 9
9	7.5	7.1 to 7.5	180 to 263	498	27.0	23.9 to 26.0	1 to 4	0 to 1
10	7.8	7.1 to 7.5	190 to 240	613	27.0	21.2 to 26.1	0 to 1	0
11	7.4	6.7 to 7.4	195 to 210	432	26.7	24.6 to 25.7	0 to 1	. 0
12	7.1	6.5 to 7.0	190 to 200	337	20.0	23.2 to 23.8	0 to 1	12

Pre-treatment results
 Ranges of values are from five post-treatment soil borings.

Table 4.8. Bionutrient test cell (T3) heterotrophic enumerations, cells/g

	A	erobic	Ana	erobic
Location/Time	4 ft depth	10 ft depth	4 ft depth	10 ft depth
T1/Pretest	>1,100,000	2400	100	10
T2/Pretest	>1,100,000	3900	10	10
T3/Pretest	>1,100,000	2400	1000	10
T3-G/3 days after injection	15,000 (2800 fungal) ^a	240	10	1
T1-J/3 days after injection	3000	ng	10	ng
T3-J/3 days after injection	46,000 (750 fungal)	240	1	ng
T1-K/30 days after injection	230	230	ng	100
T1-L/30 days after injection	230	23	10	10
T3-K/30 days after injection	23,000	23	ng	10
T3-L/30 days after injection	230	23	ng	ng

<sup>a fungal growth observed in serial dilutions in addition to bacterial growth.
b ng = no growth observed.</sup>

Note: sample results shown are two week observations, while the "ng" results are four week observations.

Source: Pfiffner 1994.

Table 4.9. Water sample results for the bionutrient test cell (T3), injection date 11/18/95

Sample location	Date sampled	Temperature, °C	pН	DO, mg/L	Electrical conductivity, μmhos	Alkalinity, mg/L	Br, mg/L	Fe, mg/L	Mn, mg/L	Cl⁻, mg/L	NO3, mg/L	SO ₄ ² -, mg/L
T3L1	11/01/94	13.2	8.5		800							
	11/19/94	14.8	7.2		1550	323	50	1.40	0.86	55.0	1.4	250
	11/20/94	12.6	7.7	3.7	1490	298	55		0.50	60.0	1.2	450
	11/21/94	14.3	7.7	4.9	1490	228	62		****	00.0	0.7	150
	12/05/94	13.4	8.5		1066	240	60				···	
	12/15/94	9.7	7.1	12.0	1290	188	46.3				•	
	02/16/95	5.1	6.6	2.1	1040	202	23.1				0.3	
	05/07/96	13.0	6.4	0.8	990						0.5	
	05/08/96			2.8				0.59				
	05/09/96	16.1	6.4	0.8	1380			OR				
T3L2	11/01/94	13.5	8.0		1650					····		
	11/19/94	14.7	7.8		1790	196	9.7	0.20	0	36.2	1.3	2375
	11/20/94	13.6	7.5	3.7	3500	194	55	0.15		54.0	1.6	2125
	11/21/94	15.2	7.6	7.0	3910	211	78				1.6	
	12/05/94	14.6	8.3		3310	218	75					•
	12/15/94	11.9	7.4	3.3	4750	270	9.9					
	02/16/95	5.8	7.0	2.2	4280	316	20.4				0.3	
	05/07/96			1.7								
	05/08/96		•		•							
	05/09/96	15.6	7.2	3.2	4430			0.37				
T3L3	11/01/94	12	7.4		3110				17.41			
	11/18/94	14.1	6.9	1.6	4190	517	0.9	0.12	8.80	3.2	0.1	2625
	11/19/94	9.3	7.0	6.4	4630	530	0.9	0.25	1.15	3.0	0.1	2375
	11/20/94	13.7	7.1	3.0	4390	518	0.8	0.28		3.0	0.1	2750
	11/21/94	15	7.1	4.2	4470	513	1			2.0	0.1	2,50
	12/05/94	14.3	7.5	5.3	3260	480	8.5	* 1			J.1	
	12/15/94	12.9	7.7	10.9	4550	520	4.7					
	02/16/95	6.6	6.9	0.6	4060	486	5.1				0.1	
	05/07/96 05/08/96	14.4	6.8	0.5	4790						V.1	
	05/09/96	16.4	6.8	3.2	5020			2.69				

Table 4.9. (continued)

Sample location	Date sampled	Temperature, °C	pН	DO, mg/L	Electrical conductivity, μmhos	Alkalinity, mg/L	Br, mg/L	Fe, mg/L	Mn, mg/L	Cl, mg/L	NO ₃ , mg/L	SO ₄ ² , mg/L
T3P1	11/18/94	14.6	6.7	5.0	3240	410	0.9	0.20	2.00	5.2	0.3	1375
	11/19/94 11/20/94	11.9	7.3	5.4	3710	395	0.8	0.33			0.1	2000
	11/21/94	15.3	,,,,		3540	388	0.9				0.1	
	12/05/94	14.7	6.9	2.5	3050	387	0.9					
	12/15/94	14.1	7.0		3960	420	0.96					
	02/16/95	7.8	7.0		4180	430	1.3				0.1	
	05/07/96	16.1	6.7	.3.0	3800							
	05/08/96	17.8	6.9	2.9	3930			2.95				

OR - over range

4-5

Table 4.10. Lime slurry test cell (T4), soil sample results

Depth, ft	1	рH	Eh, mV		Ca, ppm	TOC, ppm	% Soil	moisture
	Prea	Post ^b	Post ^b	Prea	Post ^b	- Pre ^a	Pre ^a	Post ^b
1	4.6	5.1 to 12.6	360 to 380	128	121 to 1584	6933	16.0	18.6 to 20.4
2	4.6	6.2 to 11.7	350	365	159 to 14,322	1707	17.8	17.8 to 20.1
3	4.1	4.6 to 7.0	390 to 460	322	136 to 3533	1724	21.1	20.1 to 22.4
4	4.6	4.5 to 6.0	390 to 430	378	225 to 4545	977	16.8	17.4 to 20.6
5	5.6	5.3 to 6.9	395 to 490	475	355 to 2410	889	14.6	15.0 to 20.1
6	7.0	6.6 to 11.6	370 to 390	645	464 to 3787	1187	15.3	17.3 to 21.5
7	7.6	7.3 to 9.1	280 to 350	9575	526 to 4545	420	23.8	20.2 to 22.9
8		7.1 to 11.0	305 to 325	***	3925 to 8607			23.0 to 25.6
. 9	7.6	7.3 to 11.8	320 to 340	6000	1750 to 6859	412, 430	24.0	22.3 to 27.1
10	7.8	7.4 to 12.0	295 to 305	3500	2852 to 4573	305, 412	26.9	22.9 to 26.9
11	7.0	7.3 to 7.9	305 to 330	8075	2341 to 4885	305	23.2	23.4 to 47.6
12	7.5	7.6 to 7.9	275 to 330	1838	549 to 4200	821	25.3	22.7 to 26.2

<sup>Pre-treatment results
Ranges of values are from five post-treatment soil borings.
No sample taken</sup>

4-59

Electrical SO₄2, DO, conductivity, Alkalinity, Fe, Mn, Cŀ, Sample Temperature, рH Date mg/L mg/L location °C mg/L μmhos mg/L mg/L mg/L sampled 13.2 7.5 750 T4L1 11/01/94 0.15 0.25 7.7 430 11/13/94 11.6 425 1150 23.3 11/14/94 12.3 12.3 6.5 5380 180 12.2 7.4 2100 29.8 11/15/94 175 11/16/94 13.4 12.4 5.1 5210 980 4750 1047 11/17/94 14.4 12.2 3.1 968 11/18/94 11.8 11.8 3.1 5000 100 11/19/94 12.9 4230 918 11.8 3.4 11/20/94 13.5 942 11/21/94 15.0 11.6 1.8 4650 574 12/05/94 12.1 11.7 2420 13.2 1790 280 11.7 3.5 12/15/94 10.6 02/16/95 11.0 3.1 1305 113 13.8 1980 7.7 1.4 05/07/96 14.2 2100 0.21 05/08/96 15.3 9.2 0.3 8.2 1.0 2450 0.41 05/09/96 17.5 T4L2 7.8 2350 11/01/94 14.4 7.9 20.0 1500 11/13/94 12.8 11.4 3550 1084 11/14/94 14.4 12.0 1600 0.75 17.8 1250 0.23 12.4 5.0 11/15/94 22.5 2125 1170 11/16/94 14.8 12.4 6.3 7300 1500 2.7 5360 1210 12.2 11/17/94 15.1 7160 1262 12.3 2.6 11/18/94 16.2 7170 12.1 3.1 1187 11/19/94 14.1 6980 1138 12.0 11/20/94 14.4 3580 1136 11/21/94 11.5 2.9 15.8 574 12.1 11.7 2420 12/05/94 13.2 3010 100 11.3 7.9 12/15/94 12.6 2820 9.6 3.2 33 02/16/95 4.7 6.9 2.1 4660 05/07/96 13.9 0.23 0.18 9.0 3.0 3910 05/08/96 13.5 7.4 05/09/96

Table 4.11. Water sample results for the lime slurry test cell (T4), injection date 11/13/95

4

Table 4.11. (continued)

Sample location	Date sampled	Temperature,	pН	DO, mg/L	Electrical conductivity, μ mhos	Alkalinity, mg/L	Fe, mg/L	Mn, mg/L	Cl', mg/L	SO ₄ ² , mg/L
T4L3	11/01/94	12.8	7.3	2.0	3860					
	11/13/94	14.1	7.1	3.1	4650	· 220	0.08	0.25	6.0	2600
	11/14/94	10.3	6.9	5.2	4740	570	0.13	0.25	7.0	2700
	11/15/94		7.1	1.9		626	0.05		6.8	3250
	11/16/94	14.1	7.0	1.9	4840	588		,		2900
	11/17/94	14.9	6.9	3.3	4750	627				
	11/18/94	15.5	7.1	4.1	3140	655				
	11/19/94	13.7	7.1	9.2	4620	646				
	11/20/94	14.4	7.5	2.2	2470	645				
	11/21/94	15.4	7.0	12.5	4920	741			•	
	12/05/94	14.0	7.8	4.6	2050	598				
	12/15/94	12.5	7.4	1.4	4900	685				
	02/16/95	6.6	7.2		4960	631				
	05/08/96	14.7	7.1	1.4	4740		0.01			
	05/09/96	16.3	6.8	1.5	4830		0.07			
		*								

Table 4.12. Potassium permanganate test cell (T5), soil sample results

Depth, ft	pI	I	Eh, mV	TOC, ppm		Soil sture	Manganese, ppm	
	Pre ^a	Post ^b	Post ^b	Pre	Pre ^a	Post ^b	Post ^b	
1	4.7	4.7 to 5.3	240 to 533	3483	15.7	17.4 to 20.8	9 to 344	
2	4.7	4.4 to 4.8	400 to 830	1295	19.0	19.3 to 20.8	4 to 770	
3	4.3	4.4 to 4.9	385 to 880	1103	16.9	14.8 to 21.4	3 to 89	
4	4.7	4.4 to 5.0	490 to 840	1108	12.3	14.1 to 17.4	71 to 181	
5	6.7	6.0 to 7.1	350 to 725	989	18.4	12.7 to 17.0	0 to 60	
6	7.1	6.8 to 7.0	540 to 690	405	19.8	17.1 to 19.2	0 to 14	
7	6.2	6.7 to 7.2	639 to 680	286	21.5	18.6 to 19.7	0 to 1	
8	7.8	7.4 to 7.6	515 to 688	367	24.4	21.6 to 24.2	0 to 3	
9	7.5	6.8 to 7.7	575 to 672	381	25.3	25.1 to 25.9	1 to 5	
10	7.7	6.9 to 7.6	320 to 570	396	26.3	26.1 to 27.4	1 to 7	
11	7.6	7.3 to 7.6	235 to 530	492	27.8	19.9 to 27.1	1 to 6	
12	6.9	7.4 to 8.0	252 to 460	630	22.5	22.7 to 26.9	2 to 10	

<sup>Pre-treatment results
Ranges of values are from five post-treatment soil borings.</sup>

Table 4.13. Water sample results for the potassium permanganate test cell (T5), injection date 11/16/94

Sample location	Date sampled	Temperature, °C	pН	DO, mg/L	Electrical conductivity, µmhos	Alkalinity, mg/L	Fe, mg/L	Mn, mg/L	Cl ⁻ , mg/L	NO ₃ -, mg/L	SO ₄ ²⁻ , mg/L
T5L1	11/01/94	13.1	7.6		760			- -			
	11/16/94	12.1	7.3		1560				21.8		
	11/17/94										
	11/18/94	15.0	6.4			240		4500			
	11/19/94	14.2	6.6		4270	200		3112			•
	11/20/94	14.3	6.6		2200	314		2086			
	11/21/94	15.0	6.0		4260	353		2797			
	12/05/94	13.7	7.6		2850	372		1282			
	12/15/94	11.0	6.8		2610	480		at 0		-	
	02/16/95	4.1	6.5	3.2	2210	626		§ 0			
4	05/07/96	13.6	6.2	1.7	1700						
	05/08/96	14.2	6.4	0.9	1580		0.02				
	05/09/96	15.2	6.2	0.6	1590		0.07	*		·	···
T5L2	11/01/94	15.1	8.1		1650						
	11/16/94	13.5	7.4	4.7	3280						
	11/17/94		7.0			545		17,800			
	11/18/94	15.6	7.3			250		17,700			
	11/19/94	14.9	7.2		1241	279		11,180			
	11/20/94	14.5	7.6		1027	298		8362			
	11/21/94	15.6	6.7		78 00	335		4831			
	12/05/94	14.1	7.7		4700	450		2329			
	12/15/94	12.7	7.4		4920	474		0			
	02/16/95	5.8	7.3	11.6	4980	527		10.6			
	05/07/96	12.7	6.9		4990	•					
	05/08/96	14.7	7.2	0.5	4910		0.09				
	05/09/96	14.9	7.0	1.2	5160		0.25				

4-6

Table 4.13. (continued)

Sample location	Date sampled	Temperature, °C	pН	DO, mg/L	Electrical conductivity, μ mhos	Alkalinity, mg/L	Fe, mg/L	Mn, mg/L	Cl ⁻ , mg/L	NO ₃ , mg/L	SO ₄ ² ·, mg/L
T5L3	11/01/94	12.9	7.4		3010						
	11/16/94	13.5	7.1	6.0	4320	530	0	0.6	7.8	0.1	2050
duplicate	11/16/94		7.2						6.5		
•	11/17/94		7.0			523		50.4	8.0		2750
	11/18/94	15.0	6.9			568					
	11/19/94	15.0	7.4	3.5	4420	294		1.75			
	11/20/94	14.6	7.2	2.3	4660	- 578					
	11/21/94	15.5	6.5	0.9	4660	560					
	12/05/94	14.2	7.3	6.4	3810	603		6.6			
	12/15/94	13.0	7.0	2.5	4830	588		4.89			
	02/16/95	6.7	6.8	4.3	4710	570		0			
	05/07/96	12.9	6.9	1.4	4420						
	05/08/96	15.0	7.0	4.2	4330		0.07				
e ĝi	05/09/96	17.4		1.5	4520		0.14				
T5P1	11/01/94	11.6	7.1		4370						
	11/16/94	13.7	6.7	5.7	4860	570	0.1	0	4.0	0.2	
	11/17/94		6.9			545	0		5.3		2650
	11/18/94	15.5	7.5			570					2800
	11/19/94	15.3	6.8		2500	249		0.4			
	11/20/94	14.6	7.0	6.1	4270	525					
	11/21/94	15.5	6.7	12.8	960	98		0.34			
	12/05/94	14.5	6.8	4.7	3760	562		0			
	12/15/94	12.9	7.0		5040	587		8.14			
	02/16/94	7.0	7.4		4780	565					
	05/07/96			8.3				•	•		
	05/08/96	17.0	7.1	4.6	4350		0.00				
	05/09/96	19.8	6.8	3.2	4690		0.25				

Table 4.14. Water sample results for air test cell (T6), injection date 11/19/94

Sample location	Date sampled	Temperature, °C	pН	DO, mg/L	Electrical conductivity, μ mhos	Alkalinity, mg/L	Fe, mg/L	Mn, mg/L	Cl, mg/L	NO ₃ -, mg/L	SO ₄ ² -, mg/L
T6L1	11/01/94	dry									
	11/19/94	dry									
	11/20/94	dry									
	11/21/94	dry									
	12/05/94	dry									
	12/15/94	13.0	8.3								
	02/16/95	dry 15.3									•
	05/07/96	15.3	6.3	4.7	430						
	05/08/96	15.5	6.4	1.8	430		0.13				
	05/09/96	15.8	6.1	2.6	450		0.24				
T6L2	11/01/94	dry				•					
	11/19/94	11.5	7.4		1380	275	0.12	0.4	13.5	0.2	500
	11/20/94	dry									
	11/21/94	15.5	7.2	6.1	1350						
	12/05/94	dry									
	12/15/94	13.0		4.0	2230						
	02/16/95	7.5	8.0	8.7	2190						
	05/07/96				•						
	05/08/96	14.3	7.3	2.7	1310		0.37			•	
	05/09/96	15.0	7.3	2.9	1600		0.30	-			
T6L3	11/01/94	12.9	7.8		1720						
2020	11/19/94	13.0	7.4	2.9	1300	258	0.12	0.7	5.8	0	875
	11/20/94	13.6	7.4 7.3	3.8	1170	271	0.18	1.55	5.8 3.8	0.1	875 975
	11/21/94	15.1	6.7		1070						
	12/05/94	14.2	7.3	1.7	884						
	12/15/94	13.0		4.0	2230						
	02/16/95	6.4	7.3	3.4	1938						
	05/07/96	14.4	7.0	2.8	520						
	05/08/96	17.1	7.3	4.5	460		0.01				
	05/09/96	17.6	7.1	3.4	520		0.04				

Table 4.15. Iron micropowder test cell (T7), soil sample results

Depth, ft		pН	Eh, mV	TOC, ppm	Iron, ppm
	Prea	Post ^b	Post ^b	Pre ^a	Post ^b
1	6.5	5.4 to 5.5	330 to 380	6893	3.0 to 3.5
2	4.6	5.7 to 6.4	380 to 390	1903	13.1 to OR
3	4.7	5.2 to 5.4	410 to 420	1110	0.9 to 29.5
4	4.9	5.5 to 5.6	370 to 380	1277	0.5 to 1.2
5	6.4	5.5 to 6.3	270 to 280	866	2.3 to 32.8
6	6.7	6.4 to 6.4	210 to 240	1631	1.0 to 1.0
7	6.5	6.4 to 6.5	200 to 230	531	2.4 to 3.5
8	7.4	6.4 to 6.6	150 to 200	385	0.9 to 1.2
9	7.0	6.8 to 7.1	150 to 180	383	0.2 to 1.2
10	6.9	6.5 to 6.7	135 to 165	394	1.2 to 1.7
-11	6.9	6.6 to 6.8	140 to 165	033	1.3 to 1.8
12	7.7	6.1	170	393	1.4 to 3.2

^a Pre-treatment results ^b Ranges of values are from five post-treatment soil borings. OR = over range

Table 4.16. Water sample results for iron micropowder test cell (T7), injection date 11/19/94

Sample location	Date sampled	Temperature,	pН	DO, mg/L	Electrical conductivity, μ mhos	Alkalinity, mg/L	Br ⁻ , mg/L	Fe, mg/L	Mn, mg/L	Cl', mg/L	NO ₃ , mg/L	SO ₄ ²⁻ , mg/L
T7L1	11/01/94	12.7	8.8		460							
•	11/19/94	9.2	7.6		860	200		0.1	0.4	34.0	1.1	150
	11/20/94	12.0	7.9		790	163	0.9	0.7		3.7	1.1	188
	11/21/94	15.4	7.2	6.5	590			0.1				188
	12/05/94											
	12/15/94	11.6	6.6	10.4	795			OR	2.0			
	02/16/95	4.0	6.2	11.0	833			25.5	2.7			
	05/07/96	14.7	6.1	1.8	430							
	05/08/96	15.8	5.9	1.2	280			2.98				
	05/09/96_	15.9	5.9	1.0	490			OR				
T7L2	11/01/94	14.7	9.0		1030							
	11/19/94	9.1	8.0		820	188		3.1	0.4	10.5	0.6	330
	11/20/94	13.0	8.0	5.2	1000	173	0.7	0.3	0.2	11.2	0.2	900
	11/21/94	15.8	7.4	3.3	1610			0.1	0.2			700
	12/05/94	14.8	7.3	11.5	1381			3.2	0.5			850
	12/15/94	12.7	6.9	3.0	2580			OR				
	02/16/95	4.8	7.3	5.6	1913			5.8				
	05/07/96	13.7	7.2	0.9	2140			•				
	05/08/96	13.7	7.2	1.2	2120			OR				
	05/09/96	16.4	7.2	2.4	2000			OR				

Table 4.16. (continued)

Sample location	Date sampled	Temperature, °C	pН	DO, mg/L	Electrical conductivity, μ mhos	Alkalinity, mg/L	Br ⁻ , mg/L	Fe, mg/L	Mn, mg/L	Cl [*] , mg/L	NO ₃ ·, mg/L	SO ₄ ² , mg/L
T7L3	11/01/94	13.0	7.6		2780							•
	11/19/94	13.0	7.3	1.6	3520	380		0.1	1.6	4.0	0.1	900
	11/20/94	13.3	7.3	3.1	2090	451	0.7	0.2	2.2	3.2	0.6	2432
	11/21/94	15.1	6.8	9.4	3620			0.1	2.0			2375
	12/05/94	14.8	8.5					0.4	1.5			
	12/15/94	13.2	7.2	4.9	4390			0.8	3.3			
•	02/16/95	5.8	7.0	1.7	4350			1.8	1.3			
	05/07/96	14.1	6.9	1.0	3840							
	05/08/96	14.8	7.1	3.5	3740	•		0.06				
	05/09/96	17.1	6.9	2.3	3920			0.16				

OR = over range Blanks indicate insufficient sample volume for analyses



5. Equipment Operations Observations

One of the objectives of this field test was to determine the operational characteristics of the MPIS process. This objective was accomplished by observation and documentation during the field tests. Observations included injection volumes, flow rates, back pressure, general equipment operations, and operational and maintenance problems.

The average volume of fluid injected into the soil at the CTS was about 0.27 gal/ft³, calculated in the following way:

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7,030 ft<sup>3</sup> = cell volume (effective treated cell size of 26 ft x 26 ft x 10.4 ft),
1900 gal = average volume injected (2300 gal per test cell minus 400 gal lost to surface seepage)
0.27 gal/ft<sup>3</sup> = 1900 gal/7,098 ft<sup>3</sup>.
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The approximate volume of the air-filled pores within each test cell prior to injection was estimated at about 2,000 gal or 270 ft³. This was in part, the basis for the target injection volume. If the injected fluid, which equaled approximately 1900 gal, entered the air filled pores, then the soil within each test cell should have been 100% saturated after injection. Post-treatment soil sample data indicate that the soil was not uniformly 100% saturated; thus, some of the injected fluid must have moved outside of the test cell boundaries along fractures and other preferred flow paths.

Although the average injection pressure was about 100 psi, it fluctuated between 60 and 200 psi depending on the resistance of the soil to injection. This injection pressure range is similar to that used during a previous conventional soil fracturing test conducted at the CTS and may have had similar effects (i.e., creating fractures in the fine-grained media). During fluid injection, the flow rates varied from 15 to 40 gal/min, again depending on the resistance of the soil to injection. Activities performed, production rates, and total time for each major activity are shown on Table 5.1.

Based on the work conducted, it appears that the system is capable of delivering approximately 2 gal of reagent per injector per 15-in. interval to a depth of 10.4 ft in about 5 to 6 min per injection location. The cost per day for subcontractor equipment and labor was approximately \$1300 (1994 cost). The equipment is capable of delivering reagent to between 10,500 and 16,000 ft³ of soil in an 8-h work day. Thus, the cost per cubic foot of soil at the CTS was between \$0.12/ft³ and \$0.08/ft³ (\$3.24 to \$2.16/cubic yard). The cost for reagents varied considerably, depending on the particular chemical, the purity, and the desired concentration of the injected solution. Extrapolation of these costs to a contaminated site would require a multiplier to account for additional operating requirements such as waste management, a higher level of worker protection, and other issues.

During the injections, several operational requirements were noted. The machine operates by performing an injection and then backing up 2 ft to start the next injection. Because the solution delivery hose connects to the back of the machine, the hose must be moved manually so the machine does not run over it. Much time can be spent in mixing reagent solutions and the injection progresses much more rapidly if enough solution is mixed to inject the entire area to be treated prior to starting the injection. This could require mixing almost 5,000 gal of solution for a full day of injection activities. The individual injector ports can clog with soil very easily when stop-and-go injection is performed. It is prudent to check the injector ports aboveground occasionally to see if all injectors are open. If there are clogged ports, they can be easily cleared with a piece of wire. If work is being conducted in a contaminated area, some means of containing any seepage to the ground surface should be in place prior to the start of injection to prevent spreading contamination in runoff from the injected site. This could be accomplished with a shallow trench, filter booms, and/or silt fencing. Work can be conducted in relatively restricted surroundings as the equipment is highly maneuverable and is capable of angled injections.

Generally speaking, the MPIS provides a rapid means of injecting reagent into the subsurface to a depth of 10.4 ft. There are other units available capable of injecting to 30 or 40 ft, but these are not as maneuverable and their cost is considerably more than the system tested for this demonstration. The system tested is largely nondisruptive to subsurface supports such as footers or pipe beds, and injections can be performed close to buildings and other cultural features. The system is also capable of being using an angles up to 45° from vertical.

Table 5.1. Task summary for MPIS testing

Activity	Production rate	Total time		
Pre-treatment	Pre-treatment borings,	5 d/32 borings		
characterization	6 to 7/d			
	Lysimeters, 12/d	2 d/24 lysimeters		
	Piezometers, 5/d	1 d/5 piezometers		
	SMT probes, 24/d	1 d/24 SMT probes		
	Water samples, 30/d	1 d/30 samples		
Mobilization and	Mobilization, 2 d	4 d		
shakedown	Shakedown, 2 d			
Reagent mixing	0.5 to 1 h/1000 gal batch	10 h/13,000 gal		
Test cell 1 injection	5.3 min/setup	190 min for 36 setups		
Test cell 2 injection	5.7 min/setup	205 min for 36 setups		
Test cell 3 injection	5.7 min/setup	205 min for 36 setups		
Test cell 4 injection	4.3 min/setup	155 min for 36 setups		
Test cell 5 injection	5.4 min/setup	195 min for 36 setups		
Test cell 6 injection	5 min/setup	20 min for four setups		
Test cell 7 injection	5.5 min/setup	22 min for four setups		
Post-treatment	Soil borings, 5 to 6/d	7 d/37 borings		
characterization	Water samples, 12 to 30/d	11 d/200 samples		

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6. Summary and Conclusions

The purpose of the permeation and dispersal of reactive fluids project at the PORTS CTS was to evaluate:

- the feasibility of using the MPIS for injection of treatment agents into the subsurface, and
- the relative effectiveness of injecting contrasting treatment agents for in situ remediation of low permeability soils.

The stated objectives of this project were to:

- characterize pore fracture size and continuity in the untreated soil and then determine changes in fracture size and continuity as affected by reagent or air injection,
- determine matrix effects of the various fluids released with respect to changes in soil pH, TOC, conductivity, etc.,
- determine dispersal of reactive particles in LPM,
- determine dispersal of oxidants in LPM, and
- determine the operation and maintenance characteristics of the injection tool.

Performance testing of the MPIS at the PORTS CTS has demonstrated that it is feasible to use the MPIS for injection of various reagents into relatively shallow depths of low permeability soil. However, observations suggest that the treatment agents injected advect along pre-existing preferential flow paths and only in cases of persistent agents, do they diffuse into matrix blocks. Observations of the various injections also indicate that the initial effect generally occurred in the upper more structured zone of the subsurface at the test site. Percolation of the injected media from the saturated upper soils over time resulted in subsequent, and less dramatic, changes to the deeper portions of the cells.

The relative success of injecting the various treatments appears to be similar. Post-treatment soil sampling results and boring logs indicate that the distribution patterns for the treatment agents were comparable. Thus, the slurry mixtures (i.e., lime, iron micropowder/guar gum) flowed through the injector system and penetrated the subsurface equally as well as other media (e.g., peroxide solution). From an operational standpoint, no problems occurred with the MPIS during injection of any of the treatments; however, the tank containing the iron micropowder required continuous stirring to prevent settling of the metal particles.

Pre-treatment and post-treatment soil boring logs were prepared for each test cell to assess macroscopic changes in pore fracture size and continuity. No significant changes were observed, excluding the filling of the existing fractures and macropores with the injected

media. Sample cores from each test cell were also collected for x-ray and SEM analysis to assess pre- and post-treatment fracture morphology. X-ray data were inconclusive, and SEM analyses were not completed.

Varying matrix effects were observed for the various treatment injections. No notable effects were observed for test cell 1 (water with tracers) and test cell 6 (air). The most significant effect observed in test cell 2 (hydrogen peroxide) was a significant decrease in TOC at a depth of up to 2 ft bgs; however, little change was observed at greater depths. The peroxide injection also effected a significant increase in DO in the lysimeter (water) samples. Results from test cell 2 also indicate that the peroxide reacts rapidly in the fine-grained soil and degrades in a relatively short time (a few days). No significant matrix effects were observed in test cell 3 (bionutrients with tracers) soils. However, increased nitrate levels were measured in lysimeter samples as a result of the nutrient ingredients.

Significant matrix effects were observed in test cell 4 (lime slurry). The lime injection resulted in dramatic pH increases in both soil and lysimeter samples. Alkalinity and conductivity measurements also increased substantially in the water samples. These effects appear to be fairly long lasting (at least 4 weeks). Post-treatment results from test cell 5 (potassium permanganate) indicated dramatic increases in soil Eh throughout the soil profile and a slight decrease in pH in the water samples. Post-treatment manganese concentrations in lysimeter samples remain elevated for approximately 5 weeks. Thus, of the oxidants injected, the potassium permanganate appears to react mildly but will persist longer than the peroxide. No significant matrix effects were observed for test cell 7 (iron micropowder) soils, excluding an increase in the iron concentration. However, a decrease in pH was observed in the water samples in conjunction with an increase in the soluble iron concentration.

The dispersal of the various treatment agents in low permeability soils was generally related to the success of the injection. Thus, there was a similar pattern of initial dispersal in each test cell as discussed previously. However, additional dispersal in the subsurface over time was observed for those treatments (i.e., lime, potassium permanganate, and iron) that were soluble and more persistent. This appeared to occur as affected water percolated downward through the soil.

Based on the results of the testing at the PORTS CTS, the MPIS shows promise for treatment of contaminants in shallow, low permeability soils. The injected treatment agents follow pre-existing preferential flow paths in low permeability soils (e.g., fractures) but do not appear to create new fractures. The system could be used to deliver a treatment medium into the preferential flow paths, thereby preventing further migration of contaminants, at least until the agent was exhausted or flushed out by surface recharge. Additional treatment could occur by subsequent percolation of affected water through the contaminated soil layer. The MPIS would probably be very effective at delivering solutions into unconsolidated sandy sediments or loose tailings.

Based on their inherent reactive characteristics, three of the treatments tested at the CTS have potential application for in situ remediation of VOCs in low permeability soils. These include hydrogen peroxide, potassium permanganate, and iron micropowder. Further testing of these treatments in conjunction with the MPIS is recommended for a contaminated site. The other treatments tested at the CTS are generally not effective for chlorinated solvents; however, they could be delivered with the MPIS for in situ treatment at other contaminated sites.

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7. References

American Petroleum Institute. 1995. Petroleum Contaminated Low Permeability Soil: Hydrocarbon Distribution Processes, Exposure Pathways and In Situ Remediation Technologies. Health and Environmental Sciences Dept. Publication No. 4631, September.

Case, T. 1997. Reactive permanganate grouts for horizontal permeable barriers and in situ treatment of ground water. M.S. thesis, Environmental Science & Engineering Division, Colorado School of Mines, Golden, Colorado.

Davenport, D.T., D.J. Georgopoulos, R.L. Siegrist, M.I. Morris, and O.R. West. 1994. Technology Demonstration, Assessment, and Application for a RCRA Closure: Observations and Lessons Learned in the Process. Proc. Spectrum '94, Atlanta, August, 1994.

Gates, D.D. and R.L. Siegrist. 1995. In Situ Chemical Oxidation of Trichloroethylene Using Hydrogen Peroxide. ASCE Journal of Environmental Engineering, 121(9):639-644.

Gierke, J.S., C. Wang, O.R. West, and R.L. Siegrist. 1995. "In Situ Mixed Region Vapor Stripping of Low Permeability Media: 3. Modeling of Field Tests". *Environ. Science & Technology*, **29**(9):2208-2216.

Huling, S.G. and J.W. Weaver. 1991. Dense Nonaqueous Phase Liquids. Ground Water Issue Paper, EPA/540/4-91-002. Office of Research and Development, Washington, D.C. 20460.

Johnson, R., D. Grady, T. Walden, and H. Hopkins. 1995. Remediation of Gasoline Contaminated Low Permeability Media as Evaluated at a Controlled Release Test Site. Invited presentation at the Second International Conference on Challenges and Innovations in the Management of Hazardous Wastes. Air and Waste Management Conference, May, 1995, Washington, D.C.

Johnson, R.L., and D. Grady. 1995. Remediation of Tight Soils Using Air Flushing: Summary of 1994 Field Experiments at Sarnia Ontario. Prepared for the American Petroleum Institute by Oregon Graduate Institute Center for Groundwater Research, Beaverton, Oregon.

MacDonald, J. A. and M. Kavanaugh. 1994. Restoring Contaminated Groundwater: An Achievable Goal? 1994. Environmental Science & Technology, 28(8):362A-368A.

Murdoch, L., B. Slack, B. Siegrist, S. Vesper, and T. Meiggs. 1997a. Hydraulic Fracturing Advances. *Civil Engineering*, May, pp. 10A-12A.

- Murdoch, L., W. Slack, R. Siegrist, S. Vesper, and T. Meiggs. 1997b. Advanced Hydraulic Fracturing Methods to Create In Situ Reactive Barriers. Proc. International Containment Technology Conference and Exhibition. February 9-12, 1997, St. Petersburg, FL. In press.
- ORNL. 1994. Final Work Plan for Multi-port Injector System Testing in Low-Permeability Soils at the Clean Test Site. Oak Ridge National Laboratory, Grand Junction, Colorado.
- ORNL. 1993. Environmental Technology Section (formerly Pollutant Assessments Group) Procedures Manual. ORNL-6645/R2. Oak Ridge National Laboratory, Grand Junction, Colorado.
- Pfiffner, S.M., R.L. Siegrist, K.S. Lowe, D.B. Ringleberg, and A.V. Palumbo. 1997. Sarnia: Bioremediation Processes Demonstrated at a Controlled Release Site. Symposium on In Situ and On-Site Bioremediation. April 1997. New Orleans.
- Pfiffner, S. 1994. Written communication with R. L. Siegrist, ORNL from Susan Pfiffner, University of Tennessee Center for Environmental Biotechnology, Microbial Enumerations and Activities.
- Siegrist, R. L., K. S. Lowe, L. C. Murdoch, D. A. Pickering, and T. L. Case. 1999. In Situ Oxidation by Fracture Emplaced Reactive Solids. *J. Environmental Engineering*. 125(5):429-440.
- Siegrist, R. L., K. S. Lowe, L. C. Murdoch, W. W. Slack, and T. C. Houk. 1998. X-231A Demonstration of In Situ Remediation of DNAPL Compounds in Low Permeability Media by Soil Fracturing with Thermally Enhanced Mass Recovery or Reactive Barrier Destruction. ORNL/TM-13534. Oak Ridge National Laboratory, Oak Ridge, Tennessee.
- Siegrist, R.L., N.E. Korte, D. Smuin, O.R. West, D.D. Gates, and J.S. Gunderson. 1996. In Situ Treatment of Contaminants in Low Permeability Soils: Biogeochemical Enhancement by Subsurface Manipulation. Invited presentation at First International Conference on Contaminants in the Soil Environment in the Australasia-Pacific Region. February 1996, Adelaide, South Australia.
- Siegrist, R.L. and K.S. Lowe (ed.). 1995. In Situ Remediation of DNAPL Compounds in Low Permeability Media Interim Report. Interim project report prepared by Oak Ridge National Laboratory with contributions from an ad hoc team of scientists and engineers from DOE national laboratories, academia, and industry.

- Siegrist, R.L., O.R. West, M. I. Morris, D. A. Pickering, D. W. Greene, C. A. Muhr, D. D. Davenport and J.S. Gierke. 1995a. In Situ Mixed Region Vapor Stripping of Low Permeability Media. 2. Full Scale Field Experiments. *Environ. Science & Technology*. 29(9):2198-2207.
- Siegrist, R.L., N. E. Korte, M.T. Muck, D.R. Smuin, A.D. Laase, O.R. West, D.T. Davenport, and J. Walker. 1995b. Field Evaluation of Subsurface Manipulation by Fracturing, Multipoint Injection/Dispersal, and Directional Drilling Using Unconfined Test Cells. Invited presentation at the Second International Conference on Challenges and Innovations in the Management of Hazardous Wastes. Air and Waste Management Conference, May, 1995, Washington, D.C.
- Siegrist R.L., N.E. Korte, M.T. Muck, D.R. Smuin, A.D. Laase, O.R. West, D.T. Davenport, and J. Walker. 1995c. Field Evaluation of Subsurface Manipulation by Fracturing, Permeation Dispersal, and Horizontal Well Recirculation Using Unconfined Test Cells. Invited presentation at the National Ground Water Association Annual Educational Conference, October, 1995, Indianapolis, IN.
- Siegrist, R. L., M. I. Morris, O. R. West, D. A. Pickering, C. A. Muhr, D. W. Greene, D. D. Gates, T. J. Mitchell, R. A. Jenkins, and T. McKnight. 1994a. X-231B Technology Demonstration for In Situ Treatment of Contaminated Soil: Field Evaluation of Mixed Region Vapor Stripping, Chemical Oxidation, and Solidification Processes. ORNL/TM-12261. Oak Ridge National Laboratory, Oak Ridge, Tennessee.
- Siegrist, R. L., M. I. Morris, O. R. West, D. D. Gates, D. A. Pickering, R. A. Jenkins, T. J. Mitchell, D. W. Greene, C. A. Muhr, S. E. Herbes, T. M. Gilliam, H. L. Jennings, A. J. Lucero, J. Zutman, T. McKnight, and J. S. Watson. 1995. *The X-231B Technology Demonstration for In Situ Treatment of Contaminated Soil: Field Demonstration and Testing of Vapor Stripping, Peroxidation, and Solidification*. ORNL/TM-12261. Oak Ridge National Laboratory, Oak Ridge, Tennessee.
- Siegrist, R.L., O. R. West, D. D. Gates, S. E. Herbes, M. I. Morris, T. M. Gilliam, H. L. Jennings, A. J. Lucero, J. S. Watson, D. A. Pickering, D. W. Greene, C. A. Muhr, J. Zutman, R. A. Jenkins, T. J. Mitchell, T. McKnight, D. T. Davenport. 1993. In: Post, R. G. Full-scale Field Demonstration and Testing of Physicochemical Processes for In Situ Treatment of Contaminated Soil. Proc. Waste Management '93. Feb. 28 Mar. 4 1993. Tucson, AZ. American Nuclear Society, pp. 849 854.

Smuin, D.R., R.L. Siegrist, N.E. Korte, O.R. West, J. Strong-Gunderson, and P.M. Kearl. 1995. Bromide Tracer Evaluation of a Multipoint Injection System for Soil Treatment. Presentation at American Geophysical Union, Fall, 1995 meeting.

Strong-Gunderson. J. M., and A. V. Palumbo. 1995. Laboratory Studies Demonstrate an Innovative Colloid Tracer: Real Time, Sensitive, and Non-Persistent.

Urynowicz, M. A. and R. L. Siegrist. 2000. Permeation Dispersal of Treatment Agents for In Situ Remediation in Low Permeability Media: 2. Laboratory Studies with Intact Cores. ORNL/TM-13597. Oak Ridge National Laboratory, Oak Ridge, Tennessee.

U.S. DOE. 1996. In Situ Remediation of DNAPL Compounds in Low Permeability Media Fate/Transport, In Situ Constol Technologies, and Risk Reduction. ORNL/TM-13305. Oak Ridge National Laboratory, Oak Ridge, Tennessee.

U.S. EPA. 1992. Dense Nonaqueous Phase Liquids -- A workshop Summary. EPA/600/R-92/030. Office of Research and Development, Washington, D.C. 20460.

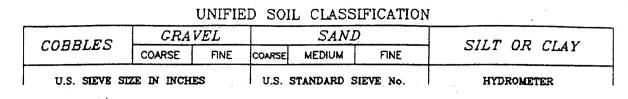
Walden, T. 1993. Remediation of Low Permeability Soils: An API Research Program. Proc. 1993 Petroleum Hydrocarbons and Organic Chemicals in Groundwater: Prevention, Detection, and Restoration. Houston, TX. *National Water Well Assn.* pp. 623-629.

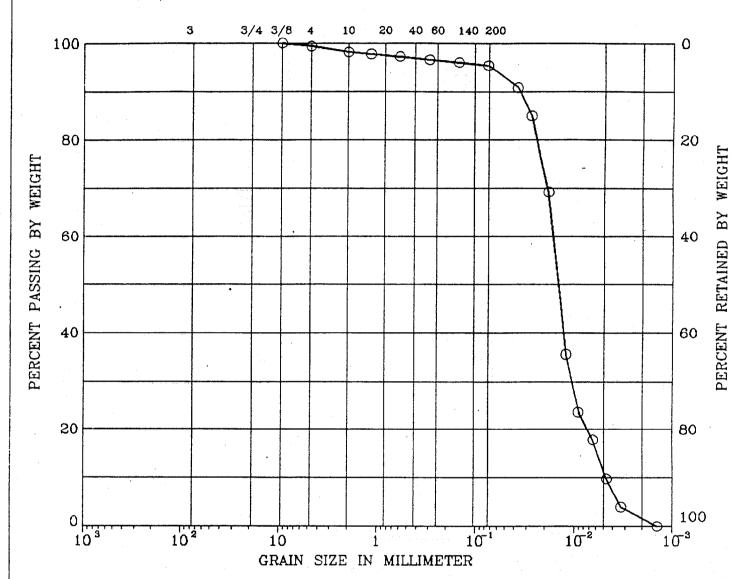
West, Or. R., R. L. Siegrist, J. S. Gierke, S. W. Schmunk, A. J. Lucero, and H. L. Jennings. 1995. In Situ Mixed Region Vapor Stripping of Low Permeability Media. 1. Process Features and Laboratory Experiments. Environmental Science & Technology. 29(9):2191 - 2197.

APPENDIX A

GEOTECHNICAL, BACKGROUND, AND GRAIN SIZE DISTRIBUTION RESULTS FOR THE CTS

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SYMBOL	BORING	DEPTH (ft)	(%)	PI (%)	DESCRIPTION	
0	BH1-05	5.0	49	31	CLAY, silty, It yelsh br, mott gr, w/ rts USC=CL	

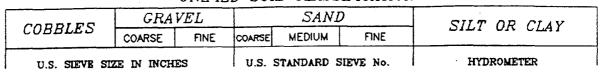
Remark: Martin Marietta Energy Systems, Inc.

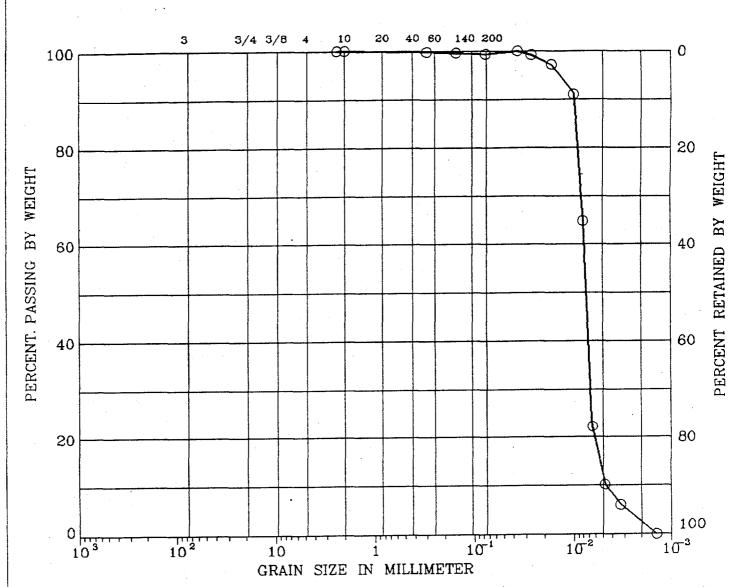
Project No.0-4267 Portsmouth - OTD Mini Characterization

ENVIRONMENTAL

GRAIN SIZE DISTRIBUTION April 5, 1994

UNIFIED SOIL CLASSIFICATION





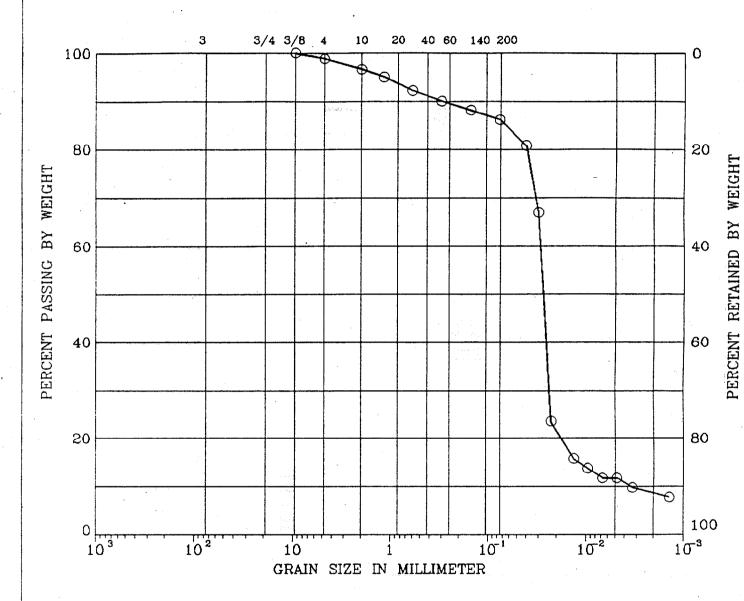
•		перти П.	ΡĪ			
SYMBOL	BORING	DEPTH LL (ft) (%)	<u>(%)</u>	DESCRIPTION		
0	BH1-13.5	13-13.5 64	38	CLAY, silty, rdsh br, mott blk	USC=CH	•

Remark: Martin Marietta Energy Systems, Inc.

Project No.0-4267	Portsmouth - OTD Mini Characterization	
OGDEN	GRAIN SIZE DISTRIBUTION April 5, 1994	

UNIFIED SOIL CLASSIFICATION

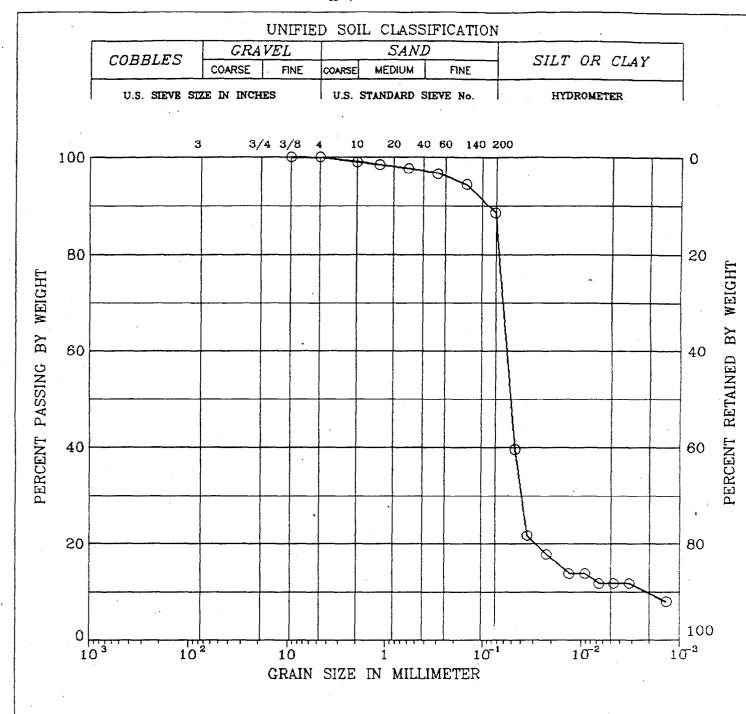
COBBLES	GRA	VEL		SANI)	SILT OR CLAY
COBBLES	COARSE	FINE	COARSE	MEDIUM	FINE	SILI OR CLAY
U.S. SIEVE SIZE IN INCHES		U.S. S	STANDARD S	IEVE No.	HYDROMETER	



SYMBOL	BORING	DEPTH LL (ft) (%)	PI (%)_	DESCRIPTION
0	BH2-05	4.5-5.0 33	14	CLAY, sity, it yish br mott gr & br w/rk fragsUSC=CL

Remark: Martin Marietta Energy Systems, Inc.

Project No.0-4267	Portsmouth - OTD Mini Characterization
OGDEN ENVIRONMENTAL	GRAIN SIZE DISTRIBUTION April 5, 1994



SYMBOL	BORING	DEPTH LL (x)	PI (%)	DESCRIPTION	
0	BH2-14	13.5-14 26	12	CLAY, silty, yellowish brown	USC=CL

Remark: Martin Marietta Energy Systems, Inc.

Project No.0-4267	Portsmouth - OTD Mini Characterization	on
OGDEN	GRAIN SIZE DISTRIBUTION	April 5, 1994

SUMMARY OF LABORATORY TEST RESULTS

					UN WEIG (PC	GHT	SPECIFIC GRAVITY		RBERG MITS		·	Project: Portsmouth ODT Mini Characterization Project Number: 0-4267-0088 Date: May 20, 1994
Hole No.	Sample No.	Sample Type*	Depth (ft)	Natural Moisture (%)	Wet	Dry	G,	Liquid Limit (%)	Plasticity Index (%)	Unified Soil Classification	Other Test	Soil Description
BH-1-05		SS	5.0	23.4			2.66	49	31	CL		CLAY, silty, light yellowish brown mottled gray with roots
BH-1-13.5		SS	13.0 - 13.5	32.4			2.63	64	38	СН		CLAY, silty, reddish-brown mottled black
BH-1-22		SS	21.5 - 22.0	21.2			2.67	27	15	CL		CLAY, silty, light yellowish brown mottled brown with weathered shale fragments
BH-2-05		SS	4.5 - 5.0	22.6			2.60	33	14	CL		CLAY, silty, light yellowish brown mottled gray and brown with rock fragments
BH-2-14	·	SS	13.5 - 14.0	22.6			2.69	26	12	CL		CLAY, silty, slightly sandy, tan
BH-2-28		SS	27.5 - 28.0	19.1			2.75	31	13	CL		CLAY, silty, dark yellowish brown with rock fragments
BH-3-09		SS	8.5 - 9.0	24.5			2.66	50	28	CL		CLAY, silty, tan mottled light gray
BH-3-12		ss	11.5 - 12.0	35.5			2.73	72	49	СН		CLAY, silty, light brown mottled gray
BH-3-19		SS	18.5 - 19.0	17.7			2. 6 8	23	11	CL		CLAY, silty, dark greenish brown with rock fragments
BH-4-07		ss	6.5 - 7.0	26.7			2.69	60	35	СН		CLAY, silty, light brown mottled gray and yellowish-brown
BH-4-15		SS	14.5 - 15.0	31.0			2.69	69	43	СН		CLAY, silty, light brown mottled gray

^{*}ST-SHELBY TUBE SAMPLE, SS-SPLIT SPOON SAMPLE, B-BAG SAMPLE

C-CONSOLIDATION
S-SIEVE OR GRAIN SIZE ANALYSIS
U-UNCONFINED COMPRESSION TEST

P-PROCTOR TEST D-DIRECT SHEAR TEST T-TRIAXIAL TEST DATA CHECKED BY

GA Technical Services

A-5

^{**}TEST RESULTS REPORTED ON OTHER SHEETS:

SUMMARY OF LABORATORY TEST RESULTS

<u></u>								· · · · · · · · · · · · · · · · · · ·				
					UN WEI (PC	GHT	SPECIFIC GRAVITY		RBERG MITS			Project: Portsmouth ODT Mini Characterization Project Number: 0-4267-0088
				·								Date: May 20, 1994
Hole No.	Sample No.	Sample Type*	Depth (ft)	Natural Moisture (%)	Wet	Dry	G,	Liquid Limit (%)	Plasticity Index (%)	Unified Soil Classification	Other Test	Soll Description
BH-4-21	·	ss	20.5 - 21.0	36.7			2.72	· 48	26	CL		CLAY, silty, brown mottled gray and reddish- brown
BH-5-08		ss	7.5 - 8.0	32.3			2.69	69	44	СН		CLAY, silty, light brown mottled gray
BH-5-13		SS	12.5 - 13.0	39.9			2.70	46	25	ĊL		CLAY, sitty, light brown mottled gray yellowish- brown and reddish-brown
BH-5-19		SS	18.5 - 19.0	20.7			2.66	26	-11	CL		CLAY, silty, yellowish-brown mottled light gray with rock fragments
BH-5-23		SS	22.5 - 23.0	30.9	·		2.70	28	€ 15	CL		CLAY, silty slightly sandy light yellowish-brown
BH-6-02	-	SS	1.5 - 2.0	25,9			2.62	41	25	CL		CLAY, silty, light yellowish-brown
BH-6-18		SS	17.5 - 18.0	21.7			2.65	26	13	CL		CLAY, silty, light yellowish-brown
BH-6-29		ss	28.5 - 29.0	14.3			2.69		-			CLAY, slity, dark brown with rock fragments
BH-7-12		ss	11.5 - 12.0	20.7			2.70	28	14	CL		CLAY, sitty, slightly sandy, light yellowish-brown
BH-7-20		ss	19.5 - 20.0	19.3			2,65	30	14	CL		CLAY, silty, slightly sandy, yellowish-brown
BH-7-26		ss	25.5 - 26.0	17.3			2.65	29	- 12	CL		CLAY, silty, dar brown

^{*}ST-SHELBY TUBE SAMPLE, SS-SPLIT SPOON SAMPLE, B-BAG SAMPLE

C-CONSOLIDATION
S-SIEVE OR GRAIN SIZE ANALYSIS
U-UNCONFINED COMPRESSION TEST

P-PROCTOR TEST D-DIRECT SHEAR TEST T-TRIAXIAL TEST DATA CHECKED BY

GA Technical Services

A-6

^{**}TEST RESULTS REPORTED ON OTHER SHEETS:

SUMMARY OF LABORA FORY TEST RESULTS

						IIT GHT ÇF)	SPECIFIC GRAVITY		RBERG MITS			Project: Portsmouth ODT Mini Characterization Project Number: 0-4267-0088 Date: May 20, 1994
Hole No.	Sample No.	Sample Type*	Depth (ft)	Natural Moisture (%)	Wet	Dry	G,	Liquid Limit (%)	Plasticity Index (%)	Unified Soil Classification	Other Test	Soil Description
BH-8-13		SS	12.5 - 13.0	33.5			2.68	61	41	СН		CLAY, silty, reddish-brown
BH-8-22		SS	21.5 - 22.0	28.5			2.72	26	12	CL		CLAY, silty, dark brown mottled dark gray with rock fragments
BH-9-09		SS	8.5 - 9.0	33.6			2.70	71	48	СН		CLAY, silty, light reddish-brown mottled gray and yellowish-brown
BH-9-21		SS	20.9 - 21.0	36.4			2.73	53	28	СН		CLAY, silty, gray mottled reddish-brown
BH-9-21.5		SS	21.0 - 21.5	29.9			2.75	51	26	СН		CLAY, silty, gray mottled greenish-brown
				·								
		·	,									
				,								

*ST-SHELBY TUBE SAMPLE, SS-SPLIT SPOON SAMPLE, B-BAG SAMPLE

C-CONSOLIDATION
S-SIEVE OR GRAIN SIZE ANALYSIS
U-UNCONFINED COMPRESSION TEST

P-PROCTOR TEST D-DIRECT SHEAR TEST T-TRIAXIAL TEST



GA Technical Services

^{**}TEST RESULTS REPORTED ON OTHER SHEETS:

MPIS Baseline Soil Characteristics: Moisture Content, Bulk Density, Porosity, pH, TOC

Pan ID	Sample Name D	epth	% Moisture based on dry wt	% Moisture based on wet wt	Wt bulk Density, g/cc	Dry bulk Density, g/cc	Dry wt % moisture	% volume porosity	% volume water-filled porosity	% volume air-filled porosity	pН	TOC, ppm	Avg. % soil moisture (wet wt)	Avg. pH	Avg. bulk density (moist), g/cc	ppm
 	T1-BE-00-01	1	21.95	18							4.26	6253		4.89		6396.63
14	T2-BE-00-01	1	21.4			median mois	t bulk				4.61	6107		•		
14	12-00-01	•				density										
27	T3-BE-00-01	1	21.82	17.91		1.83					4.87	7713				
40	T4-BE-00-01	1	19.03	15.99							4.62	6933				
53	T5-BE-00-01	1	18.66	15.72							4.67	3483				
66	T6-BE-00-01	1	21.76	17.87							4.93			<u> </u>		
79	T7-BA-00-01	1	18.21								6.5			ļ		
92	S1-BA-00-01	1	18.09						ļ.,	<u> </u>	4.68			1 40		1629.50
2	TI-BE-01-02	2	22.52								4.42			4.49		1629.30
15	T2-BE-01-02	2	24.4					<u> </u>	<u> </u>		4.3					
28	T3-BE-01-02	2	22.20								4.34					
41	T4-BE-01-02	2	21.75						<u> </u>		4.57			 		
54	T5-BE-01-02	2	23.5					ļ	ļ	 	4.74			 		
67	T6-BE-01-02	2	24.0						_		4.51			 	 	
80	T7-BA-01-02	2	22.4					ļ	ļ		4.6			ļ		
93	S1-BA-01-02	2	19.03			ļ,	ļ				4.43	+		9 4.33	1.67	1447.75
3	T1-BE-02-03	3	21.3			ļ	<u> </u>	-	<u> </u>	 ;	4.0			4.33	1.07	1447.73
16	T2-BE-02-03	3	21.8							. .	4.13				ļ	
29	T3-BE-02-03	3	26.1			<u> </u>		ļ	<u> </u>		4.23			 	 	
42	T4-BE-02-03	3	26.7			1				5 22.26	4.14			+	 	
55	T5-BE-02-03	3	19.9			7 1.	4 19.2	2 49.	2 26.8	5 22.35	4.4			-		
68	T6-BE-02-03	3	21.9			ļ					1		<u> </u>		 	
81	T7-BA-02-03	3	22.3					_			4.60			 -	+	
94	S1-BA-02-03	3	23.8					<u> </u>			4.6			6 4.6		1082.38
4*	T1-BE-03-04	4	2					<u> </u>			4.6.			4.0	<u> </u>	1002.30
17	T2-BE-03-04	4	18.			· · · · · · · · · · · · · · · · · · ·								+	 	<u> </u>
30	T3-BE-03-04	4	21.3			1				-	4.3			 		1
43	T4-BE-03-04	4	19.8				 				4.3			 	 	
56	T5-BE-03-04	4	13.7								4.9				+	
69	T6-BE-03-04	4	16.1								4.9				 	
82	T7-BA-03-04	4	16.6			 			 						+	
95	S1-BA-03-04	4	18.1	9 15.3			1 15	30.7	9 29.0	51 -0.82	4.4 2 4.8			3 5.7	3 1.88	8 1021.13
. 5	T1-BE-04-05	5	17.2											3.7	1	1
18	T2-BE-04-05	5	19											+	+	†
31	T3-BE-04-O5	5	2											+	+	
44	T4-BE-04-05	5	14.9								6.6			+	 	+
57	T5-BE-04-05	5	22.4	18.3	2 2.0	1.6	55 23.2	23 40.1	1 38.2	2/1	0.0	o _l 98	27			

MPIS Baseline Soil Characteristics: Moisture Content, Bulk Density, Porosity, pH, TOC

Pan ID	Sample Name D	epth	% Moisture based on dry wt	% Moisture based on wet wt	Wt bulk Density, g/cc	Dry bulk Density, g/cc	Dry wt % moisture	% volume porosity	% volume water-filled porosity	% volume air-filled porosity	рН	TOC, ppm	Avg. % soil moisture (wet wt)	Avg. pH	Avg. bulk density (moist), g/cc	Avg. TOC, ppm
70	T6-BE-04-05	5	14.38	12.57	1.89	1.66	14.03	39.77	23.23	16.54	6.18	866				
83	T7-BA-04-05	5	16.52	14.18	1.85	1.57	18.1	42.94	28.4	14.54	6.37					
96	S1-BA-04-05	5	18.67	15.73	1.82	1.54	17.58	43.82	27.16	16.66	4.79					
7	T1-BE-O5-06	6	20.66	17.12							6.04			6.47		994.88
20	T2-BE-05-06	6	20.91	17.3							6.89	890				
33	T3-BE-05-06	6	17.64	14.99							6.46					
46	T4-BE-O5-06	6	18.04								6.95					
59	T5-BE-05-06	6	24.59	19.74							7.07	405				
72	T6-BE-05-06	6	20.6	17.08						1	6.08	1631				
85	T7-BA-05-O6	6	15.5								6.74					
98	S1-BA-05-06	6	15.87								5.5				·	
8	T1-BE-O6-07	7	18.61									685		6.57	1.72	615.00
21	T2-BE-06-07	7	21.18	. 17.48							6.99					
34	T3-BE-06-07	7	14.35								6.22					
47	T4-BE-06-07	7	31.15								7.61	420				
60	T5-BE-06-07	7	27.29		1.72	1.36	26.17	50.42	35.68	14.74	6.23			ļ.		
73	T6-BE-06-07	7	23.09								6.67					
86	T7-BA-O6-07	7	20.81							<u> </u>	6.47					
99	S1-BA-O6-07	7	18.27	15.45							5.38	837				
9	T1-BE-07-OS	8	no core						1				20.77	7.03		558.20
22	T2-BE-07-08	8	no core													
35	T3-BE-07-08	8	27.91	21.82							6.95	305	5			
48	T4-BE-07-OB	8	no core										<u> </u>		<u> </u>	
61	T5-BE-07-08	8	31.67								7.75		.1	ļ	ļ <u>.</u>	
74	T6-BE-07-08	8	28.27					<u> </u>			6.74					
87	T7-BA-07-08	8	24.72						ļ	<u> </u>	7.44					
100	S1-BA-07-08	8	19.2								6.27					
10	T1-BE-08-09	9	31.65				<u> </u>				7.08			7.21	1.98	395.00
23	T2-BE-08-09	9	32.52				<u> </u>				7.75				ļ	
36	T3-BE-08-09	9	36.93	26.97			<u> </u>		<u> </u>		7.48				ļ	
49	T4-BE-08-09	9	31.65								7.61		0			
62	T5-BE-08-09	9	34.05		1.98	1.5	31.77	45.33	47.77	-2.43						
75	T6-BE-08-09	9	21.99						<u> </u>		6.99					
88	T7-BA-08-09	9	32.49				<u> </u>					40:				<u> </u>
101	S1-BA-08-09	9	34.17								6.29				<u> </u>	
11	T1-BE-09-10	10	35.01								7.78			7.55	5	421.00
24	T2-BE-09-10	10	33.22								7.68				<u> </u>	
37	T3-BE-09-10	10	36.88	26.94							7.82	61.	3	1	1	<u> </u>

MPIS Baseline Soil Characteristics: Moisture Content, Bulk Density, Porosity, pH, TOC

Pan ID	Sample Name	Depth	% Moisture	% Moisture based on wet	Wt bulk Density,	Dry bulk Density,	Dry wt	% volume porosity	% volume water-filled	% volume air-filled	рН	TOC,	Avg. % soil moisture	Avg. pH	Avg. bulk density	Avg. TOC, ppm
			wt	wt	g/cc	g/cc	moisture	porosity	porosity	porosity		••	(wet wt)		(moist),	
.]										 	7.00	413			g/cc	
50	T4-BE-09-10	10	36.72								7.83	412				
63	T5-BE-09-10	10	35.8	26.36							7.74	396			 	
76	T6-BE-09-10	10	39.39	28.26		i				ļ	7.37	394				
89	T7-BA-09-10	10	33.36	25.01							6.93	334				
102	S1-BA-09-10	10	34.89	25.87							7.34	450	22.00	7 43	1.00	542.20
12	T1-BE-10-11	11	34.56	25.69	1.8	1.34			46.25	 	7.56	653	23.98	7.43	1.82	543.38
25	T2-BE-10-11	11	34.75	25.79		1.29	37		47.66		7.75	425		 	ļ	
38	T3-BE-10-11	11	35.96	26.45	1.74	1.29	34.67		44.68		7.36	432				
51	T4-BE-10-11	11	30.36	23.29	1.92	1.53			38.97		7.02	305				
64	T5-BE-10-11	11	38.2	27.64		1.26			48.55		7.55	492				ļ
77	T6-BE-10-11	11	28.93	22.44	1.92	1.5					7.76	933		L		
90	T7-BA-10-11	11	24.76	19.85	1.83	1.48					6.92	493				
103	S1-BA-10-11	11	26.00	20.67	1.84	1.46	26.6	47.07	38.72	8.35	7.52	614				525.00
13	T1-BE-11-12	12	37.03	27.03	•						7.41	555		7.39	<u> </u>	525.88
26	T2-BE-11-12	12	27.76	21.73		L					7.74	640		<u> </u>		
39	T3-BE-11-12	12	24.74	19.83			<u></u>			L	7.13	'337		ļ		
52	T4-BE-11-12	12	33.62	25.16							7.54	821	4	<u> </u>	<u> </u>	
65	T5-BE-11-12	12	28.69	22.29						<u> </u>	6.9	630			ļ	
78	T6-BE-11-12	12	28.24	22.02							6.87	393		 	 	
91	T7-BA-11-12	12	34.29	25.53			<u></u> .				7.73	510		 	ļ	
104	S1-BA-11-12	12	33.20	24.96			<u></u>	<u> </u>	<u> </u>		7.81	321	<u> </u>	<u></u>	<u> </u>	<u> </u>

APPENDIX B

LITHOLOGIC LOGS FOR TEST CELL SOIL BORINGS

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or	n_	L	OAK RIDGE	NATIONAL LABORATORY	
		By:_S	R. Sturm	Date: 10/23/94	Page: 1 OF 1
Hole	No .:	S1BA	· ·	Ground Elevation: NA-	,
Tota	l Dep	th: <u>1</u>	2' Rig Typ	de: ORNL U2CRT Locat:	ion: <u>Shakedown area</u>
				Sample Type: <u>Geoprobe "Megabore 2" OD ></u>	
		PORTS	MPIS	Data Verified By:	Date:
DEPTH (FEET)		SAMPLE	LITHOLOGY	DESCRIPTION	
0 - 2 -	CS	INTV	Li malo		23/4) soft, damp , weathered surface n (10YR5/8), friable clay faces on vertical
6 -	cs			High angle fracture at 7.8 CH CLAY: brown (7.5YR5/6). so and plastic, homogenous tegray silt laminae, authegen	tiff. damp, very fat xture w/greenish
10 -	cs			at 8.1', high angle fractu CH CLAY: brown (7.5YR5/6), st	re at 10.5 & 10./
12 -				and plastic, prominent bed	partings, homogenous
14					
16					
18					
20					

OL	<u>'n</u>	L	OAK RIDG	E NATIO	NAL LABORATORY	
					Date: 11/20/94	Page: <u>1 OF 1</u>
1 .					Ground Elevation: NA	
1					U2CRT Loc	
Auge	r Siz	e: <u>NA</u>		_Sample i	Type: <u>Geograbe "Megabore 2" O</u>	0 X 4'
			MPIS		Data Verified By:	Date:
	SAMPLE TYPE	SAMPLE INTV	LITHOLOGY		DESCRIPTI	ON
0 -						
		\ /				
		$ \setminus /$			SILTY CLAY: brown (10YA5/	3) soft well
		$ \setminus $			developed root mass, fria	ble, dry
2-	cs	V		CI	SILTY CLAY: vellowish bro	WD (10VR5/8) SOME
-		$I \wedge$			Fe oxides, soft, moist, foxidized w/depth, pebbles	
		I/V			oxidized w/depth, peobles	s @ 4 . It gray mottle
		/ \				
1		/				
1 -				•		
		\ /			CTLTV CLAV: vollowich has	we lt gray mottle
_		$ \setminus $		CL	SILTY CLAY: yellowish broabund Fe & MN oxides, pet	bles @ 5.5.3" thick
_		\cup			bed, 2 – 6mm nodules high	oly oxidized
Б-	cs	$ \ \ \rangle$				
		$ / \rangle$			CL AM:	N/m 4 / 4)
-		/ \		CH	w/ lt gray inter-lams , s	scattered pebbles
		/ \			CLAY: dk yellow brown (10 w/ lt gray inter-lams, seless mottling @ 8', well plastic, fracture @ 8.1'	defined beds, stiff, w/avnsum infill
8 -					prasere, in accare e ori	w/g/podiii 111/111
		N /				
-	1	$ \setminus $				
		$ \cdot $				
10 -	cs		<u> </u>	СН	CLAY: some dry desicated cracks from drying @ 9.4 well developed bedding, p	surfaces w/1 - 2mm , 9.6, & 11.4 ,v stiff
		$ / \rangle$,	well developed bedding, p	plastic, waxy appearance
-	1	$\parallel / \parallel \setminus$	===			
		ν,				
12-		-				
-	1					
14-	1		1			
			-			
-	1					
					•	
16 -	1					•
-	1					
18	1					
	1					
20	-		,			
1	1	1	1	1	10	

cs be a compared by: S.A. cole No.: T18A col	_Rig Type: _ORNL Sample [S	Gro U2CRT Type: Ge Data Ver	und Elev	ation: N Megabore	A Locatio _2" OD X		11 #1	
Depth: 12' Juger Size: NA Doject: POHTS MPI POHI SANGLE SAMPLE EETI TYPE INTY CS CS CS 6-	_ Rig Type: _ORNL Sample [S	U2CRT Type: <u>Ge</u> Data Ver	oprobe "I	Megabore	Locatio	4'		
TOJECT: NA MPIE SAMPLE INTV C1 CS CS CS	Sample	Type: <u>Ge</u> Data Ver	oprobe "I	Megabore :	S. OD X	4'		
CS C	тнолову	Data Ver		: <u> </u>			e:	-
CS CS CS CS	TH0L06Y							
0 - CS 2 - CS 5 - CS	CL	CTI TV						
2 - CS	CL							
2 - CS 6 -	CL	CTITY						
2 - CS 6 -	CL	CTI TV						
2 - CS 6 -	Secretary Control of the Control of	DIFIL (CLAY: di	k brown	(7.5YR3	/4) soft,	damp ,	
4 - CS	· · · · · · · · · · · · · · · · · · ·	abund i	humus, i	raat bu	ndles, T	opsoil		
4 - CS								
CS 6-							•	
CS 6-		CTLTV	CI AV- 100	ellowiel	h heawn	(10YR5/8)	friabl	_
CS 6-		damp. (occasio	nal Fe	stainin	ig from 2-	-5 ', apen	Ç .
CS 6-		root p	ores. b	locky s	tructure			
5-						•		
6-								
6-								
						-		
8-		LOST C	ORE 6'	TO 8'				
8-						•		
8-								
8-								
		- CL AV-	hrown (7 5YB5/	6) sti	ff damn	verv fa	t
		and pl	astic.	homogen	ous text	ff, damp, ure, pror	nounced b	èd
	===	at 9.4	gs <1mm . 10.2	. tnick, !'. 10.3	nigh ar ! and !	igle fract [1.5].	ures	
cs \								
				•				
10								
/ \	====							
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12								
4								
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16 -								
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18 -	la de la companya de							
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20 -								

OI.	· N -	L	OAK RIDGE	NATIONAL LABORATORY	
Prep	ared	By:_S	.A. Sturm	Date: <u>10/23/94</u> Page: <u>1 OF 1</u>	
Hole	No.:	T188		Ground Elevation: <u>NA</u>	
Tota	l Dep	th: <u> </u>	2 <u>. </u>	pe: ORNL U2CRT Location: Test Cell #1	_ -
				Sample Type: <u>Geoprobe "Megabore 2" OD X 4'</u>	—
			MPIS	Data Verified By:Date:	
DEPTH (FEET)	SAMPLE TYPE	SAMPLE INTY	LITHOLOGY	DESCRIPTION	
(reet)		INTV	LIMALOS	CL SILTY CLAY: dk brown (7.5YR3/4) soft. damp, abund humus, root bundles, Topsoil CL SILTY CLAY: yellowish brown (10YR5/8), friable damp, open root pores, blocky structure, scattered sandstone pebbles, argillaceous infill in verticle dessication fracts	
6-	cs			 CH CLAY: brown (7.5YA5/6), stiff, damp, very fat and plastic, homogenous texture, w/intbdd gray silt laminae, authigenic gypsum crystals 	
10				parallel continuous bedding, 10mm thick mudstone pebble bed at 9'	
12					
14					
16					
18	_				

epared	d By: S	A. Sturm	NATIONAL LABORATORY Oate: 10/23/94	Page: 1 OF 1
			Ground Elevation: NA	Tak tahun 1940 terkiti makasan kenali 1966 di Lumbur 1991 di
otal De	enth: 1	2' Rig Ty	DE: ORNL U2CAT LO	cation: <u>Test Cell #1</u>
ider S	ize: NA		Sample Type: <u>Geoprobe "Megabore 2"</u>	OD X 4'
_			Data Verified By:	Date:
EPTH SAMPLI	E SAMPLE	LITHOLOGY	DESCRIP	TION
	100			maniferant. Budung makeumken i In komine nakantuka Italia (1999) i ji
0 -				
				EVD2/4) coft dama
1	11//		CL SILTY CLAY: dk brown (7 abund humus, root bundle	es, weathered Surrace
	-1		of subjacent unit. Tops	oil
2 - C	s X			
	-1/			/40VDT /0\
-	- / \		CL SILTY CLAY: yellowish by damp, common fe stainin	g, open root pores.
	//		damp, common fe stainin blocky structure, lt gr	ay mottle
4-	(
	- []	/		
4	-1 \ /		•	
	\/			
Б- C	s X		CH CLAY: brown (7.5YA5/6). and plastic, homogenous	stiff. damp, very fat texture. w/greenish
	- / \		gray silt laminae, auth at 7.9', mudstone pebbl	egenic gypsum crystals
4	- [/ \	(<u>==</u>	at 7.9 , mudstone pebbl	e bed at 5
	/			
8-		#===		
ł	1		,	
4	-1 \ /	/ <u>===</u>		
	1 \/			
10 - 0	cs X			
}	-1/			
+	/ \	\===	CH CLAY: brown (7.5YA5/6), and plastic, prominent	stiff, damp, very fat bed partings
	/		dia plasera, prominent	The part of the pa
12 -	-			
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	-	-		
A 50	***			
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Drena	E E	By: S			NAL LABORATORY Page: 1 0F 1
					Date: 10/23/94 Page: 1 UF 1Ground Elevation: NA
					U2CAT Location: Test Cell #1
					Type: Geoprobe "Megabore 2" OD X 4'
-					Data Verified By:
DEPTH S	SAMPLE	SAMPLE	LITHOLOGY		DESCRIPTION
(FEET)	TYPE	INTV	23.1102.001		DESCRIPTION
0 - 1 2 - 1	cs				SILTY CLAY: dk brown (7.5YR3/4) soft, damp, abund humus, root bundles, weathered surface of subjacent unit, Topsoil SILTY CLAY: yellowish brown (10YR5/8), friable damp, common Fe staining, open root pores, blocky structure, lt gray mottle
4		\			
					Becoming stiff increased clay content occasional sandstone pebbles
5-	cs	\bigwedge		•	
8 -				СН	CLAY: brown (7.5YR5/6), stiff, damp, very fat and plastic, with intrbdd greenish gray silt laminae, flecks of Mn oxide, parallel continuous bedding, becoming very homogenous
10	cs	\backslash			
-		$/\setminus$			
12-					
14-					
16 -					
18 -					
20 -					

Borehole Summary Information OAK RIDGE NATIONAL LABORATORY Page: 1 0F 1 Prepared By: S.A. Sturm _____ Date: <u>10/23/94</u> Hole No.: TiBE __Ground Elevation: NA Total Depth: 12' Rig Type: ORNL U2CRT ____Location: Test Cell #1 Auger Size: NA Sample Type: Geoprobe "Megabore 2" 0D X 4' __Data Verified By:___ Date: Project: PORTS MPIS DEPTH SAMPLE (FEET) TYPE DESCRIPTION INTV CL SILTY CLAY: dk brown (7.5YR3/4) soft, damp , abund humus, root bundles, weathered surface of subjacent unit, Topsoil 2 -CS CL SILTY CLAY: yellowish brown (10YR5/8), friable damp. mod stiff, common Fe staining, open root pores, blocky structure, lt gray mottle, occassional sandstone pebbles CH CLAY: brown (7.5YR5/6), stiff, damp, very fat and plastic, homogenous texture, w/greenish gray silt laminae, authegenic gypsum crystals at 8.1', mudstone pebble bed at 9' cs Б 10 -CS CH CLAY: brown (7.5YR5/6), very stiff, damp, very fat and plastic, < 1mm bedding 12 14 16 -18

20

OL	·N _	Ļ	OAK RIDG	E NATIONAL LABORATORY
Prep	ared	Ву:_ Р	.M. Schlos	serDate: <u>11/13/94</u> Page: <u>1 OF 1</u>
1				Ground Elevation: NA
i				pe: ORNL U2CRT Location: Test Cell #1
1				_Sample Type: Geoprobe "Megabore 2" QD X 4'
	SAMPLE	PORTS		Data Verified By:Date:
(FEET)	TYPE	INTV	L17HOLOGY	DESCRIPTION
0 -				CL SILTY CLAY: brown (10YR5/3), 10" with well developed root mass.
2-	cs			CL SILTY CLAY: yellowish brown (10YR5/8), friable damp, abundant Fe staining from 2–5°, some scattered 2–4mm pebbles.
6-	Cs			CL SILTY CLAY: as above, with abundant Mn oxides in root pores, mottled light gray along per- imeter of root pores, friable, moist.
8-				CL SILTY CLAY: abundant root pores with infill of light gray as above, scattered Fe nodules 2-4mm mottled light gray throughout.
10 -	cs			CL CLAY: strong brown (7.5YR5/6), mottled light gray (7.5YRN7/), very stiff, plastic, very gyp- siferous at 9.5', no apparent voids.
12-				CL CLAY: as above, well defined bedding surfaces, mottled light gray in vertical areas, 2-4mm pebbles scattered throughout, stiff, moist, waxy apperance.
-				
14-				
_				
15 -				
18 -				
20 -				

ornl OAK RIDGE NATIONAL LABORATORY Page: 1 0F 1 Date: <u>11/13/94</u> Prepared By: R.M. Schlosser __Ground Elevation:<u>NA</u> Hole No.: T1-BH __Location: <u>Test Cell #1</u> Total Depth: 12' Aig Type: ORNL U2CRT Sample Type: <u>Geoprobe "Megabore 2" OD X 4'</u> Auger Size: NA Project: PORTS MPIS Data Verified By:_ DEPTH SAMPLE (FEET) TYPE DESCRIPTION LITHOLOGY O CL SILTY CLAY: brown (10YR5/3). 10"-11" well developed root mass, becoming yellowish brown (10YR5/8) at 1', abundant Fe staining at 2.3'. 2 CS CL SILTY CLAY: yellowish brown (10YR5/8). mottled light gray along root pores, friable. 4 CL SILTY CLAY: as above, becoming mottled both brown and light gray throughout, abundant remnant pores, scattered Mn oxides and Fe staining, some scattered Fe nodules. CS Б. CL CLAY: yellowish brown (7.5YR5/6) mottled light gray (7.5YRN7/), blocky, plastic, stiff, waxy apperance, scattered very fine roots, well defined bedding planes, HIGH ANGLE FRACTURES at approximately 9.5' and 10.5' FRACTURE at 11' has light gray clay infill. Very mottled throughout, some remnant roots, very stiff, plastic, waxy apperance, becoming completely brown (7.5YR5/6) with no mottle at 11.5'. 8 cs 10 -12 14 15 18 20

UI II.			E NATIONAL LABORATORY
			ser Date: <u>11/13/94</u> Page: <u>1 OF 1</u>
			Ground Elevation: NA
			pe: DRNL U2CAT Location: Test Cell #1
			Sample Type: <u>Geoprobe "Megabore 2" OD X 4"</u>
Project:_	PORTS	T	Data Verified By:Date:
(FEET) TYPE	INTV	LITHOLOGY	DESCRIPTION
0 -			
	\ /		CL SILTY CLAY: brown (10YR5/3), abundant roots, mottling, becoming yellowish brown at 1', abundant Fe staining beginning at 1.5', soft,
	$ \setminus / $		friable
z - cs	X		
	$ / \rangle$		(10,000,000,000,000,000,000,000,000,000,
	/ \		CL SILTY CLAY: yellowish brown (10YA5/8), mottled. friable, root hairs, scattered Fe staining.
4-			occassional nodules, abundant Fe staining at 5
	1	\	
-	$ \rangle /$		CL CLAY: brownish yellow (10YR6/6) mottled light brownish gray, abundant root pores, some Mn
6 - CS	$ \vee $		staining along pores, some staining on desiccated surfaces, friable
07 03			
	/		
	V - V		
8-			CL CLAY: dark brown (10YR2/2) becoming stiff. very mottled as above, abundant Mn staining along pores, some staining on desiccated
	1\ /		surfaces, friable
	$ \cdot \rangle /$		·
10 - CS	$\int \int$		CL CLAY: brownish yellow (10YA5/8) no mottling, very stiff, plastic, waxy appearance, damp
	$ \cdot $		High angle fracture at 11.5
	V '		
12 -			
14-			
16-			
18 -			
20 -			

OAK RIDGE NATIONAL LABORATORY Page: 1 0F 1 __Date:<u>11/13/94</u> Prepared By: R.M. Schlosser ___Ground Elevation: NA Hole No.: T1-BJ Location: <u>Test Cell #1</u> Total Depth: 12' Rig Type: ORNL U2CRT Auger Size: NA Sample Type: Geoprobe "Megabore 2" OD X 4' Date: Project: PORTS MPIS __Data Verified By:___ DEPTH SAMPLE (FEET) TYPE SAMPLE INTV DESCRIPTION Ò CL SILTY CLAY: brown (10YR5/3), abundant roots very friable CL SILTY CLAY: yellowish brown (10YR5/8), aboreous as above, very soft, some scattered two - 3 mm pebbles 2 -CS CL SILTY CLAY: Lt brownish yellow (10YR6/4) mottled yellowish brown (10YR5/6) very soft, friable abundant Fe staining. CL SILTY CLAY: Lt. brownish yellow (10YR6/4) abundant Mn and Fe staining along root pores, occasional nodules Б-CS CL CLAY: brownish yellow (10YR5/8) friable, some scattered Fe nodules, friable, blocky structure 8 CL CLAY: brownish yellow (10YR5/8), mottled lt gray on bedding surfaces cs siltstone in desiccation surfaces. 10 CL CLAY: as above with abundant well developed bedding surfaces, very stiff, waxy appearance 12 16 20

orn

UL.	5 H _	1.	OAK RIDG	E NATIONAL LABORATORY
Prep	ared	By:5	G.A. Sturm	Date: 12/13/94 Page: 1 OF 1
ł.				Ground Elevation: NA
Tota	l Dep	th:1	<u>12'</u> Aig Ty	/pe: ORNL U2CRT Location: Test Cell #1
				_Sample Type: <u>Geoprobe "Megabore 2" OD X 4'</u>
	ect:	PORTS	7	Data Verified By:Date:
	TYPE	INTV	LITHOLOGY	DESCRIPTION
0-				
		\ /		
-		$ \setminus /$		CL SILTY CLAY: dk brown (7.5YR3/4) soft, damp,
		$ \cdot \cdot $		abund humus, root bundles, Topsoil
2-	CS	ΙX		
		$ / \rangle$		
-		/ \		CL SILTY CLAY: yellowish brown (10YR6/6), friable damp, open root pores, blocky structure:
		/		damp, open root pores, blocky structure, Becoming v stiff, increasing silt content, limonite staining, abund Fe nods 5' to 7'
4-				Tamonia de Octaviang, abene la libes d'est
		\ /	***************************************	
		$ \setminus / $		
6-	cs	ΙV		
		$ \wedge $		
-		/		
		// \		
8-				CH CLAY: brown (7.5YR5/6) , stiff, damp, very fat and plastic, homogenous texture, w/intbdd
,		\ /		gray silt laminae, no visible fractures or
-		$ \setminus / $		alterations
		$ \ \ \ $		
10 -	cs	$ \lambda $		
_		$ / \rangle$		
		// \		
12-				
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15 -				
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40				
18-		ŀ		
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Or	n.	OAK	RIDGE	E NATIONAL LABORATORY	
Prep	ared	By: <u>S.R. S</u>	turm	Date: <u>12/13/94</u> Page: <u>1 OF 1</u>	9 5 GMG 3 55
		T1BL		The state of the s	
Tota	l Dep	th: <u>12'</u> F	Rig Typ	rpe: ORNL U2CAT Location: Test Cell #1	2 45 - 1
Auge	r Siz	e: <u>NA</u>		_Sample Type: <u>Geoprobe "Megabore 2" OD X 4'</u>	Bara Carl
		PORTS MPIS	<u> </u>	Data Verified By:Date:	
DEPTH (FEET)		INTV LITH	OLOGY	DESCRIPTION	
	cs cs	SAPPLE LITH	0.06Y	CL SILTY CLAY: dk brown (7.5YR3/4) soft, damp, abund humus, root bundles, Topsoil CL SILTY CLAY: yellowish brown (10YR6/6), friable damp, open root pores, blocky structure. Becoming v stiff, increasing silt content, limonite staining, abund Fe nods 5 to 7. CH CLAY: brown (7.5YR5/6), stiff, damp, very fat and plastic, homogenous texture, w/intbdd gray silt laminae, no visible fractures or alterations	
16					
18	_				
20	4				

OL.	11 <u>-</u>			GE NATIONAL LABORATORY
				Date: 10/22/94 Page: 1 OF 1
				Ground Elevation: NA
				ype: ORNL U2CRT Location: Test Cell #2
				Sample Type: <u>Geoprobe "Megabore 2" OD X 4'</u>
			MPIS	Data Verified By:Date:
OEPTH (FEET)		SAMPLE INTV	LITHOLOGY	DESCRIPTION
0 -				
٦		N /		
		\ /		CL SILTY CLAY: dk brown (7.5YR3/4) soft, damp ,
		$[\] / [$		abund humus, root bundles, weathered surface of subjacent unit, Topsoil
2 -	cs	V		or subjacent unit, Topsoff
		$ \wedge $		
		$ / \rangle$		CL SILTY CLAY: yellowish brown (10YR5/8), friable
İ		I/ \		damp, mod stiff, common Fe staining, open_root pores, blocky structure, lt gray
4-				mottle, occassional sandstone pebbles
7		1	/	
		$ \setminus /$		Fe nodules up to 10 mm, verticle dessication
		$ \setminus $		planes with Mn oxide dendritic staining
5 -	cs	ΙV		
.		$ \wedge $		
		I/Λ		
		/ \		<u>-</u>
8 -		<u> </u>		CH CLAY: brown (7.5YA5/6), stiff, damp, very fat and plastic, homogenous texture, w/greenish
		\		and plastic, homogenous texture, w/greenish gray silt laminae, authegenic gypsum crystals
4		$ \setminus /$		gray silt laminae, authegenic gypsum crystals at 7.9', mudstone pebbles in gray silt bed at 9.2'
		$ \cdot $		3.2
10 -	cs	X		<u> </u>
		I / I		
-		/ \		
		/ '		<u></u>
12-		 	+	=
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14-				
-				
15 -				
-	1			
18 -	ł			
-	1			
20 -	-			

OAK RIDGE NATIONAL LABORATORY Prepared By: S.A. Sturm Page: 1 OF 1 _Date:<u>10/22/94</u> __Ground Elevation: NA Hole No.: T2BB Total Depth: 12' Rig Type: ORNL U2CRT Location: <u>Test Cell #1</u> Auger Size: NA Sample Type: Geoprobe "Megabore 2" OD X 4' Project: <u>PORTS MPIS</u> Data Verified By:_ Date: OEPTH SAMPLE (FEET) TYPE DESCRIPTION LITHOLOGY INTV 0 -CL SILTY CLAY: dk brown (7.5YR3/4) soft. damp , abund humus, root bundles, weathered surface of subjacent unit, Topsoil cs 2 -CL SILTY CLAY: yellowish brown (10YR5/8), friable damp. mod stiff, common Fe staining, open root pores, blocky structure, lt gray mottle, scattered Fe nodules CH CLAY: brown (7.5YR5/6), stiff, damp, very fat and plastic, homogenous texture, w/greenish gray silt laminae cs 6 8 $\ensuremath{\text{CLAY:}}$ as above with 2 - 4 mm beds of silt with dessication fractures 10 -CS 12 -14 15 18 20

1	OL.	.[]	L	OAK RIDG	E NATIONAL LABORATORY	
	Prep	ared	By: S	.R. Sturm	Date: 10/22/94 Page: 1 OF 1	
- {	Hole	No.:	T2BC		Ground Elevation: NA	
	Tota	l Dep	tn: <u>1</u>	2' Rig Ty	pe: <u>ORNL U2CRT Lacation: Test Cell #2</u>	
	Auge	r Siz	e: <u>NA</u>		Sample Type: <u>Geoprobe "Megabore 2" OD X 4</u>	
	-	_		MPIS	Data Verified By:Date:	
	DEPTH (FEET)	SAMPLE TYPE	SAMPLE INTV	LITHOLOGY	DESCRIPTION	
	0 -					
			Λ			
			\setminus		CL SILTY CLAY: dk brown (7.5YR3/4) soft, damp,	~
			$ \setminus $		abund humus, root bundles, weathered surface	
ĺ	2	cs	I V		of subjacent unit, Topsoil	
	_		$I \wedge$			
				*************	CL SILTY CLAY: yellowish brown (10YR5/8), friab	le .
			/ \		damp, mod stiff, common Fe staining,	••
	4-		<u> </u>		open root pores, blocky structure, lt gray mottle, scattered Fe nodules	
			N /			
			$ \rangle /$		en en en en en en en en en en en en en e	
			$ \setminus $		in the state of th	
	6-	, CS	J V			•
			$ \ \ $			
	_				 CH CLAY: brown (7.5YR5/6), stiff, damp, very fa and plastic, homogenous texture, w/greenish 	t
			// \		and plastic, homogenous texture, w/greenish gray silt laminae, flecks of Mn oxide	
	8-		(——		3 . 2, 2. 2 2. 2. 2. 2. 2. 2. 2. 2. 2. 2. 2. 2. 2.	
	·		N /			
	-	ļ	$1 \setminus I$			
			$ \cdot \cdot $			
	10 ~	cs	X		Mudstone pebbles at 10.5'	
			$ / \rangle$			
	-		I/ \setminus		High angle fracture at 11.4'	
			// \			
	12 -	1	-		· .	
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	14-					
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	16 -	1				
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OAK RIDGE NATIONAL LABORATORY Page: <u>1 OF 1</u> ____Date:<u>10/22/94</u> Prepared By: S.A. Sturm __Ground Elevation:__NA Hole No.: T2BD Location: Test Cell #2 Total Depth: 12 Rig Type: ORNL U2CRT Sample Type: <u>Geoprobe "Megabore 2" OD X 4'</u> Augen Size: NA _Data Verified By:___ Date:_ Project: PORTS MPIS DEPTH SAMPLE (FEET) TYPE SAMPLE INTV DESCRIPTION 0 CL SILTY CLAY: dk brown (7.5YR3/4) soft, damp, abund humus root bundles, weathered surface of subjacent unit, Topsoil CL SILTY CLAY: yellowish brown (10YA5/8), friable damp. mod stiff, common Fe staining, open root pores, blocky structure, lt gray mottle, scattered Fe nodules up to 20 mm 2 cs CH CLAY: brown (7.5YR5/6), stiff, damp, very fat and plastic, homogenous texture, w/greenish gray silt laminae, authegenic gypsum crystals at 8 6 CS В cs 10 Prominent bed partings, silt with dessication fracturing at 10.6 and 11.4 $^{\prime}$ 12 14 16 18

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ren	ared			SE NATIONAL LABORATORY Date: 10/22/94 Page: 1 OF 1	
				Ground Elevation: NA	
				ype: ORNL U2CRT Location: Test Cell #2	
				Sample Type:Geoprobe "Megabore 2" QD X 4'	
				Data Verified By:Date:	
ОЕРТН	SAMPLE	SAMPLE	LITHOLOGY	DESCRIPTION	
(FEET)	TYPE	INTY			
0 -				CL SILTY CLAY: dk brown (7.5YR3/4) soft. damp , abund humus. root bundles, weathered surface of subjacent unit. Topsoil	
2-	cs			CL SILTY CLAY: yellowish brown (10YR5/8), friable damp, mod stiff, some Fe nodules, open root pores with Mn infill, blocky structu	
4 -		(<u>'</u>			
		\ /		·	
- 5 -	cs			CH CLAY: brown (7.5YR5/6), stiff, damp, very fat and plastic, homogenous texture, w/greenish gray silt laminae	
0 -					
-		/ \			
		// /			
8 -	-				
	l	1\ /			
-	1	$ \cdot $			
		$ \ \ $			
10 -	cs	ΙX			
		$ / \setminus$		Prominent bed partings, silt with dessication fracturing	
12		<u>/</u>	<u> </u>		
16					
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14	1				•
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UL	<u> </u>			NATIONAL LABORATURY
Prepa	ared	Ву:_ Р	M. Schloss	pr Date: 11/15/94 Page: 1 OF 1
		T2-B		
Tota	l Dep	th: <u>1</u>	<u>2'</u> Aig Ty	
				Sample Type: Geoprobe "Megabore 2" OD X 4' Data Verified By:
	SAMPLE	SAMPLE	MPIS	DESCRIPTION
(FEET)	TYPE	INTV	CI Macair	
0-				
		\ /		(40VDE/7) 40"-44" with poots
1 1	cs	$ \setminus $		CL SILTY CLAY: brown (10YR5/3), 10"-11" with roots very wet from surface penetration of rain
		$ \cdot \rangle $		CL SILTY CLAY: yellowish brown (10YR5/8), mottled
2-		ΙŽ		itaba abau with poot mass occassioned for .
		$ / \rangle$		staining, becoming much more stained with depth occassional Mn staining, soft, friable, damp.
		// \		Mn staining increasing with depth, mottling very light gray
4-				
4-		1		
				CL SILTY CLAY: as above, becoming denser, firm
	cs	$ \setminus $		
5-		Y		
-		/		
		// \		
8-				CL CLAY: yellowish brown (7.5YA5/6) mottled light
		1		CL CLAY: yellowish brown (7.5YA5/6) mottled light gray (7.5YAN7/), damp, very firm, scattered nodules, becoming less mottled with depth @ 9.5 clay layer 5mm thick, scattered remnant
-	cs			root masses.
		$ \ \ $		CL CLAY: yellowish brown (7.5YR5/6) well defined
10 -	1	1 1		bedding planes, stiff, fat, damp, slight waxy
•		1/		appearance
•	1	/ \	LEE .	
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	1 .			
20	1			

OAK RIDGE NATIONAL LABORATORY Prepared By: R.M. Schlosser ____ Date: <u>11/16/94</u> Page: 1 OF 1 __Ground Elevation: NA Hole No.: T2BG Location: <u>Test Cell #2</u> Total Depth: 12' Aig Type: ORNL U2CAT Auger Size: NA Sample Type: Geoprobe "Megabore 2" 0D X 4' Data Verified By:___ Project: PORTS MPIS Date: DEPTH SAMPLE (FEET) TYPE LITHOLOGY DESCRIPTION INTV CL SILTY CLAY: brown (10YR5/3). 10"-11" with roots becoming yellow brn lt gray mottle, some pebs. highly oxidized Fe and Mn @ 3.5', soft, friable lt gray mottle on vertical planes— old fracs? cs CL SILTY CLAY: yellowish brown (10YR5/8), abund pebs @ 5, firmer, damp, strong Fe staining, becoming less silty, dk gray brn alteration in root vesicles, blocky structure cs CL CLAY: yellowish brown (7.5YA5/4) mottled light gray (7.5YAN7/), firm, plastic, occ gypsiferous zones, some large crystal development in voids, strong red oxides on vertical micro-fractures, gray layer w/mudstone on desiccated surface 8 10 CS CL CLAY: yellowish brown (7.5YR5/6) well defined bedding planes, stiff, fat, damp, slight waxy appearance, gypsum in voids mottled strong brn 12 14 15 18

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OAK RIDGE NATIONAL LABORATORY Page: 1 OF 1 Prepared By: A M Schlosser Date: 11/16/94 __Ground Elevation: NA Hole No.: T2-BH Location: <u>Test Cell #2</u> Total Depth: 12' Aig Type: ORNL U2CRT Sample Type: <u>Geoprobe "Megabore 2" OD X 4'</u> Auger Size: NA _Data Verified By:__ Project: <u>POATS MPIS</u> DEPTH SAMPLE (FEET) TYPE DESCRIPTION LITHOLOGY 0 CL SILTY CLAY: brown (10YR5/3), 10"-11" with roots very wet from pooled surface water CL SILTY CLAY: brownish yellow (10YR6/8), mottled reddish brown from Fe staining and lt gray mottle, occasional root hairs, friable, Mn staining on vertical frac surfaces ÇS 2 -CL SILTY CLAY: as above, dense, abund Fe oxides, less silty, damp, scattered 1 — 3mm pebbles cs CL CLAY: yellowish brown (10YR5/4) mottled light gray (10YR6/1), gray clay on vertical fractures (.3 mm), stiff, damp CL CLAY: yellowish brown (10YR5/4) well defined bedding planes, stiff, waxy appearance, v fat high angle fracture @ 11' 10 cs 12 -14 16 18

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U				GE NATIONAL LABORATORY
1				<u>sserDate:11/16/94Page:1_OF_1</u>
1				Grauna Elevation: NA
Tota	1 Der)th:1	2' Aig Ty	ype: ORNL U2CRT Location: Test Cell #2
Auge	r Siz	e: <u>N</u> A	·	Sample Type: <u>Geoprobe "Megabore 2" OD X 4'</u>
		PORTS	MPIS	Data Verified By:Date:
DEPTH IFEETI	SAMPLE TYPE	SAMPLE INTV	LITHOLOGY	DESCRIPTION
2	cs			desiccated surface, gypsiferous zones @ 7.9'and at 8.0' some crystal development in vugs 3-5mm, CL STETYDEPAT: 8570wA (15985/3), 8 - 10" with roots organic rich, pooled surface water CL SILTY CLAY: brownish yellow (10Y86/8), mottled
4-				lt gray , some Fe oxides, friable, damp, slight gypsiferous
6-	CS			CL SILTY CLAY: as above, friable, abund Fe oxides a few scattered 2 – 3 mm well rounded pebbles
8-				CL CLAY: yellowish brown (10YR5/4) mottled light gray (10YR6/1), gray clay on vertical micro-frac
10 -	cs			CL CLAY: yellowish brown (10YR5/4) well defined bedding planes, stiff, waxy appearance, v fat high angle fracture @ 10, more homogenous @ 10.5
14-				
16 -		-		
18 -				
20 -				

OAK RIDGE NATIONAL LABORATORY Date: 11/16/94 ___Page:__1_0F_1 Prepared By: R.M. Schlosser _ Ground Elevation: <u>NA</u> Hole No.: T2-BJ Total Depth: 12' Rig Type: ORNL U2CRT Location: Test Cell #2 Auger Size: NA Sample Type: Geoprobe "Megabore 2" OD X 4' Data Verified By:_ Project: <u>PORTS MPIS</u> DEPTH SAMPLE SAMPLE (FEET) TYPE INTV DESCRIPTION 0 -CL SILTY CLAY: brown (10YR5/3), 8 - 10" with roots organic rich, wet from surface water CL SILTY CLAY: brownish yellow (10YR6/8), mottled lt gray , Fe staining throughout, soft, friable scattered oxidized s.s. pebbles cs CL SILTY CLAY: as above, friable, abund Fe oxides a few scattered 1-3mm pebbles. It gray mottle CS 5 CL CLAY: yellowish brown (10YR5/4) mottled light gray (10YR6/1), low angle fracture 1-3mm wide with lt gray silt infill, very firm, blocky structure, slighty waxy appearance, gypsiferous @ 9.0' - 9.5 8 CL CLAY: yellowish brown (10YR5/4) gray reduction occuring on vertical .2 - .5 mm bands, v fat waxy appearance, more homogenous @ 11.5' cs 10 -12-16 18 20

OI.	11.			E NATIONAL LABORATORY	
				Date: <u>12/13/94</u> Page: <u>1 OF 1</u>	
1				Ground Elevation: NA	
				pe: <u>ORNL U2CRT</u> Location: <u>Test Cell #2</u>	
				Sample Type: <u>Geoprobe "Megabore 2" OD X 4'</u>	
	ect:_		T	Data Verified By:Date:	
	TYPE	INTV	LITHOLOGY	DESCRIPTION	
0-			(20.000)		
		\ /			
-		$ \rangle /$		CL SILTY CLAY: dk brown (10YR5/3) soft. damp	,
		$ \cdot \cdot $		abund humus, root bundles. Topsoil	
2-	cs	X		CL SILTY CLAY: brownish yellow (10YR6/6), fri damp, lt gray mottle, blocky structure,	able
		$ / \rangle$		Fe staining common, scattered oxidized	
-		 / \		sandstone pebbles	
		/ \			
4-					
		$ \rangle /$			
		$ \setminus $			
6-	cs	I Y	**************************************		
-		$ f \rangle$			
ļ		// \			
8-	-	(CH CLAY: brown (7.5YR5/6), stiff, damp, very and plastic, homogenous texture, w/intbdd	/ fat
		\setminus		gray silt laminae, horizontal fracture wi additional moisture at 7', bedded gypsum a	th
•		$ \setminus $		additional moistore at 7, bedded gypsum a	36 0
		$\mid \bigvee$			
10 -	cs	$ \ \ \rangle$			
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or	n	L	OAK RIDG	NATIONAL LABORATORY	
			.A. Sturm		
Hole	No.:	ТЗВА		Ground Elevation: NA	
Tota	l Dep	th: <u> </u>	<u>2'</u> Rig Ty	De: ORNL U2CAT Location: Test Cell #3	. 1 1. 191
				Sample Type: Geoprobe "Megabore 2" OD X 4'	
	ect:_	PORTS	MPIS	Data Verified By:Date:	
(FEET)		INTV	LITHOLOGY	DESCRIPTION	
2-	cs			CL SILTY CLAY: dk brown (7.5YR3/4) soft, damp , abund humus, root bundles, Topsoil	
4-	Cs			CL SILTY CLAY: yellowish orange (10YR6/6), mod stiff, damp, common Fe nods, with some Mn nods root pores with Mn infill	
8-	cs			CL SILTY CLAY: It brown (10YR5/6), with It gray silt laminations. mod stiff, some yellowish orange clay inter-laminae, moist, light gray silt laminations up to 5mm at 11	
12				CH CLAY: yellowish red (5YR4/6), damp, stiff, fat and plastic, with lt gray silt laminae up to 3m oxidized Mn O2 nodules forming crystalline druzs around a soft mineral core	nm -
16	-				
18	-				

DED OAK BIDGE NATIONAL LABORATORY

Hole No. T3BB	Ground Elevation: NA
	Type: ORNL U2CRT Location: Test Cell #3
	Sample Type: Geoprobe "Megabore 2" OD X 4'
	Data Verified By:Date:
DEPTH SAMPLE SAMPLE LITHOLOGY	DESCRIPTION
0	
	7 EVD2/4)
	CL SILTY CLAY: dk brown (7.5YR3/4) soft, damp , abund humus, root bundles, Topsoil
2 - cs V	
[] [s \	
	CL STITY CLAY: vallowich hoows (40VD5/9) mad
	CL SILTY CLAY: yellowish brown (10YR5/8), mod stiff, damp, common Fe nods, with some Mn nods
4 .	
	Decreasing nodules
	- - -
6 - CS	
	CH CLAY: 1t yellow brown (10YR6/4), with 1t gray
	silt laminations, mod stiff, damp, laminar bedding, fat, mod plastic
8-	
10 - CS X	
	CH CLAY: 1+ vollow boo (2.5VD6/4) damp very fat
	CH CLAY: Lt yellow brn (2.5YR6/4), damp, very fat and plastic, with lt gray silt laminae.
12	pronounced bed partings
14-	
15	
18 -	
30	
20 -	

or	n.	L OAK	DGE NATIONAL LABORATORY	
Prepa	ared		m Date: 10/20/94 Page:_	1 OF 1
		T3BC		The second secon
			Type: ORNL U2CRT Location: Test	Cell #3
			Sample Type: <u>Geoprobe "Megabore 2" OD X 4'</u>	
	ect:_	PORTS MPIS		Date:
(FEET)		INTV LITHO	DESCRIPTION	en communication and armong to the contraction of t
2 -	cs		CL SILTY CLAY: dk brown (7.5YR3/4) so abund humus, root bundles, Topsoil CL SILTY CLAY: yellowish brown (10YR5, stiff, damp, common Fe nods, with	/8), mod
6-	cs		CH CLAY: lt yellow brown (10YR5/4), w silt laminations, stiff, damp, lam bedding, fat, mod plastic	ith lt gray inar
10 -	-		CH CLAY: Lt yellow brn (2.5YA6/4), da and plastic, v. stiff, lt gray sil pronounced bed partings	mp. very fat t laminae.
14-				
15				
18-				
20	4			

ole No.: otal Dep uger Sız	_ <u>T3BD</u> th: <u>12'</u> Rig_Ty e:NA	Date: 10/21/94 Page: 1 OF 1
otal Dep uger Siz roject: DEPTH SAMPLE TYPE	th: 12' Rig Ty e: NA PORTS MPIS SAMPLE TRUE SE	ype: ORNL U2CRT Location: Test Cell #3 Sample Type: Geoorobe "Megabore 2" OD X 4' Data Verified By: Date:
uger Siz	e: NA PORTS MPIS	Sample Type: <u>Geoprobe "Megabore 2" QD X 4'</u> Data Verified By:Date:
POJECT:	PORTS MPIS	Data Verified By:Date:
PEPTH SAMPLE FEET) TYPE	SAMPLE	
	(
0 -	<u> </u>	
2 - CS		CL SILTY CLAY: dk brown (7.5YR3/4) soft, damp , abund humus, root bundles, Topsoil
4 -		CL SILTY CLAY: yellowish brown (10YR6/8). mod stiff, damp, common Fe nods, some are dusky red. root pores w/ fine sand infill
6 - CS		CH CLAY: yellow brown (10YR5/6), with pale olive (5y6/3) mottle, stiff, damp, laminar bedding, coarse sand lens at 7.5' & 7.8'
8-		
10 - CS		CH CLAY: Lt yellow brn (2.5YR6/4), damp, slighty
12 -		plastic with 1t gray silt laminae. pronounced bed partings
14-		
_		
15 -		
18 -		

OAK RIDGE NATIONAL LABORATORY

.B. Sturm Oate: 10/20/94

		7	.H. Sturm	Bate: 10/20/94 Page: 1 0/14
				그는 사람들은 사람들이 살아보고 있다면 살아보고 있다면 사람들이 되었다면 살아보고 있다면 살아보고 싶다면 살아보고 싶다면 살아보고 싶다면 살아보고 싶다면 살아보고 싶다면 살아보고 있다면 살아보고 싶다면 살아보고 싶다면 살아보고 싶다면 살아보고 싶다면 살아보고 싶다면 살아보고 싶다면 살아보고 싶다면 살아보고 싶다면 살아보고 싶다면 살아보고 싶다면 살아보고 싶다면 살아보고 싶다면 살아보고 싶다면 살아보고 싶다
				The first of the conventional property and the conventional property of the conventional convent
			MPIS	Sample Type: <u>Geoprobe "Megabore 2" OD X 4'</u> Data Verified By:
DEPTH	SAMPLE	SAMPLE	LITHOLOGY	DESCRIPTION
IFEET	TYPE	INTV	LI INOCOG!	DESCRIPT TON
0	-			
		\ /		
	-	$ \setminus / $		CL SILTY CLAY: dk brown (7.5YR3/4) soft, damp , abund humus, root bundles, Topsoil
		- \/		applie Homos, 7000 ballotes, 1050011
.2	- cs	X	**************************************	
	-	/ \		CL SILTY CLAY: yellowish brown (10YR5/8), mod stiff, damp, common Fe nods, Fe oxide on blocky
		// \		structure clay faces
4	1			
		1		
'	1	$ \rangle /$		
j		$ \cdot \rangle /$		
6	- cs	ΙX		Clay as above but larger Fe nodules that are dusky red (10R3/4)
] / \		dddky 763 (1010) 4)
	-	I/ \setminus		
		// \		
l e	4			CH CLAY: dk yellow brown (10YR4/4), very stiff, damp, very fat and plastic, with lt gray silt
		1\ /		laminations
	4	$\mathbb{N}/$		
		$ \cdot $		
10	- cs) X		
		$ / \rangle$		
	-	I/ \setminus		CH CLAY: Lt yellow brn (2.5YA6/4), damp, very fat and plastic, with lt gray silt laminae,
1.		// /		pronounced bed partings
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OPPI OAK RIDGE NATIONAL LABORATORY

Prep	ared	By: R	.M. Schlos		~~~			9/94			OF 1		
			2' Rig Ty									}	
								ore 2"	0D X 4				
	ect:_	PORTS	MPIS LITHOLOGY		Data Ve	ritied		ESCRIP	TION	Da	te:		
(FEET)	TYPE	INTV	CTINOCOS					LJChir	1011			<u></u>	
2-	CS				SILTY stiff, heavy	CLAY: damp, fe sta	root yellow	ish brite side bundle	es, we rown (tainir s @ 4'	4) soft from (10YR5/8 ng 2 - 3 , lt gr	injed 3), mo 3'. fr	tion d ciable	
_													
6 -	cs			СН	CLAY: and Mr vession	yellow n oxide cles to	wish br es @ 5 o 7', t	own 1 - 5.5 pecomi	t gray ', sca ng fin	mottle attered mer, wa	e, abu root axy	nd Fe hairs	
8-								,		<i>(</i> •)		24	
10 -	cs			CH	CLAY: as ab	dk ye. ove, wi	llow br ell de	rown (fined	10YH4/ gray t	(4), mo peds at	8.4°	lt gray @ 8.6'	y
-		$\left / \right $			lost	core 10	0.1'-	12.0'					•
12 -		\(\frac{1}{2}\)							•				
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(or	<u>n</u>	<u> </u>	OAK RIDG	SE NATIONAL LABORATORY
. [Prepa	ared	By:R	.M. Schlos	Ser Date: 11/19/94 Page: 1 OF 1
	Hole	No.:	<u> </u>		Ground Elevation: NA
					ype: ORNL U2CRT Location: Test Cell #3
	Auger	r Siz	e: <u>NA</u>		Sample Type: <u>Geoprobe "Megabore 2" OD X 4'</u>
		ect:_		MPIS	Data Verified By:Date:
	(FEET)		SAMPLE INTY	LITHOLOGY	DESCRIPTION
	0 -				CL SILTY CLAY: dk brown (7.5YR3/4) soft, damp , established root system to 12", friable
	2 -	CS			CL SILTY CLAY: yellowish brown (10YR5/8), mod stiff, damp, abund Fe staining 1.5'– 4', friable Mn oxides and Fe nods from 3' – 5.5', oxides on vertical fractures and root vesicles
	4 -	·			homogenous at 10'- 12', waxy appearance, damp, low angle fractures at 7'& 7.5'
	6 -	cs			CH CLAY: yellowish brown lt gray mottle, becomes more defined @ 8.5' w/ mottling on vertical micro-fracs, firm, blocky structure, lower
	8 -				
	10 -	cs			CH Clay: yellowish brn 10YR5/8, as above, v stiff, well defined gray bed @ 9', 3 – 6mm thick
	12-				
	14-				
	16 -			-	
,	18 -				
	20 -				

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Borehole Summary Information

OAK RIDGE NATIONAL LABORATORY

	sserDate:_ <u>11/19/94</u> Page:_ <u>1_0F_1</u>
Hole No.: <u>T3BH</u>	Ground Elevation: NA
Total Depth: 12' Rig T	
	Sample Type: Geoprobe "Megabore 2" OD X 4'
Project: PORTS MPIS DEPTH SAMPLE SAMPLE LITHOLOGY	Data Verified By:Date:
(FEET) TYPE INTY	DESCRIPTION
0 -	
	CL SILTY CLAY: dk brown (7.5YR3/4) soft. damp . some yellow brown mottling, friable
2 - CS /	CL SILTY CLAY: yellowish brown (10YR5/8), mod stiff, damp, abund Fe staining 3'— 4', blocky structure, scattered Fe ox and pebbles 4.5' 5.5'
	structure, stattered Fe ox and peobles 4.5 5.5
4-	
6- CS X	CH CLAY: yellowish brown lt gray mottle, some Mn oxides, scattered 2–3mm peobles @ 7.5 well rnd
	becoming firm, Mn oxide in remnant root vesicles high angle fracture at 8.5
8-	
	CH CLAY: dk yellow brown (10YR4/4), mottled lt gray on beds and on vertical micro-fracs, trace roots @ 9', very stiff, well defined bedding, waxy
10- CS	
12-	CH ClAY: as above also mottled brownish yellow
16 -	
18-	
20 -	

OL.	11	<u>L</u>		SE NATIONAL LABORATORY
			M. Schlos	一个一个一个一个一个一个一个一个一个一个一个一个一个一个一个一个一个一个一个
				Ground Elevation: NA
				ype: ORNL U2CRT Location: Test Cell #3
Auge	r Siz	e: <u>NA</u>		Sample Type: <u>Geoprobe "Megabore 2" OD X 4'</u>
			MPIS	Data Verified By:Date:
	SAMPLE TYPE	SAMPLE INTV	LITHOLOGY	DESCRIPTION
2 -	cs			CL SILTY CLAY: dk brown (10YR5/3) soft. damp , abund roots, organic rich CL SILTY CLAY: yellowish brown (10YR5/8), lt gray mottle, some Fe staining @ 1' common by 4', soft
	cs			friable, red oxidized peobles. less silt w/depth CL as above, abund peobles 6 – 12mm dia., angular, heavy Fe staining
5 - 8 -				CL SILTY CLAY: yellowish brn, lt gray mottle on bed ding and some in vertical fractures, gypsiferous at 7.5', high angle fracture at 7.5'.
10 -	cs			CH CLAY: yellowish brown (10YR5/8), mottled lt gray on beds, very thin distinct bedding, plastic, moist
12-		<u> </u>		
14-				
16 -				
18 -				
20 -				

OFTIL OAK RIDG	SE NATIONAL LABORATORY
Prepared By: A.M. Schlos	serDate: <u>11/19/94</u>
1	Ground Elevation: NA
	ype: <u>ORNL U2CRT</u> Location: <u>Test Cell #3</u>
	Sample Type: <u>Geoprobe "Megabore 2" OD X 4'</u>
ent Laure Laure 5	Data Verified By:Date:
(FEET) TYPE INTY LITHOLOGY	DESCRIPTION
0-	
	CL SILTY CLAY: dk brown (10YR5/3) soft, damp , abund roots, organic rich
2 - CS \	CL SILTY CLAY: yellowish brown (10YR5/8), lt gray mottle, Fe staining throughout, friable, soft
] / \	
4	
	CL SILTY CLAY: as above, _abund pebbles @ 5 - 6.5
	CL SILTY CLAY: as above, abund pebbles @ 5'- 6.5' slight fracture at 6.7', less silty @ 6.5. gypsiferous @ 7.5', friable, stiffer @ 7'
6- CS X	
	CH CLAY: yellowish brown (10YA5/8), mottled lt gray on beds, some lt gray on vertical micro fracs.
	lt gray reduced mudstone on dessicated surface @ 11.5', plastic, waxy appearance
8	
10 - CS X	
12	
14-	
16 -	
18 -	
20 -	

DED | DAK RIDGE NATIONAL LABORATORY

Prens	red		.A. Sturm	Date: 12/13/94 Page: 1 OF 1
i i		-	.A. SCOM	Ground Elevation: NA
				pe: ORNL U2CRT Location: Test Cell #3
				Sample Type: <u>Geogrape "Megabore 2" OD X 4'</u>
Proje	ect:_	PORTS	MPIS	Data Verified By:Date:
	SAMPLE TYPE	SAMPLE INTV	LITHOLOGY	DESCRIPTION
0 -				
-	cs	$ \cdot $		CL SILTY CLAY: dk brown (10YR5/3) soft, damp, abund humus, root bundles, Topsail
2-				CL SILTY CLAY: brownish yellow (10YR6/6), friable damp. It gray mottle, some Mn oxides Fe staining common, scattered oxidized sandstone pebbles
4				Sauracous bennies
	•	\setminus		
6-	CS	$ \bigvee $		CL CLAY: color change to 10 YR5/6. increasing clay content with olive green laminae, stiff, plastic
				content with office green familiae, Stirr, prosere
8-				CH CLAY: brown (7.5YR5/6), stiff, damp, very fat and plastic, plastic texture, w/intbdd
10-	cs			gray silt laminae, near vertical fracture at 6.7 becoming reddish at 11' – 12'
12 -				
_				
14-				
_				
16 -				
18 -				
20 -				

		DGE NATIONAL LABORATORY	
repared	By: <u>S.H. Stur</u>	m Date: <u>12/13/94</u> Page: <u>1 DF 1</u> Ground Elevation: NA	
		Type: ORNL U2CRT Location: Test Cell #3	
		Sample Type: Geoprobe "Megabore 2" OD X 4'	
		Data Verified By:Date:	
EPTH SAMPLE	SAMPLE LITTER DOW		
FEET) TYPE .	INTV		
0-	***********		
	/ /		
4		CL SILTY CLAY: dk brown (10YR5/3) soft. damp abund humus, root bundles, Topsoil	•
2 - CS	\ \ \ \ \ \ \ \ \ \ \ \ \ \ \ \ \ \ \ 	CL SILTY CLAY: brownish yellow (10YR6/6), fr	iable
	//	damp, lt gray mottle, some Mn oxides Fe staining common2.5 – 6.5', scattered o	xidized
-	/ \	sandstone pebbles	
	/ \	1970) - 1971 - 1971 - 1971	
4			
	\ /		
-			
	\/		
6 cs	\ \ \ \ \ \ \ \ \ \ \ \ \ \ \ \ \ \ \		
+	/ \ 		
8-		CH CLAY: brown (7.5YR5/6) , stiff, damp, ver and plastic, plastic texture, w/intbdd_	y fat
	\\	gray silt laminae, gypsum crystals at 8.8	3 '
. 1	1\ / ===		•
	1 // ====		
10 - CS		<u></u>	
		<u> </u>	
1		CH CLAY: becoming red brown	
12-			
1 .			
			•
14-	1		
1			
16 -			
1			
18 -			
1			

OAK RIDGE NATIONAL LABORATORY

Prepared By: S.A. Sturm	Date: 10/23/94 Page: 1 OF 1
Hole No.: T4BA	Ground Elevation: NA
Total Depth: 12' Aig Ty	
Auger Size: NA	_Sample Type: <u>Geoprobe "Megabore 2" QD X 4'</u>
Project: <u>PORTS MPIS</u>	Data Verified By:Date:
DEPTH SAMPLE SAMPLE LITHOLOGY (FEET) TYPE INTY	DESCRIPTION
0 -	
	CL SILTY CLAY: dk brown (7.5YR3/4) soft damp.
	friable, abund humus, root bundles, Topsoil
2 - CS /	
	CL SILTY CLAY: yellowish brown (10YR5/8), very
/ \-	stiff, dry, common Fe nods, some Mn flecks
4-	
	High angle fractures with argillaceous faces @ 5.5, 6.3, 7.5
	2 3.3 , 2.3 , 7.2
6- CS	
	(10)007.40
1 1 1/ \====	CH CLAY: lt yellow brown (10YR5/4), intbdd lt gray silt laminations, stiff, damp, laminar bedding,
	fat, mod plastic, authegenic gypsum crystals beds @ 7.0' and 7.5'
8-	
10 - CS V	Mudstone pebbles @ 10.5'
	·
12-	CH CLAY: Lt yellow brn (2.5Y86/4), damp, very fat
	and plastic, v. stiff, pronounced bed parting
14-	
16 -	
18 -	
1	
20 -	
=	

ornl OAK RIDGE NATIONAL LABORATORY Prepared By: S.A. Sturm Date: 10/24/94 __ Page: <u>1 OF 1</u> __Ground Elevation: NA Hole No.: T4BB Total Depth: 12' Rig Type: ORNL U2CRT Location: <u>Test Cell #4</u> Auger Size: NA Sample Type: Geoprobe "Megabore 2" OD X 4' Project: <u>FORTS MPIS</u> __Data Verified By:___ Date: DEPTH SAMPLE SAMPLE (FEET) TYPE INTV DESCRIPTION LITHOLOGY 0 -CL SILTY CLAY: dk brown (7.5YR3/4) soft, damp, friable, abund humus, root bundles, breaks into stair step planar faces 5 -CS 4 CL SILTY CLAY: yellowish brown (10YR5/8), very stiff, dry, common Fe nods, some Mn flecks High angle fractures with argillaceous faces @ 5.5', 5.9'& 7.2' 5 -CS CH CLAY: It yellow brown (10YR5/4), intbdd lt gray silt laminations, stiff, damp, laminar bedding, fat, mod plastic, authegenic gypsum crystals beds @ 7.0', lost core 7.1' to 8.0' 8

stiff, dry, common Fe nods, some Mn flecks'

High angle fractures with argillaceous faces @ 5.5', 5.9'\$ 7.2'

CH CLAY: It yellow brown (10YR5/4), intbdd it gray silt laminations stiff, damp, laminar bedding, fat, mod plastic, authegenic gypsum crystals beds @ 7.0', lost core 7.1' to 8.0'

High angle fractures 98.3', 8.4', 8.5', 8.6'and

CH CLAY: brn (7.5YR5/4), damp, very fat and plastic with intbdd yellow brn and red laminae, variegated

10- CS

11- CS

CH CLAY: brn (7.5YR5/4), damp, very fat and plastic with intbdd yellow brn and red laminae.

OAK RIDGE NATIONAL LABORATORY Prepared By: S.A. Sturm ____ Date: <u>10/24/94</u> ___Page:<u>_1_0F_1</u> _Ground Elevation: NA Hole No.: T4BC Total Depth: 12' Aig Type: ORNE U2CRT ___Location: <u>Test Cell #4</u> Sample Type: <u>Geoprobe "Megabore 2" OD X 4'</u> Auger Size: NA____ Date: Project: PORTS MPIS Data Verified By:_ DEPTH SAMPLE SAMPLE (FEET) TYPE INTY DESCRIPTION 0 CL SILTY CLAY: dk brown (7.5YR3/4) soft, damp, friable, abund humus, root bundles, Topsoil 2. CS CL SILTY CLAY: yellowish brown (10YR5/8), very stiff, damp, common Fe nods, some are dusky red CL Clay: brn (7.5YR5/6) with greenish gray silt laminae, fat and plastic, damp CS Б CH CLAY: It yellow brown (10YA5/4), intbdd lt gray silt laminations, stiff, damp, laminar bedding, fat, mod plastic, authegenic gypsum crystals beds @ 7.2', 7.4' and 7.6' CH CLAY: brn (7.5YR5/6) fat, plastic, stiff, damp, homogenous 10 cs CH CLAY: brn (7.5YR5/6) damp, mod plastic, stiff, variegated by yellow (10YR7/8) and red laminae 12 -14 15 18 20

)		D.//	D 8+115#	Date: <u>10/23/94</u> Page: <u>1 0F 1</u>
				Ground Elevation: NA
				De: ORNL U2CRT Location: Test Cell #4
				Sample Type: Geoprobe "Megabore 2" OD X 4'
				Data Verified By:Date:
DEPTH S	AMPLE	SAMPLE	LITHOLOGY	DESCRIPTION
(FEET)	TYPE	INTV		
2 -	cs			CL SILTY CLAY: dk brown (7.5YR3/4) soft, damp . friable, abund humus, root bundles, Topsoil
4-				CL SILTY CLAY: yellowish brown (10YR5/8), with lt gray mottle, damp, friable, common Fe staining, becoming dry and hard at 4'
5-	CS			High angle fractures at 6.7', 7.0' and 7.1', latter two intersect and have xtal gypsum infill
8 -				CH CLAY: It yellow brown (10YR5/4), intbdd lt gray silt laminations, stiff, damp, laminar bedding, fat, mod plastic, authegenic gypsum crystals beds @ 7.9
10-	cs			Dendritic Mn oxide staining on silt laminae partings
12-				CH CLAY: brn (7.5YR5/4), damp, very fat and plastic, v. stiff, pronounced bed partings, homogenous
14-				
16 -				
18				

ornl

<u>VI</u>	<u> </u>	<u> </u>	OAK RIDG	E NATIONAL LABORATORY
Prep	ared	By: <u>S</u>	<u>.R. Sturm</u>	Date: 10/23/94 Page: 1 OF 1
1				Ground Elevation: NA
1				pe: ORNL U2CRT Location: Test Cell #4
Auge	r Siz	e: <u>NA</u>		Sample Type: <u>Geoprobe "Megabore 2" OD X 4'</u>
	****		MPIS	Data Verified By: Date:
OEPTH (FEET)	SAMPLE TYPE	SAMPLE INTV	LITHOLOGY	DESCRIPTION
0-		<u> </u>		
		N /		
_		\ /		CL SILTY CLAY: dk brown (7.5YR3/4) soft, damp,
		$ \setminus $		friable, abund humus, root bundles, Topsoil
2-	cs	$ \ \ $		
-		Λ		
_		/ \		CL CILTY CLAY: vellowish boown (10VP5/P) with it
		/ \		CL SILTY CLAY; yellowish brown (10YR5/8), with lt gray mottle, friable, damp, common Fe staining.
4-		<u>/</u>		becomes hard and dry by 4 with abund fe nods
1 4		\		
		\ <i> </i>		. High pools forsture at 6 4'
.		$ \setminus $		High angle fracture at 6.1'
5-	CS	V		CH CLAY It wellow brown (40VDE (4) intended it good
	63	1 /		CH CLAY: It yellow brown (10YR5/4), intbdd lt gray silt laminations, stiff, damp, laminar bedding,
		$ / \rangle$		fat, mod plastic, authegenic gypsum crystals beds @ 7.0
-		// \		
		/ \		LOCT CORT 7 0 0 0 0
8-				LOST CORE 7.0 - 8.0'
		\ /		
-]	$ \cdot $		
		V		
10 -	CS.	$ \ \ \rangle$		
		I/V		
-	1]/ \		
		/ \		
12-	1			CH CLAY: brn (7.5YR5/4), damp, very fat and plastic, v. stiff, pronounced 1mm bed
	ļ		1	partings, homogenous
	1			
14-	1			
	1			
	ļ			
16 -	1			
-	1			
16 -	1	1		
-	1			
20 -	1			
	1	1	1 i	

repa	red	By:_R	.M. Schloss	erDate: <u>11/14/94</u>	
				Ground Elevation: NA	
otal	Dec	th: 1	2' Aig Ty	e: ORNL U2CRT Location: Test Cell #4	
uger	Siz	e: <u>N</u> A		Sample Type: <u>Geoprobe "Megabore 2" OD X 4'</u>	
				Data Verified By:Date:	
PTH	SAMPLE	SAMPLE	LITHOLOGY	DESCRIPTION	
EET)	TYPE	INIV			
0 -					
		1\ /			
\dashv				CL BIBTY CLAY: brown (10YR5/3) soft, damp . friable, CaCo3 in root pores	
		$ \ \ $			
2-	CS	X		CL SILTY CLAY: yellowish brown (10YR5/8), with life staining throughout, roots as above	t.
		$ / \rangle$		To Starring the bagness, vests at the	
+		I/ \setminus			
		/ \			
4 -				CL SILTY CLAY: as above, abund Fe oxides, Mn oxidenter on open pores, and some CaCo3, 4'-8'	des
		1\ /		on open pores, and some edges,	
+		1 /			
		$ \cdot \cdot $			
Б	CS	I X		CH CLAY: yellow brown (7.5YR5/4), some silt and oxide on desicated surfaces, lt gray mottle throughout interval, very firm, plastic, occ	Mn
		1/		throughout interval, very firm, plastic, occ	
-		1/\			
		/ \	\ <u>===</u>	·	
е-					
		1	/====		
4		$ \cdot $ /			
		$ \cdot $			
10-	cs	X		CH CLAY: yellow brn (7.5YR5/4), well defined bed	
				CH CLAY: yellow brn (7.5YR5/4), well defined bed partings @ 10'. 10.2', 10.6', & 10.9', It gra mottle throughout decreasing w/depth, high an	gle
-		$\parallel / \parallel \setminus$		fracture @ 11.5'	
			\ 		
12-		-	<u> </u>		
-					
14-					
_					
16 -	1				
				•	
_	-				
18 -	1				
_	1				
			1	·	

OAK RIDGE NATIONAL LABORATORY Prepared By: R.M. Schlosser Date: 11/14/94 __Ground Elevation: NA Hole No.: T4BG Total Depth: 12' Rig Type: ORNL U2CRT Location: <u>Test Cell #4</u> Auger Size: NA Sample Type: Geoprobe "Megabore 2" OD X 4' Project: <u>PORTS MPIS</u> _Data Verified By:___ DEPTH SAMPLE (FEET) TYPE DESCRIPTION CL SILTY CLAY: brown (10YR5/3) soft, damp . friable, CaCo3 from injection in root pores CL SILTY CLAY: yellowish brown (10YR5/8), with lt Fe staining throughout, soft, friable 2-CS CL SILTY CLAY: as above, about Fe oxides, some red Fe nodules CH CLAY: brown (7.5YR5/4). mottled lt gray Б. CS on desicated surfaces, abund Mn & Fe oxide, some scattered red nodules, lost core 7'- 8' 8 CH CLAY: yellow brn (7.5YR5/4), gypsiferous, some Fe nodules, desication surface @ 10.5'desication cracksw/ lt gray silt infill 10 cs 12-14 16 18-20

Prepared By: B.M. Schlosser Oste: 11/14/94 Page: 1 OF 1 **Note No.: 128	VI.		1	OAK RIDG	SE NATIONAL LABORATORY
Total Depth: 11	Prep	ared	Ву: <u>Р</u>	.M. Schlos	<u>ser</u> Date: <u>11/14/94</u> Page: <u>1 OF 1</u>
Augustic System (1978) Project: FORTS MPIS Bata Verified By: CL SILTY CLAY: Drown (19785/3) soft. damo friable, CaCO3 from injection in root pores CL SILTY CLAY: vellowish brown (19785/8), with it fe staining w/occasional Mn staining & oxides CL SILTY CLAY: vellowish brown w/ it gray mottle grace CaCo3 from injection, occasional Fe nods gray CH CLAY: brown (7.5YR5/4), mottled it gray On desicated surfaces, abund Mn & Fe staining and homogenous at 10' CH CLAY: vellow brn (7.5YR5/4), silt on erosional surface @ 9', 9.5', & 9.8', some silt fragments CH CLAY: yellow brn (7.5YR5/4), silt on erosional surface @ 9', 9.5', & 9.8', some silt fragments	Hole	No .:	<u> T4B</u> H		Ground Elevation: NA
Project: PORTS MPTS Date Verified By: DESCRIPTION CL SILTY CLAY: brown (10YR5/3) soft, damp. friable. CaCo3 from injection in root pores CL SILTY CLAY: yellowish brown (10YR5/8), with it fresteining w/occasional Mn staining & oxides CL SILTY CLAY: yellowish brown w/ it gray mottle trace CaCo3 from injection, occasional Fe nods CH CLAY: brown (7.5YR5/4), mottled it gray on desicated surfaces, abund Mn & Fe staining and homogenous at 10' CH CLAY: yellow brn (7.5YR5/4), silt on erosional surface @ 9', 9.5', & 9.8', some silt fragments	Tota	l Dep	th: <u>1</u>	1' Aig Ty	ype: ORNL U2CRT Location: Test Cell #4
Serving First New Community Colors of the Color of the Co	Augei	- Siz	e: <u>NA</u>		_Sample Type: <u>Geoprobe "Megabore 2" OD X 4'</u>
CL SILTY CLAY: brown (10YR5/3) soft. damp. friable, Cac03 from injection in root pares CL SILTY CLAY: yellowish brown (10YR5/8), with it free staining w/occasional Mn staining & oxides CL SILTY CLAY: yellowish brown w/ it gray mottle frace Cac03 from injection, occasional Fe nods CH CLAY: brown (7.5YR5/4), mottled it gray on desicated surfaces, abund Mn & Fe staining and homogenous at 10' CH CLAY: yellow brn (7.5YR5/4), silt on erosional surface @ 9'. 9.5', & 9.8', some silt fragments	Proje	ect:_	PORTS	MPIS	Data Verified By:Date:
CL SILTY CLAY: brown (10YR5/3) soft, damp, friable, CaCO3 from injection in root pores CL SILTY CLAY: yellowish brown (10YR5/8), with lt fe staining w/occasional Mn staining & oxides CL SILTY CLAY: yellowish brown w/ lt gray mottle trace CaCO3 from injection, occasional Fe nods e 5. CH CLAY: brown (7.5YR5/4), mottled lt gray on desicated surfaces, abund Mn & Fe staining and homogenous at 10. CH CLAY: yellow brn (7.5YR5/4), silt on erosional surface @ 9', 9.5', & 9.8', some silt fragments				LITHOLOGY	DESCRIPTION
CL SILTY CLAY: brown (10YR5/3) soft, damp, friable, CaCo3 from injection in root pores CL SILTY CLAY: yellowish brown (10YR5/8), with lt fe staining w/occasional Mn staining & oxides CL SILTY CLAY: yellowish brown w/ lt gray mottle trace CaCo3 from injection, occasional Fe nods CH CLAY: brown (7.5YR5/4), mottled lt gray on desicated surfaces, abund Mn & Fe staining and homogenous at 10' CH CLAY: yellow brn (7.5YR5/4), silt on erosional surface @ 9', 9.5', & 9.8', some silt fragments					
friable, CaCO3 from injection in root pages CL SILTY CLAY: yellowish brown (10YR5/8), with lt Fe staining w/occasional Mn staining & oxides CL SILTY CLAY: yellowish brown w/ lt gray mottle trage CaCO3 from injection, occasional Fe nods CH CLAY: brown (7.5YR5/4), mottled lt gray on desicated surfaces, abund Mn & Fe staining and homogenous at 10' CH CLAY: yellow brn (7.5YR5/4), silt on erosional surface @ 9', 9.5', & 9.8', some silt fragments	"				
friable, CaCO3 from injection in root pages CL SILTY CLAY: yellowish brown (10YR5/8), with lt Fe staining w/occasional Mn staining & oxides CL SILTY CLAY: yellowish brown w/ lt gray mottle trage CaCO3 from injection, occasional Fe nods CH CLAY: brown (7.5YR5/4), mottled lt gray on desicated surfaces, abund Mn & Fe staining and homogenous at 10' CH CLAY: yellow brn (7.5YR5/4), silt on erosional surface @ 9', 9.5', & 9.8', some silt fragments			\ /		
CL SILTY CLAY: yellowish brown (10YR5/8), with lt Fe staining w/qccasional Mn staining & oxides CL SILTY CLAY: yellowish brown w/ lt gray mottle trace Cacca from injection, occasional Fe nods CH CLAY: brown (7.5YR5/4), mottled lt gray on desicated surfaces, abund Mn & Fe staining and homogenous at 10' CH CLAY: yellow brn (7.5YR5/4), silt on erosional surface @ 9', 9.5', & 9.8', some silt fragments	1 1		$ \setminus / $		CL SILTY CLAY: brown (10YA5/3) soft, damp, friable CaCo3 from injection in root pures
Fe staining w/occesional Mn staining & oxides CL SILTY CLAY: yellowish brown w/ lt gray mottle trace CaCo3 from injection, occasional Fe nods e 5 CH CLAY: brown (7.5YR5/4), mottled lt gray bn desicated surfaces, abund Mn & Fe staining and homogenous at 10' CH CLAY: yellow brn (7.5YR5/4), silt on erosional surface @ 9', 9.5', & 9.8', some silt fragments			\/		·
CL SILTY CLAY: yellowish brown w/ lt gray mottle trace CaCo3 from injection. occasional Fe nods e 5 CH CLAY: brown (7.5YR5/4). mottled lt gray on desicated surfaces, abund Mn & Fe staining and homogenous at 10' CH CLAY: yellow brn (7.5YR5/4), silt on erosional surface @ 9', 9.5', & 9.8', some silt fragments 12- 14- 16- 18-	5-	CS	ΙX		CL SILTY CLAY: yellowish brown (10YR5/8), with lt
trace CaCo3 from injection, occasional Fe nods @ 5. CH CLAY: brown (7.5YR5/4). mottled lt gray on desicated surfaces, abund Mn & Fe staining and homogenous at 10. CH CLAY: yellow brn (7.5YR5/4). silt on erosional surface @ 9', 9.5', & 9.8', some silt fragments 12- 14- 16-			$ / \rangle$		To Staining WydeadSibilai i'w Staining G dwidds
trace CaCo3 from injection, occasional Fe nods @ 5' CH CLAY: brown (7.5YR5/4), mottled lt gray on desicated surfaces, abund Mn & Fe staining and homogenous at 10' CH CLAY: yellow brn (7.5YR5/4), silt on erosional surface @ 9', 9.5', & 9.8', some silt fragments 12- 14- 16- 18-	i d		I/ \setminus		
trace CaCo3 from injection, occasional Fe nods @ 5' CH CLAY: brown (7.5YR5/4), mottled lt gray on desicated surfaces, abund Mn & Fe staining and homogenous at 10' CH CLAY: yellow brn (7.5YR5/4), silt on erosional surface @ 9', 9.5', & 9.8', some silt fragments 12- 14- 16- 18-			// \		
CH CLAY: brown (7.5YR5/4), mottled lt gray on desicated surfaces, abund Mn & Fe staining and homogenous at 10' CH CLAY: yellow brn (7.5YR5/4), silt on erosional surface @ 9', 9.5', & 9.8', some silt fragments 12- 14- 18-	4-		(CL SILTY CLAY: yellowish brown w/ lt gray mottle
on desicated surfaces, abund Mn & Fe staining and homogenous at 10 CH CLAY: yellow brn (7.5YR5/4), silt on erosional surface @ 9', 9.5', & 9.8', some silt fragments			N /		
on desicated surfaces, abund Mn & Fe staining and homogenous at 10 CH CLAY: yellow brn (7.5YR5/4), silt on erosional surface @ 9', 9.5', & 9.8', some silt fragments	-		$ \rangle /$		
on desicated surfaces, abund Mn & Fe staining and homogenous at 10 CH CLAY: yellow brn (7.5YR5/4), silt on erosional surface @ 9', 9.5', & 9.8', some silt fragments			$ \cdot $		
and homogenous at 10' CH CLAY: yellow brn (7.5YR5/4), silt on erosional surface @ 9', 9.5', & 9.8', some silt fragments 12- 14- 18-	6-	cs	X		CH CLAY: brown (7.5YR5/4). mottled lt gray
and homogenous at 10' CH CLAY: yellow brn (7.5YR5/4), silt on erosional surface @ 9', 9.5', & 9.8', some silt fragments 12- 14- 18-			/\		
and homogenous at 10' CH CLAY: yellow brn (7.5YR5/4), silt on erosional surface @ 9', 9.5', & 9.8', some silt fragments 12- 14- 18-	-				
CH CLAY: yellow brn (7.5YR5/4). silt on erosional surface @ 9', 9.5'. & 9.8', some silt fragments			// \		on desicated surfaces, abund Mn & Fe staining
14-	8-		<u> </u>		and homogenous at 10'
14-			N: /		
14-	-]\./		
14-			$ \cdot \rangle /$		
14-	10 -	cs	\ \ \		CH CLAY: yellow brn (7.5YR5/4), silt on erosional
14- 16- 18-			$ \ \ $		surface @ 9', 9.5', & 9.8', some silt fragments
14- 16- 18-	4		/		
14- 16- 18-			// \		·
18-	12-			<u>\</u>	
18-					
18-					
18-					
18-	14-				
18-					
18-			-		
18-					
18-	15				
	10				
	1				
20-	18 -				•
20-					
20-					
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OAK RIDGE NATIONAL LABORATORY __Page:<u>1 0F 1</u> Prepared By: R.M. Schlosser ___ Date:<u>11/14/94</u> Ground Elevation: NA Hole No.: T4BI Total Depth: 12' Rig Type: ORNL U2CRT ____Location: Test Cell #4 Auger Size: NA Sample Type: Geoprobe "Megabore 2" OD X 4" Project: <u>PORTS MPIS</u> _Data Verified By:___ Date: DEPTH SAMPLE SAMPLE (FEET) TYPE INTY DESCRIPTION LITHOLOGY CL SILTY CLAY: brown (10YR5/3) well developed root mass, some CaCo3 from injection around roots, soft, friable CL SILTY CLAY: yellowish brown (10YR5/8), with lt Fe staining, some CaCo3 on bedding surfaces, soft, friable, vertical micro-fracs w/ CaCo3 CS CL SILTY CLAY: yellowish brown . CaCo3along small vertical fracture planes, high angle fracture @ 5', 3 - 5mm angular Fe stained sandstone liths becoming firm, scattered 1 - 2mm Fe? nods cs CH CLAY: brown (7.5YR5/4), mottled lt gray 7.5YRN7/ firm, slightly plastic, gypsiferous zones @ 7.2 $^{\circ}$ 8 gypsum filled vugs @ 11.5', 2 - 5mm thick CH CLAY: yellow brn (7.5YR5/4), mottled bands @ 9.5 10 -CS CH CLAY: yellow brown, very firm, fat & plastic, waxy appearance. 12 14 16 18 20

OAK RIDGE NATIONAL LABORATORY Prepared By: R.M. Schlosser Date: 11/14/94 ___Page:<u>__1_0F_1</u> Ground Elevation: NA Hole No.: T4BJ Total Depth: 12' Aig Type: ORNL U2CAT Location: <u>Test Cell #4</u> Auger Size: NA Sample Type: Geoprobe "Megabore 2" OD X 4' Project: <u>PORTS MPIS</u> ___Data Verified By:__ Date: DEPTH SAMPLE (FEET) TYPE DESCRIPTION TYPE INTY CL SILTY CLAY: brown (10YR5/3) well developed root mass, some CaCo3 from injection around roots, soft, friable CL SILTY CLAY: yellowish brown (10YR5/8), abund Fe staining, abund CaCo3 along bedding fractures occasional Mn staining cs CL SILTY CLAY: yellowish brown , CaCo3 along horiz bedding planes, some CaCo3 along remnant & root vesicles, strong Mn staining @ 5 CS CH CLAY: brown (7.5YR5/4), mottled lt gray 7.5YRN7/ high angle fracture @ 6', bedding fracture @ 7' very firm, plastic, occasional gypsiferous zones waxy appearance, lt gray bed at 9', 3 - 4mm silt stone bed @ 9.5', vug w/ gypsum @ 9.8' 10 CS $\ensuremath{\mathsf{CH}}$ CLAY: yellow brown, less mottling, dense, fat waxy appearance. 12 14 16 -18 -

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Dr	n _	L	OAK RIDGE	NATIONAL LABORATORY
Prep	ared	By:_S	A Sturm	Date: <u>12/13/94</u> Page: <u>1 OF 1</u>
				Ground Elevation: NA
Tota	l Dep	th: <u>1</u> 3	2' Rig Typ	oe: ORNL U2CRT Location: Test Cell #4
Auge	r Siz	e: <u>NA</u>		Sample Type: Geoprobe "Megabore 2" OD X 4'
			MPIS	Data Verified By:Date:
DEPTH (FEET)	SAMPLE TYPE	SAMPLE INTV	LITHOLOGY	DESCRIPTION
0 -				
-		\ /		
		$ \setminus $		CL SILTY CLAY: dk brown (10YR5/3) soft, damp ,
		$ \cdot $		abund humus, root bundles, Topsoil
2-	cs	V		CL SILTY CLAY: brownish yellow (10YR6/6). friable
_		$ \ \ \rangle$		damp, it gray mottle, some Mn uxides
	1			sandstone pebbles, bed parting at 4.5' no infill
		// \		
. 4-	1			
		1		
		\ /		
İ		$ \setminus $		
Б.	cs	Y		
		$ \ \ \rangle$		
	-	I/ \setminus		CH CLAY: brown (7.5YR5/6) , stiff, damp, very fat
		// \		and plastic, plastic texture, w/intbdd gray silt laminae, gypsum crystals at 6.5 & 7 mudstone pebbles at 9.8
8	4			. mudstone pebbles at 9.8'
		1		
	-	$1 $ \ 1		
		$ \cdot \rangle /$		
10	cs) X		
		1/		
	-	$1/ \setminus$		CH CLAY: becoming red brown
		// \		
12	4	 	+===	
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				Date: <u>12/13/94</u> Page: <u>1 OF 1</u>
ole !	No.:	T4BL		Ground Elevation: NA
otal	Dep	th: <u>1</u>	2' Aig Ty	pe: ORNL U2CRT Location: Test Cell #4
uger	Siz	e: <u>NA</u>		Sample Type: <u>Geoprobe "Megabore 2" OD X 4'</u>
			MPIS	
DEPTH S		SAMPLE INTV	LITHOLOGY	DESCRIPTION
(FEE1)	ITPE	1014	-	
0 -		\ 		
1	÷	\ /		
4		$ \setminus / $		CL SILTY CLAY: dk brown (10YR5/3) soft, damp .
		$ \setminus $	****	abund humus, root bundles, Topsoil
s -	cs	l X		CL SILTY CLAY: brownish yellow (10YR6/6), friable
		$ \ / \rangle$		damp, it gray mottle, some Mn oxides Fe staining common 1'– 5', scattered oxidized sandstone pebbles, bed parting at 3.2' with
				sandstone pebbles, bed parting at 3.2° with
7		/ \		lime precipitates
		/ \		
4-		\		
		\ /		
4		$ \cdot $		
		$ \setminus $		
6-	CS	ΙX		
		/\		
		/ \		CH CLAY: brown (7.5YA5/6) , stiff, damp, very fat
1		// \		and plastic plastic texture w/intbdd
		/ \		gray silt laminae, gypsum crystals at 7.8', low angle fracture at 6.2'- no infill
8-				
		\ /	====	CH CLAY: becoming red brown
4		$ \setminus /$		
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10 -	cs	X		
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	# # _	D C	D STUDE N	ATIONAL LABORATORY
			.H. Sturm	Ground Elevation: NA
) I e	NO .:	136A	O' Dia Topo:	ORNL U2CRT Location: Test Cell #5
ita.	. c:-	LN: <u>1</u>	ear Hig Type.	mple Type: Geoprobe "Megabore 2" OD X 4'
			MPIS	Data Verified By:Date:
РТН	SAMPLE	SAMPLE	LITHOLOGY	DESCRIPTION
EET)	TYPE	INTV	ET INCLUS	
0-				
.		\ /		
-		$ \setminus / $		CL SILTY CLAY: dk brown (7.5YR3/4) soft, damp . friable, abund humus, root bundles, Topsoil
		$ \cdot \cdot $		Trianie, abbita Hamas, 1886 Saliessa, 1994
2 -	cs	X		
		$ \ / \ $		
4		I/V		CL SILTY CLAY: yellowish brown (10YR5/8), with lt gray mottle, friable, damp, common Fe staining,
		/ / \		becomes hard and dry by 3'with abund Fe nods
4-		<u> </u>		
		1		
-		$ \rangle /$		High angle fracture at 5.2'and 6.3'
		$ \cdot $		
6 -	cs	X		CH CLAY: It yellow brown (10YR5/4), intbdd pale olive silt laminations, stiff, damp, very
		$ / \rangle$		fat, mod plastic, parallel discontinous bedding,
-	ł	/		
		\/ \		
8 -		<u> </u>	1	LOST CORE 7.0 - 8.0'
		\		
	-	$ \cdot $		•
		$ \cdot \cdot $		
10	cs	\		CH CLAY: as above with dendritric Mn oxide on bed
		1/		partings .
	-	-/- \		
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	4			
14	-	'		
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repared		. Sturm	NATIONAL LABORATORY Date: 10/22/94 Page: 1 OF 1	
ole No	T588		Ground Elevation: NA	
			De: OANL U2CAT Location: Test Cell #5	
			Sample Type: Geoprobe "Megabore 2" OD X 4'	
	PORTS M		Data Verified By:Date:	
PTH SAMPLE	SAMPLE	LETHOLOGY	DESCRIPTION	
ET) TYPE	INTV		2233777730	
0 -				
	\			
+	1 / [5]		CL SILTY CLAY: dk brown (7.5YR3/4) soft, damp, friable, abund humus, root bundles. Topsoil	
			miable, abund numus, root bungles, ippsoil	
2 - cs				
	/ \			
4.	1/ / 麗		CL SILTY CLAY: yellowish brown (10YR5/8), friab	le
	// \ 陸		blocky structure, abund Fe nods <1mm, become dry and hard by 4′	5
4-			,	
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	\			
€ cs			CH CLAY: It yellow brown (10YR5/8), intbdd lt d	rav
			CH CLAY: It yellow brown (10YR5/8), intbdd lt g silt laminations, stiff, damp, laminar beddi fat, mod plastic, occasional greenish gray	ng,
4	1/\E		(5G5/1) silt laminae w/ dendritic Mn oxide	
	/ \F			
8			LOST CORE 7.0 - 8.0	
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o d cs				
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1	/ \E			
_	N F	===	CIL CLAVA BOT /2 EVDE / 4	
5-			CH CLAY: brn (7.5YR5/4), damp, very fat and plastic, v. stiff, pronounced 1mm bed	
			partings, homogenous	
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rn			NATIONAL LABORATORYDate: 10/22/94 Page: 1 0F 1
		S.A. Sturm	The second secon
	.: <u>T58</u>		Ground Elevation: NA : ORNL U2CRTLocation: Test Cell #5
otal D	epth:_	<u>12'</u> Rig Type	
			ample Type: <u>Geoprobe "Megabore 2" OD X 4'</u>
		S MPIS	Data Verified By:Date:
EPTH SAMPL			DESCRIPTION
0 -			
	Λ	A	
	1	/	CL SILTY CLAY: dk brown (7.5YR3/4) soft, damp.
1	$-1 \setminus I$		friable, abund humus. root bundles, Topsoil
	. I V		
5 - C	s \		
	/ \	\	CL SILTY CLAY: yellowish brown (10YR5/8), increased
1	- 1/	\	clay becomes hard and dry by 4 with about he not
	V	\ 	up to 2mm
4-			
	\	/ ::::	
+		/	
6- 0	s X		CH CLAY: It yellow brown (10YR5/4), intbdd lt gray silt laminations, stiff, damp, paralell laminar
	/\	\ ====	bedding, fat, plastic
4		\ <u>==</u>	
	/		
в-	<u> </u>	+==	LOST CORE 7.0 - 8.0'
	1		
1	\	/ ===	
10 -	s X		
	- /		
		\	
	1/	\E==	
12-	<u> </u>		CH CLAY: brn (7.5YR5/4), with lt gray silt laminae
			easily split at bed partings, stiff, damp, uneven parallel beds
			and voir por aller book
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147			
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Orn I OAK RIDGE NATIONAL LABORATORY

Prepared By: S.R. Sturm Date: 10/22/94 Page: 1 OF 1

Hole No : ISBD Ground Elevation: NA

_Ground Elevation:_NA Hole No.: T58D Total Depth: 12' Rig Type: ORNL U2CRT Location: Test Cell #5 Auger Size: NA Sample Type: Geoprobe "Megabore 2" OD X 4' Project: <u>PORTS MPIS</u> ____Oata Verified By:___ Date: DEPTH SAMPLE SAMPLE (FEET) TYPE INTV DESCRIPTION LITHOLOGY CL SILTY CLAY: dk brown (7.5YR3/4) soft, damp, friable, abund humus, root bundles, Topsoil CS CL SILTY CLAY: yellowish brown (10YR5/8), friable but stiff, damp, common Fe nodules < 1mm becomes hard and dry by 3' High angle fracture w/ clay skins @ 5.8' CH CLAY: It yellow brown (10YR5/8), intbdd It gray silt laminations, stiff, damp, laminar bedding, fat, mod plastic Б CS LOST CORE 7.0 - 8.0' CH CLAY: yellow brn (10YR5/4), damp, very fat and plastic, w/ intbdd dk brn clay and green gray silt laminae, lt gray mudstone pebbles in a dessicated zone at 11.9' 10 -CS 12 -15 -18 -

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repa	ared		.B. Sturm	E NATIONAL LABORATORYOate: 10/21/94 Page: 1 OF 1
				Ground Elevation: NA
ntai	1 Den	th: 13	e' Bia Tv	pe: ORNL U2CRT Location; Test Cell #5
uga.	1 BCp	a. NA		Sample Type: Geoprobe "Megabore 2" OD X 4'
			MPIS	Data Verified By:Date:
	SAMPLE	SAMPLE	LITHOLOGY	DESCRIPTION
(FEET)	TYPE	INTV	LITHOLOGY	
0-	cs			CL SILTY CLAY: dk brown (7.5YR3/4) soft, damp , friable, abund humus, root bundles, Topsoil
4	•			CL SILTY CLAY: yellowish brown (10YR5/8), with lt gray mottle, friable, damp, common Fe staining, becomes hard and dry by 3 with abund Fe nods
-				High angle fracture at 6.1'and 6.2' CH CLAY: It yellow brown (10YR5/4), intbdd pale
6-	CS	$\left \right $		ch CLAY: It yellow brown (10185/4), Income below the colive silt laminations, stiff, damp, very fat, mod plastic, parallel discontinous bedding,
8-	-			LOST CORE 7.6' – 8.0' High angle fracture at 9.0' @ 9.5'
10 -	cs			CH CLAY: as above with increased silt lams and zones of friable silt with dessication fractures
12		//		
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20				

ULTI			E NATIONAL LABORATORY	
			<u>Date: 11/17/94 Page: 1 OF 1</u>	
1			Ground Elevation: NA	
			pe: ORNL U2CRT Location: Test Cell #5	
			Sample Type: <u>Geoprobe "Megabore 2" QD X 4'</u>	
Project:_			Data Verified By:Date:	
(FEET) TYPE	INTV	FITHOLOGY	DESCRIPTION	
0 -	\	,		
	Λ /			
-	$ \cdot $		CL SILTY CLAY: brown (10YR5/3), some purple stain- ing from KMNO4 in upper 8", from surface pen-	
	$ \ \ $		etration, friable.	
2 - CS	X			
	$ / \rangle$		CL SILTY CLAY; as above. KMNO4 staing @ 3°, 4.5°	
	/ \		and at 4 7	
4-	<u> </u>		CL SILTY CLAY: Yellowish brown. (10YR6/8). mottled slightly light gray, friable. Fe staining	
-].	1		throughout.	
	1 \ 1			
	$ \cdot \cdot $			
5 - CS	1 X		LOST 4.7'-8.0'.	
	+/			
	1/\		·	
	/ \	\	at all we well evide because most look light apply plans	
8-			CL CLAY: yellowish brown mottled light gray along bedding fraxtures at 9.2, 9.4, and 9.5, firm.	
	\ /		stiff. waxy apperance, becoming more homogeneous less reduction at 10.0'.	
10 - CS	\			
	$ \ \ \rangle$			
	$ \cdot $			
	// /	VEE E	·	
12-	-			
10			·	
14-				
16 -				
18 -				
20 -				

OAK RIDGE NATIONAL LABORATORY __ Page:<u>1 0F 1</u> Prepared By: R.M. Schlosser ___ Date:__11/17/94 __Ground Elevation: NA Location: <u>Test Cell #5</u> Total Depth: 12' Aig Type: ORNL U2CRT Auger Size: NA Sample Type: Geoprobe "Megabore 2" OD X 4' Project: <u>PORTS MPIS</u> Date: _Data Verified By:_ DEPTH SAMPLE (FEET) TYPE DESCRIPTION INTV 0 -CL SILTY CLAY: brown (10YR5/3), abund roots CL SILTY CLAY: yellowish brown (10YR5/8), very strong KMNO4 along fracture at 3.1, some Fe & Mn staining cs 2 -CL SILTY CLAY: as above, lt KMNO4 staining at 3.5 (staining 1085/4 weak red) some Mn oxides @ 4' CL SILTY CLAY: yellowish brown, strong Fe stained pebbles @ 5.5', abundant Mn oxides 4' - 5.5', pebbly zone @ 5.6' stained w/KMNO4, lt staining @ 5.7', 6.1', & 6.9', strong staining & fracs @ 7.1' & 7.5', less silt becoming clay @ 6.5' CS 6 8 CH CLAY: yellow brn (7.5YR5/4), mottled lt gray. lt gray zone 4 - 6mm wide @ 8.5°, KMNO4 along bedding @ 8.7°, 9°, 10°, 10.2°, bed parting at 10 -CS some weak red staining @ 11.5' 12 -14 16 18 20

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<u> </u>		E NATIONAL LABORATORY
	,	ser Date: <u>11/17/94</u>
	: <u>T58H</u>	Ground Elevation: NA
		ype: ORNL U2CRT Location: Test Cell #5
		Sample Type: Geoprobe "Megabore 2" OD X 4'
Project:_ DEPTH SAMPLE	CHO C	Data Verified By:Date:
(FEET) TYPE	INTV LITHOLOGY	DESCRIPTION
0-	(11)	
	\ /	
		CL SILTY CLAY: brown (10YR5/3), abund roots
		top 6", KMNO4 staining from surface seepage, friable, soft, moist
2 - C5	\ \ \ \ \ \ \ \ \ \ \ \ \ \ \ \ \ \ \	CL SILTY CLAY: vellowish brown (10YR5/8), verv
	\ \ \ \ \ \ \ \ \ \ \ \ \ \ \ \ \ \ \	strong KMNO4 staining 2cm wide at 2.9', lt' staining along fractures @ 3.9'& 4.2', lt gray
		mottle
	// \ 	
4	\ \	CL SILTY CLAY: as above, abundant Mn & Fe oxides
	\ / 	CL SILTY CLAY: as above, abundant Mn & Fe oxides occasional 2 - 3mm pebbles, strong KMNO4 stain in pebbly zones @ 4.8', 5.5', 5.8', & 7.8'
	\ / :: ::::::	In peoply zones e 4.0 , 3.3 , 3.0 , 4 7.0
	\	
5 - CS	V	
	\ \ \ \ \ \ \ \ \ \ \ \ \ \ \ \ \ \ \	
		CH CLAY: yellowish brown, mottled lt gray , lt
	// \ ====	gray reduced zone 3 – 8mm wide @ 8.5 , occasionl roots
8-		roots
	\ /===	
	1\/ ===	
10 - CS		CH CLAY: yellow brn (7.5YR5/4), mottled lt gray,
10 00		heroming darker w/depth. moist. firm. well
	1/\====	defined beds, KMNO4 staining @ 8.7', 8.9', & 9'
12-		
157		
14-		
147		
45		
16 -		
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18 -		
20 -		

OAK RIDGE NATIONAL LABORATORY Page: 1 0F_1 Prepared By: R.M. Schlosser Date: 11/17/94 Ground Elevation: NA Hole No.: T58I Total Depth: 12' Aig Type: ORNL U2CRT _Location: Test Cell #5 Auger Size: NA Sample Type: Geoprobe "Megabore 2" OD X 4' Project: <u>PORTS MPIS</u> Data Verified By: DEPTH SAMPLE SAMPLE (FEET) TYPE INTV DESCRIPTION CL SILTY CLAY: brown (10YR5/3), well developed root mass, some Fe staining, some faint KMNO4 staining in upper 2 CL SILTY CLAY: yellowish brown (10YR5/8), Fe staining and Mn oxides, roots, KMNO4 staining very strong @ 2.8', friable, soft, scattered pebbles @ 3.5', mottled vertically and horzontal w/lt gray silt cs CL SILTY CLAY: as above, abundant pebbles @ 4.2'
Mn & Fe staining and oxides, KMNO4 staining on
fracture @ 3.8'; staining on bedding @ 6.1', 6.3 б cs CH CLAY: yellowish brown lt gray mottle, occasional Fe stained pebbles 5-6 . KMNO4 staining 10R4/4 - 10R5/4 weak red to red 8 gray-orn mudstone on desicated surface. very gypsiferous from 10.2' - 10.5', some crystal development, very well defined beds CH CLAY: yellow brn (7.5YR5/4), KMNO4 staining @ 8.2 10 -CS 12-14 16 18 20

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UL				E NATIONAL LABORATORY
Prep	ared	Ву: <u>Р</u>	.M. Schlos	ser Date: <u>11/17/94</u> Page: <u>1 OF 1</u>
1				Ground Elevation: NA
				pe: ORNL U2CRT Location: Test Cell #5
1				Sample Type: <u>Geoprobe "Megabore 2" OD X 4'</u>
	ect:	PORTS		Data Verified By:Date:
(FEET)		INTV	LITHOLOGY	DESCRIPTION
0 -		ļ		
		\ /		·
-		$ \setminus / $		CL SILTY CLAY: brown (10YR5/3), well developed
	CS	$ \setminus $		root mass, some Fe staining, becoming yellow brn @ 1.5', dark KMNO4 stain at 3', 2.5cm wide, abund pebbles @ 3.5'strongly oxidized, soft to 3'
2 -		ΙX		pebbles @ 3.5'strongly oxidized, soft to 3' becoming firm
		$ / \rangle$		
-				
		/ \		
4-				CL SILTY CLAY: yellowish brown (10YR5/8), Fe staining and Mn oxides, roots, KMNO4 staining
		\ /		very strong @ 5', 6', 6.8', lt staining @ 7.5' & at 8', root development to 7', firm, lt gray
'	cs	$ \cdot $		vertical & horizontal mottling, roots w/reduced area @ 7', blocky structure, KMNO4 staining weak
Б-		V		red
		$ \wedge $		
_		$ / \rangle$		
		/ \		
8-				CH CLAY: yellowish brown, lt gray mottle some mottle
		\ /		CH CLAY: yellowish brown, lt gray mottle some mottle along old root vesicles, dark KMNO4 staining @ 9', 9.2', 9.6', gypsiferous @ 10.2 w/lt gray
-		$ \rangle / $		prown @ 10.4 . becoming more dense w/depth, well
	. cs	$ \cdot \rangle /$		defined bedding , mudstone on desicated surface @ 10', w/gypsum infill in voids, KMNO4 staining weak red. 10A5/4.
10 -		X		
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orn.	OAK RID	GE NATIONAL LABORATORY
Prepared	By: S.A. Sturm	
	: _T58K	Ground Elevation: <u>NA</u>
Total Dep	pth: <u>12'</u> Aig T	ype: <u>ORNL U2CRT</u> Location: <u>Test Cell #5</u>
		Sample Type: <u>Geoprobe "Megabore 2" CD X 4'</u>
	PORTS MPIS	Data Verified By:Date:
DEPTH SAMPLE (FEET) TYPE	SAMPLE LITHOLOGY INTV	DESCRIPTION
0 -		
	/	
		CL SILTY CLAY: dk brown (10YR5/3) soft, damp , abund humus, root bundles, Topsoil
		abund humus, root bundles, (bpsoli
2 CS	\ \ \ \ \ \ \ \ \ \ \ \ \ \ \ \ \ \ \	CL SILTY CLAY: brownish yellow (10YR6/6), friable,
	/	damp. It gray mottle, some Mn oxides Fe staining common 1'- 5', 10-20mm oxidized s.s. pebbles at 4'-5', bed parting at 5' no visible
4	1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1	pebbles at 4'-5', bed parting at 5' no visible alterations
Ì	/ \	
4-		
	/	
4		
5 - CS	\ \ <u></u>	
4		CH CLAY: brown (7.5YR5/6) , stiff, damp, very fat and plastic, plastic texture, w/intbdd
	/ \===	gray silt laminae
8 -		CH CLAY: red brown (2.5YR4/4), v. stiff, plastic, prominent bed partings at 11.4 & 11.7 not
		prominent bed partings at 11.4 & 11.7 not altered
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10 - CS		
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<u>) [</u>	<u>n</u>			E NATIONAL LABORATORY
				Date: 12/13/94 Page: 1 OF 1
				pe: ORNL U2CRT Location: Test Cell #5
				Sample Type: <u>Geoprobe "Megabore 2" OD X 4'</u>
Эгоје Вертн (PORTS		Data Verified By:Date:
(FEET)		INTV	LITHOLOGY	DESCRIPTION
0 -				CL SILT Y CLAY : dk brown (10YR5/3) soft, damp , abund humus, root bundles, Topsoil
2-	cs			CL SILTY CLAY: brownish yellow (10YR6/6), friable, damp, lt gray mottle, some Mn oxides Fe staining common 1'- 5', 10-20mm oxidized s.s. pebbles at 4'-5'
4 -	•			
6-	CS			CH CLAY: brown (7.5YR5/6), stiff, damp, very fat and plastic, w/intbdd gray silt laminae, 45 degree open fracture@ 6.8 - not altered CH CLAY: red brown (2.5YR4/4), v. stiff, plastic, homogenous
8 -				
10 -	cs			
12 -				
14-				
_				
15 -				
18 -				
20 -				

-			t	sorenoie	Summary
ornl	OAK I	RIDGE	NATIONAL	LABORATOR	IY
	20 C A C A C		Acres 10 April 20 Apr	constitution of the experience was although the	Commence of the Control of the Contr

			A. Sturm	Date: 10/21/92 Page: 1 OF 1
				Ground Elevation:NA
				Sample Type: Geoprobe "Megabors 2" OD X 4:
1			MPIS	Data Verified By:Date:
	SAMPLE	SAMPLE INTV	LITHOLOGY	DESCRIPTION
0-				CL SILTY CLAY: dk brown (7.5YR3/4) soft, damp , friable, abund humus, root bundles, Topsoil
4 -				CL SILTY CLAY: yellowish brown (10YR5/8), blocky structure, friable, damp, abundant Fe nodules at 4 to 5
6-	cs			High angle fracture with clay infill at 7 certain the second of the second continues and particles of the second continues and place of the second continues and the second continues are second continues and the second continues are second continues and second continues are second continues and continues are second continues and continues are second continues are second continues and continues are second c
g -	-			
10 -	CS			CH CLAY: yellow brn (10YR5/4), intbdd pale olive silt. as above with high angle fractures at 10.6° @ 11.5°, both with clay infill.
12-				
15 -				
18				
20 -				

UL				E NATIONAL LABORATORY
				Date: 10/21/94 Page: 1 OF 1
1				Ground Elevation: NA
1				pe: ORNL U2CRT Location: Test Cell #6
Auge	r Siz	e: <u>N</u>		Sample Type: <u>Geoprobe "Megabore 2" OD X 4'</u>
1	_		MPIS	Data Verified By:Date:
(FEET)	SAMPLE TYPE	SAMPLE INTV	LITHOLOGY	DESCRIPTION
0 -				
		Λ,		
		$ \setminus $		CL SILTY CLAY: dk brown (7.5YR3/4) soft, damp . friable, abund humus, root bundles, Topsoil
		$ \setminus $		friable, abund humus, root bundles, Topsoil
2-	cs	ΙX		
		$ \ / \ $		
-		$ / \setminus$		CL SILTY CLAY: yellowish brown (10YR5/8), blocky
		ا/		structure, mod stiff, damp, abundant Fe nods' at 3' to 4'
4-				
		\	/	
-		$ \setminus $		High angle fractures with clay infill and Mn oxide at 5', 6.3', 6.5'and 6.8'
		$ \ \ $		
Б-	cs	ΙX		CH CLAY: It yellow brown (10YR5/4), intbdd pale olive silt laminations, stiff, damp, very
		I / I		fat, mod plastic, parallel discontinous bedding,
-	-	I/ \		
		/ '	\ <u></u>	
8 -		\vdash		
		1\ /		
-		$ \setminus $		
		V		
10 -	cs	$ \ \ \rangle$		
		$ / \rangle$		CH CLAV: pod (2 EVD4/R) damo btiff thin laminar
		// /		CH CLAY: red (2.5YR4/8), damp, stiff, thin laminar bedding easily split on bed partings with Mn
12 -		<u></u>		oxide, dendritic staining
-				
			1	
14-				
-	-			
16 -	1			
-	1	1		
18 -	1			
-	1			
20 -	1			

OL	n	L		SE NATIONAL LABORATORY
1				Date: 10/21/94 Page: 1 OF 1
1				
1				ype: ORNL U2CRT Location: Test Cell #6
ł				Sample Type: <u>Geoprobe "Megabore 2" OD X 4'</u>
	ect:_	PORTS	MPIS	Data Verified By:Date:
(FEET)	TYPE	INTV	LITHOLOGY	DESCRIPTION
2-	cs			CL SILTY CLAY: dk brown (7.5YR3/4) soft, damp , friable, abund humus, root bundles, Topsoil
4-	-			CL SILTY CLAY: yellowish brown (10YR5/8), blocky structure, soft, damp, common Fe nods, scattered friable s.stone pebbles
5-	cs			High angle fractures with clay infill at 5.8. CH CLAY: It yellow brown (10YR5/4), intbdd pale olive sit laminations, stiff, damp, very for model laminations.
8-				fat, mod plastic, parallel discontinous bedding,
10	cs			CH CLAY: red (2.5YR4/8), damp, stiff, thin laminar bedding easily split on bed partings with Mn oxide, dendritic staining
12				CH CLAY: It yellow brn (10YR5/4), intbdd pale olive laminae with Mn oxide on bed partings creating tri- color effect, parallel discontinous varves <1 mm, occasional v fine sand laminae
14				
15	4		e sile was differin	
. 18	1			
20	1			

OAK RIDGE NATIONAL LABORATORY ____ Date: <u>10/21/94</u> Page: <u>1 OF 1</u> Hole No.: T6BD Ground Elevation: NA

CL SILTY CLAY: dk brown (7.5YR3/4) soft, damp, friable, abund numus, root bundles, Topsoil CL SILTY CLAY: yellowish brown (10YR6/8), blocky structure, soft, damp, common Fe nods 4' to 6' root pores with clay infill, argillaceous "skins" also apparent on blocky structures CH CLAY: lt yellow brown (10YR5/4), intbdd pale olive silt laminations, stiff, damp, very fat, mod plastic, parallel discontinous bedding.	ugei	- Siz	e: <u>NA</u>		Sample Type: <u>Geoprobe "Megabore 2" OD X 4'</u>
CL SILTY CLAY: dk brown (7.5YR3/4) soft, damp, friable, abund humus, root bundles, Topsoil CL SILTY CLAY: yellowish brown (10YR5/4), blocky structure, soft, damp, common Fe nods 4 to 6 noot pores with clay infill, argillaceous skins also apparent on blocky structures CH CLAY: lt yellow brown (10YR5/4), intbdd pale olive silt laminations, stiff, damp, very fat, mod plastic, parallel discontinous bedding, CH CLAY: red (2.5YR4/8), damp, stiff, thin laminar bedding, rare Mn oxide stains, very pronounced bed parting top of hard red clay, erosional?	noje	ect:_	PORTS	MPIS	Data Verified By:Date:
CL SILTY CLAY: dk brown [7.5YR3/4] soft, damp, friable, abund numus, root bundles, Topsoil CL SILTY CLAY: yellowish brown (10YR6/8), blocky structure, soft, damp, common Fe nods 4 to 6 root pores with clay infill, argillaceous "skins" also apparent on blocky structures CH CLAY: lt yellow brown (10YR5/4), intbdd pale olive silt laminations, stiff, damp, very fat, mod plastic, parallel discontinuous bedding. CH CLAY: red (2.5YR4/8), damp, stiff, thin laminar bedding, nare Mn oxide stains, very pronounced bed parting top of hard red clay, erosional?	EPTH	SAMPLE	SAMPLE	T	
CL SILTY CLAY: dk brown (7.5YR3/4) soft, damp friable, abund humus, root bundles, Topsoil CL SILTY CLAY: yellowish brown (10YR6/8), blocky structure, soft, damp, common Fe nods 4 to 6 root ponce with clay infill argillaries. "skins" also apparent on blocky structures CH CLAY: lt yellow brown (10YR6/4), intbdd pale clive silt laminations, stiff, damp, very fat, mod plastic, parallel discontinous bedding. CH CLAY: red (2.5YR4/8), damp, stiff, thin laminar bedding, rare Mm oxide stains, very pronounced bed parting top of hard red clay, erosional?					
friable, abund numus, root bundles, Topsoil CL SILTY CLAY: yellowish brown (10YRF/8) blocky structure. Soft, damp, common Fe nods 4 to 6 root pores with clay infill, argillaceous "skins' also apparent on blocky structures CH CLAY: lt yellow brown (10YRF/4), intbdd pale clive slit laminations, stiff, damp, very fat, mod plastic, parallel discontinous bedding, and plastic, parallel discontinous bedding, rare Mn oxide stains, very pronounced bed parting top of hard red clay, erosional?	0 -				
friable, abund numus, root bundles, Topsoil CL SILTY CLAY: yellowish brown (10YRF/8) blocky structure. Soft, damp, common Fe nods 4 to 6 root pores with clay infill, argillaceous "skins' also apparent on blocky structures CH CLAY: lt yellow brown (10YRF/4), intbdd pale clive slit laminations, stiff, damp, very fat, mod plastic, parallel discontinous bedding, and plastic, parallel discontinous bedding, rare Mn oxide stains, very pronounced bed parting top of hard red clay, erosional?			1\ /		
CL SILTY CLAY: yellowish brown (10YR6/8), blocky structure, soft, damp, common Fe nods 4' to 6 root pores with clay infill, angilaceous "skins' also apparent on blocky structures 6- cs CH CLAY: lt yellow brown (10YR5/4), intbdd pale olive silt laminations, stiff, damp, very fat, mod plastic, parallel discontinous bedding. CH CLAY: red (2.5YR4/8), damp, stiff, thin laminar bedding, rare Mn oxide stains, very pronounced bed parting top of hard red clay, erosional? CH CLAY: red (2.5YR4/8), damp, stiff, thin laminar bedding, rare Mn oxide stains, very pronounced bed parting top of hard red clay, erosional?	4		\mathbb{N} / \mathbb{N}		CL SILTY CLAY: dk brown (7.5YR3/4) soft damp,
CH CLAY: lt yellow brown (10YR5/4), intbdd pale clive silt laminations, stiff, damp, very fat, mod plastic, parallel discontinous bedding. CH CLAY: red (2.5YR4/8), damp, stiff, thin laminar bedding, rare Mn oxide stains, very pronounced bed parting top of hard red clay, erosional?	İ		$1 \setminus I$		friable, abund humus, root bundles, Topsoil
CH CLAY: lt yellow brown (10YR5/4), intbdd pale clive silt laminations, stiff, damp, very fat, mod plastic, parallel discontinous bedding. CH CLAY: red (2.5YR4/8), damp, stiff, thin laminar bedding, rare Mn oxide stains, very pronounced bed parting top of hard red clay, erosional?	2 -	cs	1. Y		CL STITY CLAY: vellowish brown (10786/8) blocky
CH CLAY: lt yellow brown (10YR5/4), intbdd pale clive silt laminations, stiff, damp, very fat, mod plastic, parallel discontinous bedding. CH CLAY: red (2.5YR4/8), damp, stiff, thin laminar bedding, rare Mn oxide stains, very pronounced bed parting top of hard red clay, erosional?	.		$\perp \wedge$		structure, soft, damp, common Fe nods 4' to 6'
CH CLAY: lt yellow brown (10YR5/4), intbdd pale olive silt laminations, stiff, damp, very fat, mod plastic, parallel discontinous bedding. CH CLAY: red (2.5YR4/8), damp, stiff, thin laminar bedding, rare Mn oxide stains, very pronounced bed parting top of hard red clay, erosional?			/ \		root pores with clay infill, argillaceous "skips" also apparent on blocky structures
CH CLAY: lt yellow brown (10YR5/4), intbdd pale olive silt laminations, stiff, damp, very fat, mod plastic, parallel discontinous bedding. CH CLAY: red (2.5YR4/8), damp, stiff, thin laminar bedding, rare Mn oxide stains, very pronounced bed parting top of hard red clay, erosional?	7]/ \		and dead apparent on blocky attactores
CH CLAY: lt yellow brown (10YR5/4), intbdd pale olive silt laminations, stiff, damp, very fat, mod plastic, parallel discontinous bedding. CH CLAY: red (2.5YR4/8), damp, stiff, thin laminar bedding, rare Mn oxide stains, very pronounced bed parting top of hard red clay, erosional?			// \		
CH CLAY: red (2.5YR4/8), damp, stiff, thin laminar bedding, rare Mn oxide stains, very pronounced bed parting top of hard red clay, erosional?	4-	•			
CH CLAY: red (2.5YR4/8), damp, stiff, thin laminar bedding, rare Mn oxide stains, very pronounced bed parting top of hard red clay, erosional?			1\ /		
CH CLAY: red (2.5YR4/8), damp, stiff, thin laminar bedding, rare Mn oxide stains, very pronounced bed parting top of hard red clay, erosional?	4		$ \setminus /$		
CH CLAY: red (2.5YR4/8), damp, stiff, thin laminar bedding, rare Mn oxide stains, very pronounced bed parting top of hard red clay, erosional?	Ì		$ \setminus $		
CH CLAY: red (2.5YR4/8), damp, stiff, thin laminar bedding, rare Mn oxide stains, very pronounced bed parting top of hard red clay, erosional?	_		V		OH OLAV. 14
fat, mod plastic. parallel discontinous bedding. CH CLAY: red (2.5YR4/8), damp, stiff, thin laminar bedding, rare Mn oxide stains, very pronounced bed parting top of hard red clay, erosional? 12- 14-	٦٦	US			<pre>um uLay: It yellow brown (10YH5/4), intbdd pale</pre>
CH CLAY: red (2.5YR4/8), damp, stiff, thin laminar bedding, rare Mn oxide stains, very pronounced bed parting top of hard red clay, erosional?			$ / \rangle$		fat, mod plastic, parallel discontinous bedding,
CH CLAY: red (2.5YR4/8), damp, stiff, thin laminar bedding, rare Mn oxide stains, very pronounced bed parting top of hard red clay, erosional?	+				
CH CLAY: red (2.5YR4/8), damp, stiff, thin laminar bedding, rare Mn oxide stains, very pronounced bed parting top of hard red clay, erosional?			// /		
CH CLAY: red (2.5YR4/8), damp, stiff, thin laminar bedding, rare Mn oxide stains, very pronounced bed parting top of hard red clay, erosional?	8 -		<u> </u>		
CH CLAY: red (2.5YR4/8), damp, stiff, thin laminar bedding, rare Mn oxide stains, very pronounced bed parting top of hard red clay, erosional?			\ /	====	
CH CLAY: red (2.5YR4/8), damp, stiff, thin laminar bedding, rare Mn oxide stains, very pronounced bed parting top of hard red clay, erosional?			\		
CH CLAY: red (2.5YR4/8), damp, stiff, thin laminar bedding, rare Mn oxide stains, very pronounced bed parting top of hard red clay, erosional?	٦		$ \setminus $		
CH CLAY: red (2.5YR4/8), damp, stiff, thin laminar bedding, rare Mn oxide stains, very pronounced bed parting top of hard red clay, erosional?			$ \ \ $		
bedding, rare Mn oxide stains, very pronounced bed parting top of hard red clay, erosional?	10 -	CS	X ·		
bedding, rare Mn oxide stains, very pronounced bed parting top of hard red clay, erosional?			/\		
bedding, rare Mn oxide stains, very pronounced bed parting top of hard red clay, erosional?	4				CH CLAY: red (2.5YA4/8), damp, stiff, thin laminar
14 -			/ \		bedding, rare Mn oxide stains, very pronounced
16 -	12		/\		bed parting top of hard red tlay, erosional?
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Borehole Summary Information

OAK RIDGE NATIONAL LABORATORY Page: 1 0F 1 __ Date:<u>10/21/94</u> Prepared By: S.R. Sturm __Ground Elevation:__NA Hole No.: TSBE Total Depth: 12' Rig Type: ORNL U2CRT _Location: <u>Test Cell #6</u> Auger Size: NA Sample Type: Geoprobe "Megabore 2" OD X 4' Project: <u>PORTS MPIS</u> _Data Verified By:_ Date: SAMPLE INTV DEPTH SAMPLE (FEET) TYPE DESCRIPTION LITHOLOGY 0 -CL SILTY CLAY: dk brown (7.5YR3/4) soft, damp , friable, abund humus, root bundles, Topsoil CL SILTY CLAY: yellowish brown (10YR6/8), blocky structure, soft, damp, common Fe nods 4' to 5. 2. CS CH CLAY: It yellow brown (10YR5/4), intbdd pale olive silt laminations, stiff, damp, very fat, mod plastic, parallel discontinous bedding, 6 -CS 8 -Saturated, perched water on red clay 10 cs CH CLAY: red (2.5YR4/8), damp, stiff, thin laminar bedding, rare Mn oxide stains, very pronounced bed parting top of hard red clay, erosional ? 12-14 16 -18 20

repared	By: S.A. Sturm	GE NATIONAL LABORATORY Date: 10/24/94 Page: 1 OF 1
ole No.:	T78A	Ground Elevation:_NA
otal De	pth: <u>12'</u> Rig	Type: ORNL U2CRT Location: Test Cell #7
uger Si	ze: <u>NA</u>	Sample Type: <u>Geoprobe</u> "Megabore 2" OD X 4'
	PORTS MPIS	Data Verified By:Date:
DEPTH SAMPLE (FEET) TYPE	SAMPLE LITHOLOGY	DESCRIPTION
0 -		
1		CL SILTY CLAY: dk brown (7.5YR3/4) soft, damp . friable, abund humus, root bundles, Topsoil
2 - CS		CL SILTY CLAY: yellowish brown (10YR5/8), blocky structure, damp, common Fe flecks, mod stiff Fe oxide infill in root pores—limonite
4 -		
-		
6 - CS		CH CLAY: lt yellow brown (10YR5/4), intbdd pale olive silt laminations, stiff, damp, very fat, mod plastic, high angle fractures 6.7', 7.5'
8 -		LOST CORE 7.5' to 8'
10 - CS		
12-		CH CLAY: red (2.5YR4/8), damp, stiff, thin laminar bedding, rare Mn oxide stains, very pronounced bed parting top of hard red clay, erosional ?
4		
14-		
16 -		

Borehole Summary Information

OAK RIDGE NATIONAL LABORATORY Prepared By: R.M. Schlosser ___Date: <u>11/20/94</u> Page: 1 0F 1 _Ground Elevation: <u>NA</u> Hole No.: T7BB Total Depth: 12' Rig Type: ORNL U2CRT Location: Test Cell #7 Sample Type: <u>Geoprobe "Megabore 2" OD X 4'</u> Project: PORTS MPIS Data Verified By:___ Date: DEPTH SAMPLE (FEET) TYPE DESCRIPTION INTV 0 CL SILTY CLAY: brown (10YR5/3) soft, well , developed root bundles, friable CL SILTY CLAY: yellowish brown (10YR5/8), mod stiff, damp, abund Fe staining 2-3, friable heavy Fe stained pebbles @ 4', lt gray mottle becoming more mottled with depth, occ. roots 2 CS CL SILTY CLAY: yellowish brown lt gray mottle, abundant pebbles 5' - 5.8', some Fe nodules from 4.5', MN oxides occasional CH CLAY: dk yellow brown (10YR4/4). mottled lt gray as above, slighty plastic, moist, CS 6 CH CLAY: brn (7.5YR5/2) very firm, plastic, well developed beds w/ly gray clay on bed surfaces, stiff, moist, waxy appearance 10 -CS CH CLAY: yellow brn varieagated w/lt gray grn & lt brownish gray, stiff plastic becoming red brown 5YR4/4 to yellowish red 5YR4/6, stiff, homogenous 12. 16 -18 20

Borehole Summary Information ornl OAK RIDGE NATIONAL LABORATORY Prepared By: R.M. Schlosser ___ Date:<u>11/20/94</u> __ Page: <u>1 OF 1</u> _Ground Elevation:<u>NA</u> Hole No.: T7BC Total Depth: 12' Rig Type: ORNL U2CRT Location: <u>Test Cell #7</u> Auger Size: NA Sample Type: Geoprobe "Megabore 2" OD X 4' Project: <u>PORTS MPIS</u> Data Verified By:__ Date: DEPTH SAMPLE (FEET) TYPE DESCRIPTION 0 CL SILTY CLAY: brown (10YR5/3) soft, well developed soil w/root mass CL SILTY CLAY: yellowish brown (10YR5/8), some Fe oxides, soft, moist, friable, becoming stiff CS CL SILTY CLAY: yellowish brown lt gray mottle, some Fe & MN oxides, some Guar-gum @ 5.5'in fractures, becoming v stiff, moist, blocky structure, Guargum in void @ 6.2' cs CH CLAY: dk yellow brown (10YR4/4), mottled lt gray as above, increasing mottle w/depth, fracture @ 7' CH CLAY: It gray green clay mottled yellow brn LOST CORE 11' - 12' 10 CS 12 -14 15 18

20

Borehole Summary Information

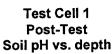
- #				GE NATIONAL LABORATORY Date: 12/14/94 Page: 1 OF 1
		,	S.A. Sturm	and the second of the second o
)	Ground Elevation: NA
				ype: ORNL U2CRT Location: Test Cell #7
				Sample Type: Geoprobe "Megabore 2" OD X 4'
	SAMPLE	PORTS	MPIS	Data Verified By:Date:
	TYPE	INTV	LITHOLOGY	DESCRIPTION
2-	cs			CL SILTY CLAY: dk brown (10YR5/3) soft, damp, abund humus, root bundles, Topsoil CL SILTY CLAY: brownish yellow (10YR6/6), friable, damp, lt gray mottle, open root pores some w/roots Fe staining common 3' - 5', 10-20mm oxidized s.s. pebbles at 4'-5'
4-				
5-	CS			CH CLAY: yellow brown (10YR5/6), stiff, damp, very fat and plastic, w/intbdd gray silt laminae, 45 degree open fracture @ 7.2 shows Fe staining and recent hydration of surrounding clay
8 -				
10 -	cs			CH CLAY: red brown (2.5YR4/4), v. stiff, plastic, intbdd lt gray silt laminae, numerous bed partings, no visible alterations
12-		_		
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epared	By:_S.R. Śtur	DGE NATIONAL LABORATORY
		Type: ORNL U2CAT Location: Test Cell #7
		Sample Type: Geophobe "Medabore 2" OD X 4'
	PORIS MPIS	
P7H SAMELE	SAMPLE LITHDLOGY	Data Verified By:Date:
EET) TYPE	INTY ETHOCOS	DESCRIPTION
0 -		
	\	
4		CL SILTY CLAY: dk brown (10YR5/3) soft, damp.
cs		abund humus, root bundles, Topsoil
2-	\	CL SILTY CLAY: brownish yellow (10YR6/6), friable,
		damp, lt gray mottle, Fe staining common, high angle fracture 1' to 3' with Guar gum infill,
	/ \	m angle fracture 1 to 3 with Guar gum infill, comparate 5'-6'
	/ \	
4	V V::	
	A	
.]	\	
cs		
_	/ V	
5 -		CH CLAY: yellow brown (10YR5/6), stiff, damp, very fat and plastic, w/intbdd gray silt laminae, 3
`		high angle / vertical fractures @ 6.1 6.3 and at 7, each with Guar gum infill
1		T ac / , each with oder gom in iii
	/ \===	-
8		
	\	
cs		
(3)		
0-		CH CLAY: red brown (2.5YR4/4), v. stiff, plastic,
		CH CLAY: red brown (2.5YR4/4), v. stiff, plastic, intbdd lt gray silt laminae, numerous bed partings 9'– 11', no visible alterations
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APPENDIX C

DATA TREND GRAPHS FOR VARIOUS GEOCHEMICAL PARAMETERS IN SOIL AND SOIL-PORE WATER

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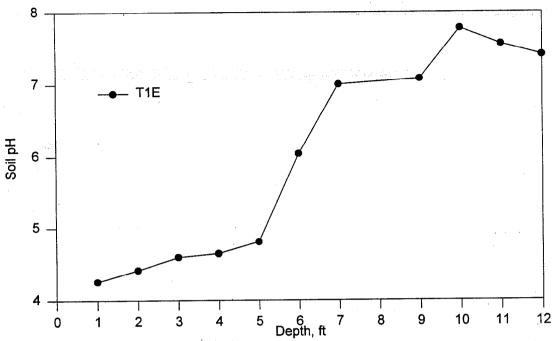


Fig. C.1. Test cell 1 - post-treatment soil pH.

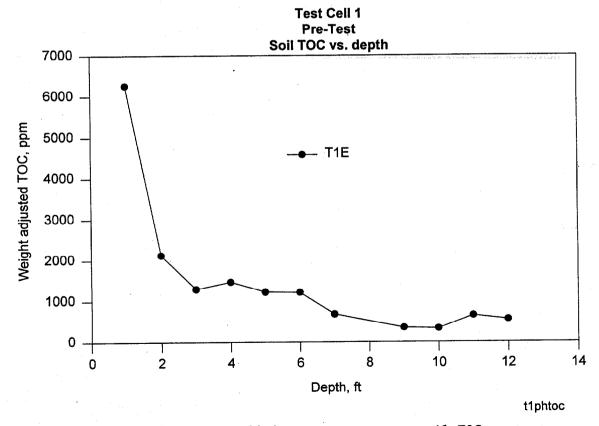


Fig. C.2. Test cell 1 - pre-treatment soil TOC content.



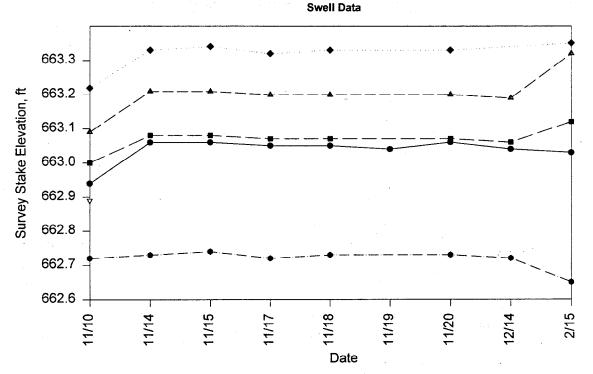


Fig. C.3. Test cell 1 - post-treatment soil swell data. Test Cell 1

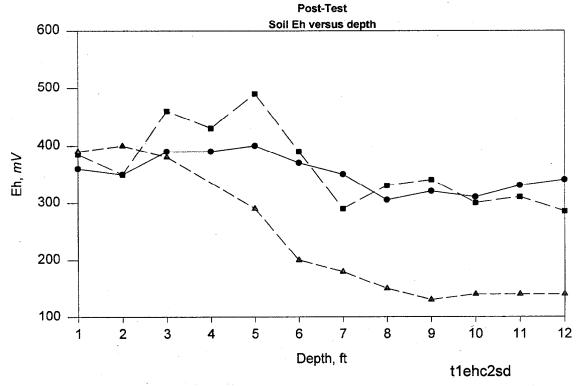
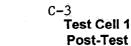


Fig. C.4. Test cell 1 - post-treatment soil Eh.



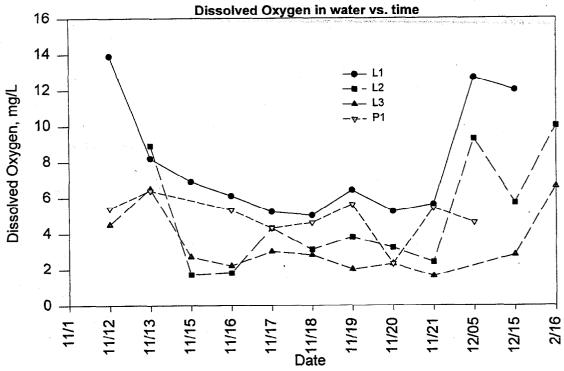


Fig. C.5. Test cell 1 - post-treatment water DO levels.

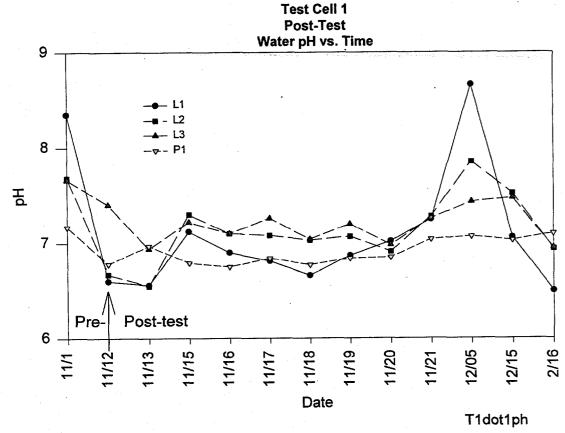


Fig. C.6. Test cell 1 - post-treatment water pH.

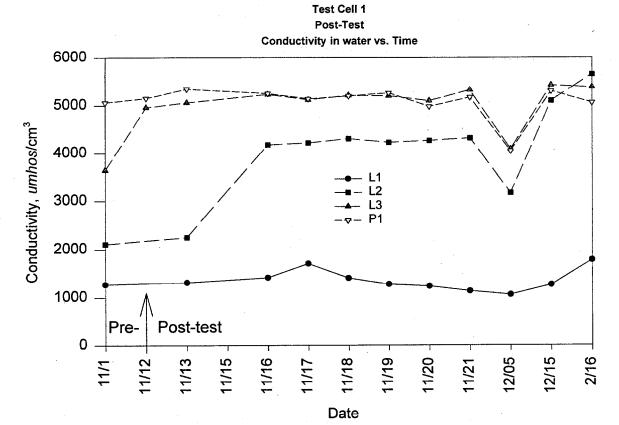


Fig. C.7. Test cell 1 - post-treatment water conductivity levels.

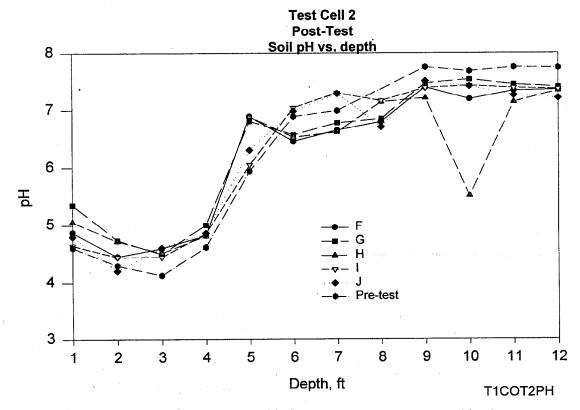


Fig. C.8. Test cell 2 - post-treatment soil pH.

Test Cell 2

Post-Test Nitrate vs. depth 3 -Ġ 2 NO₃, ppm 1 0 2 3 4 5 6 7 8 9 10 11 12 Depth, ft

Fig. C.9. Test cell 2 - post-treatment soil nitrate levels.

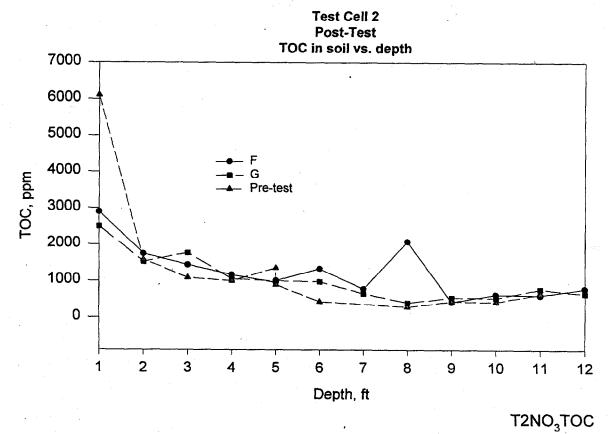


Fig. C.10. Test cell 2 - post-treatment soil TOC content.

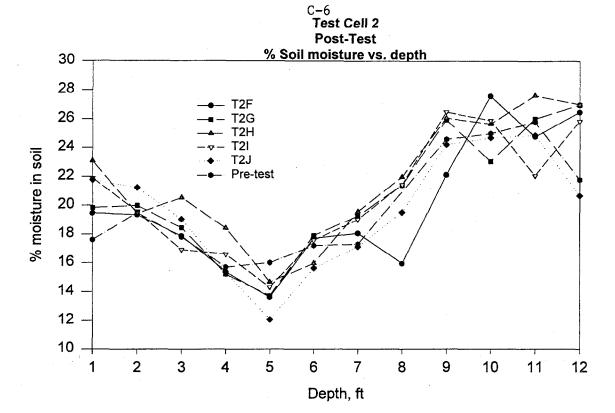


Fig. C.11. Test cell 2 - post-treatment soil moisture content.

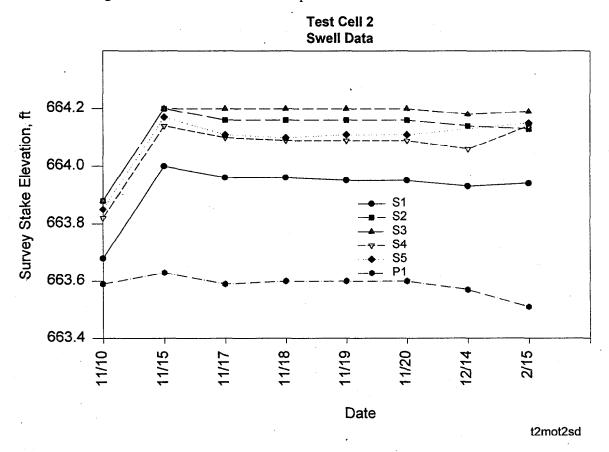


Fig. C.12. Test cell 2 - post-treatment soil swell data.

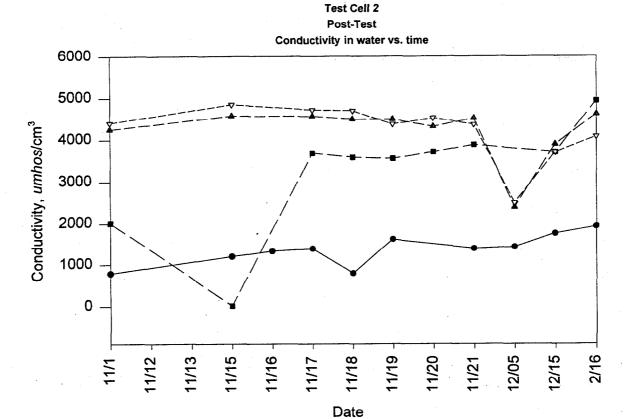


Fig. C.13. Test cell 2 - post-treatment water conductitivity levels.

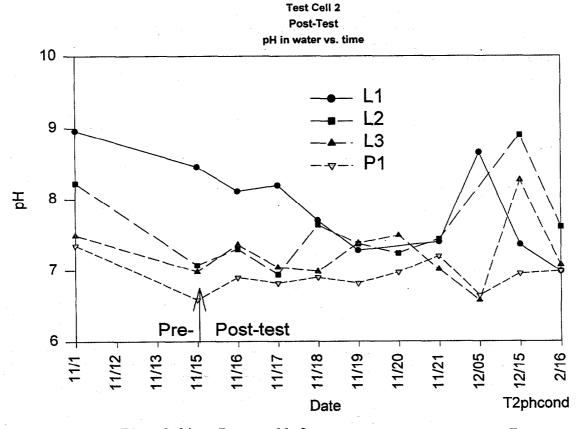


Fig. C.14. Test cell 2 - post-treatment water pH.

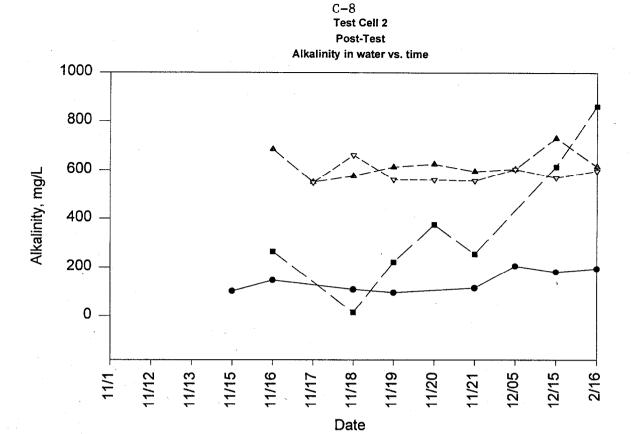


Fig. C.15. Test cell 2 - post-treatment water alkalinity levels.

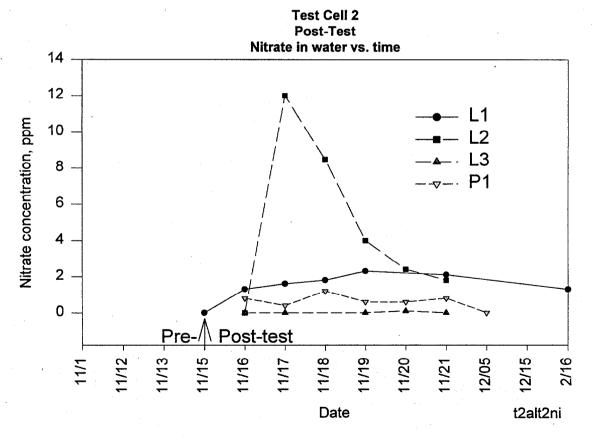


Fig. C.16. Test cell 2 - post-treatment water nitrate levels.

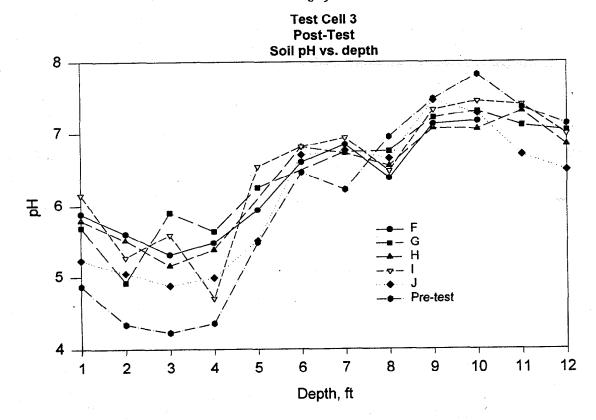


Fig. C.17. Test cell 3 - post-treatment soil pH.

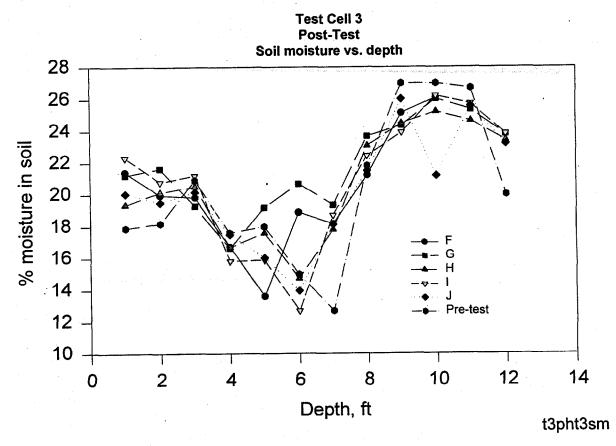


Fig. C.18. Test cell 3 - post-treatment soil moisture levels.



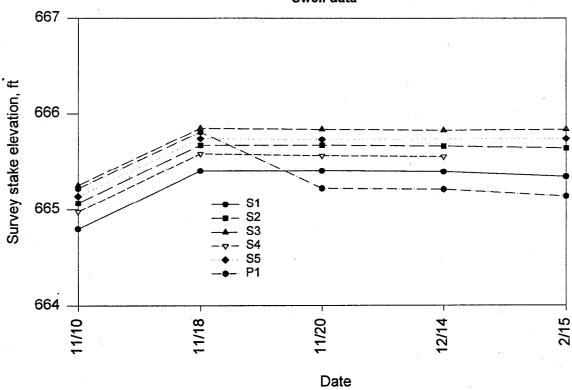


Fig. C.19. Test cell 3 - post-treatment soil swell data.

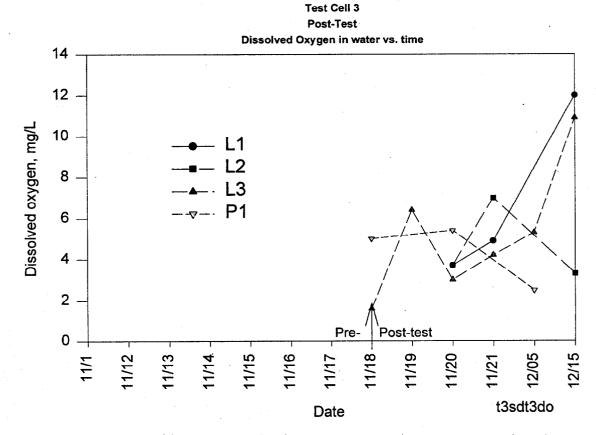


Fig. C.20. Test cell 3 - post-treatment water DO levels.

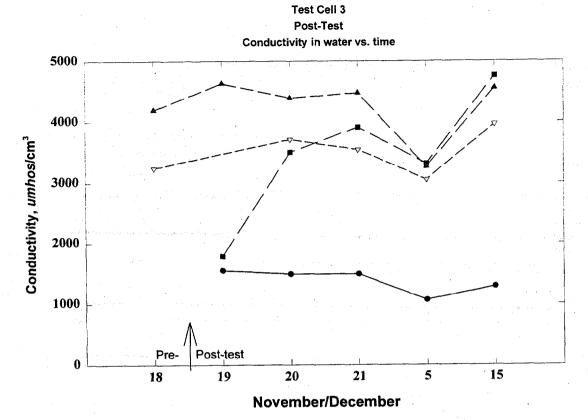


Fig. C.21. Test cell 3 - post-treatment water conductivity levels.

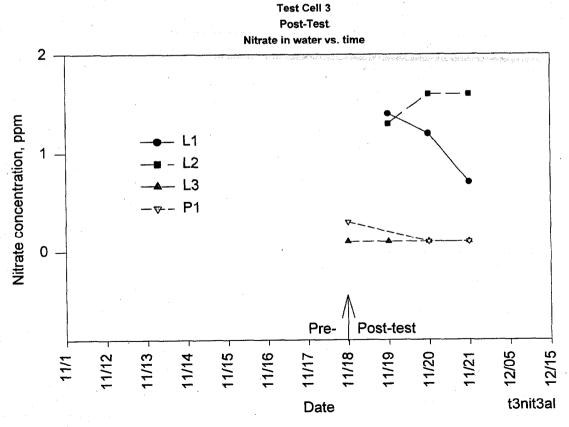
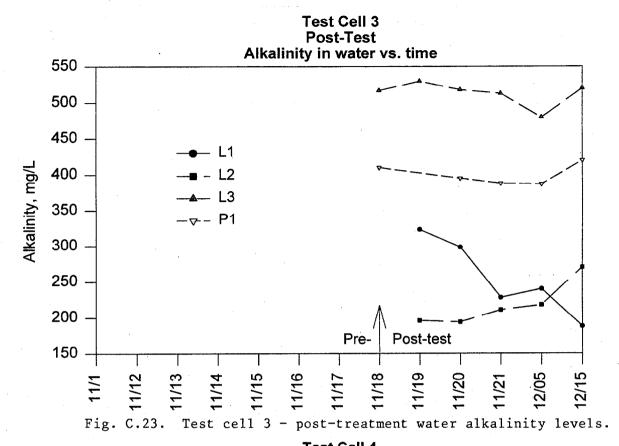


Fig. C.22. Test cell 3 - post-treatment water nitrate levels.



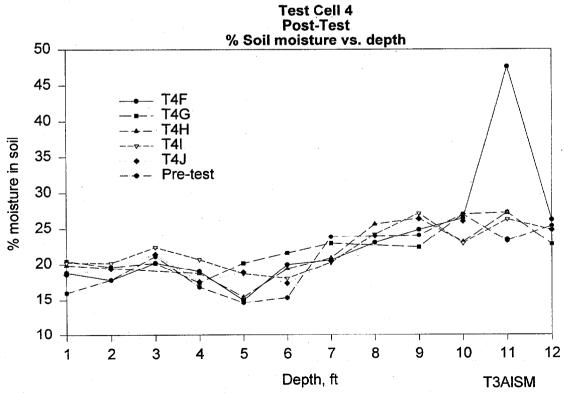


Fig. C.24. Test cell 4 - post-treatment soil moisture levels.



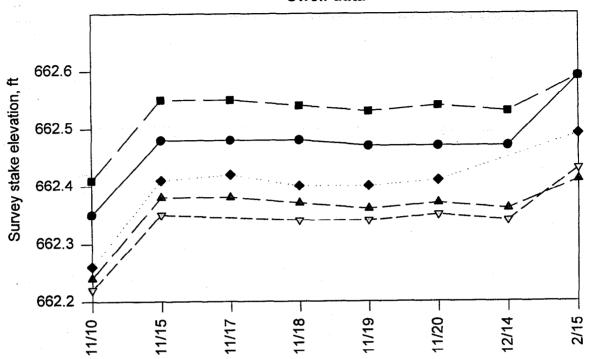


Fig. C.25. Test cell 4 - post-treatment soil swell data.

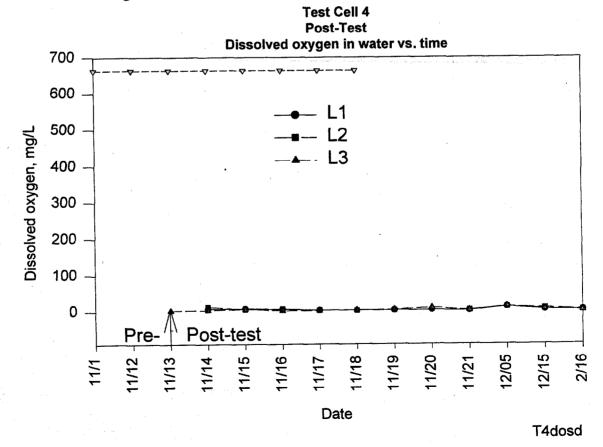


Fig. C.26. Test cell 4 - post-treatment water DO levels.

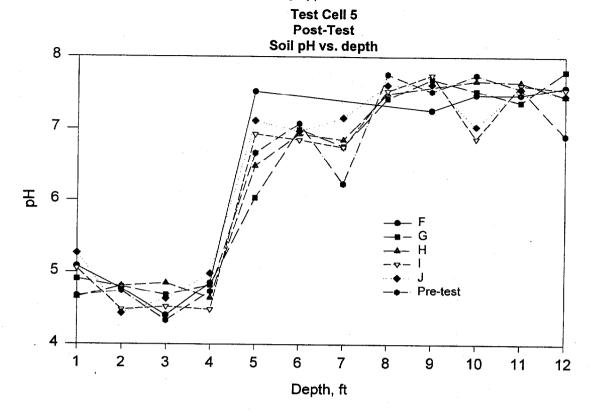


Fig. C.27. Test cell 5 - post-treatment soil pH.

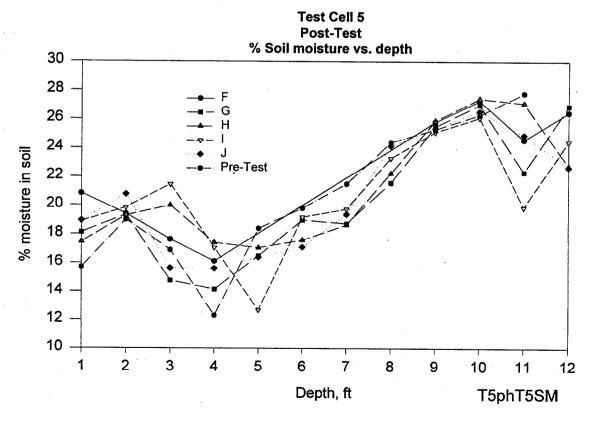


Fig. C.28. Test cell 5 - post-treatment soil moisture levels.

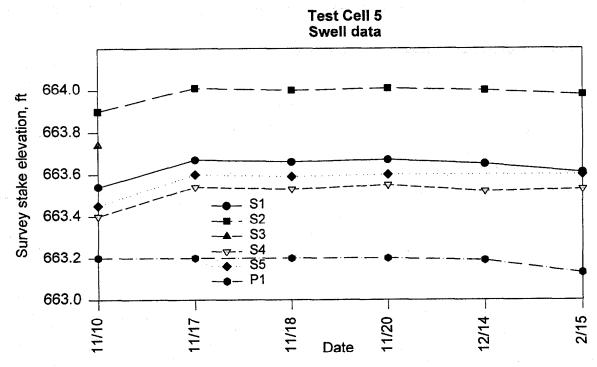


Fig. C.29. Test cell 5 - post-treatment soil swell data.

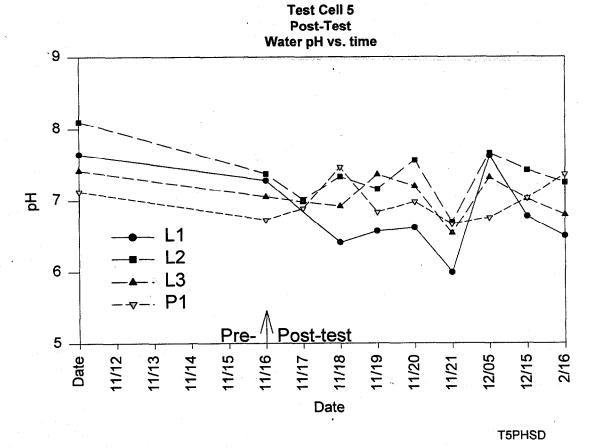


Fig. C.30. Test cell 5 - post-treatment water pH.

Test Cell 5 Post-Test Conductivity in water vs. time

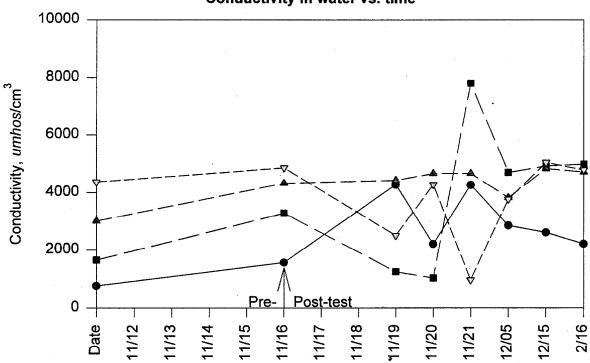


Fig. C.31. Test cell 5 - post-treatment water conductivity levels.

Test Cell 5

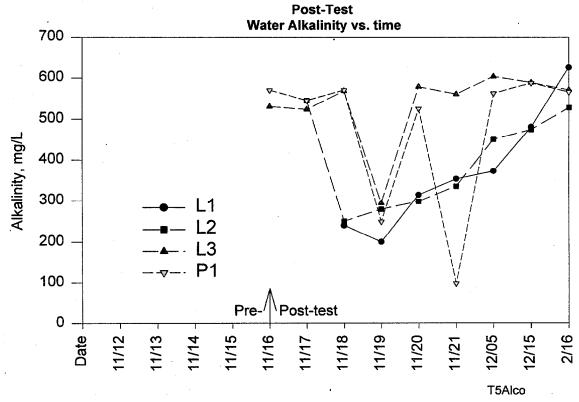


Fig. C.32. Test cell 5 - post-treatment water alkalinity levels.

Test Cell 6 Swell data

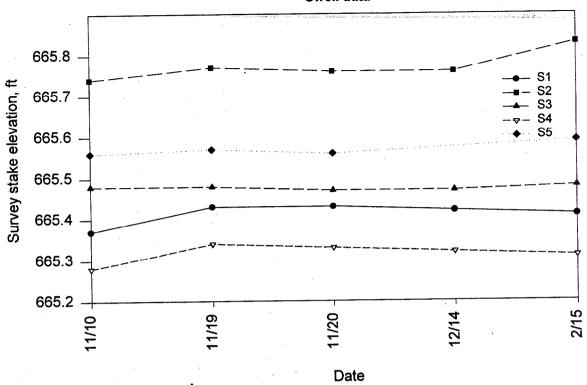


Fig. C.33. Test cell 6 - post-treatment soil swell data.

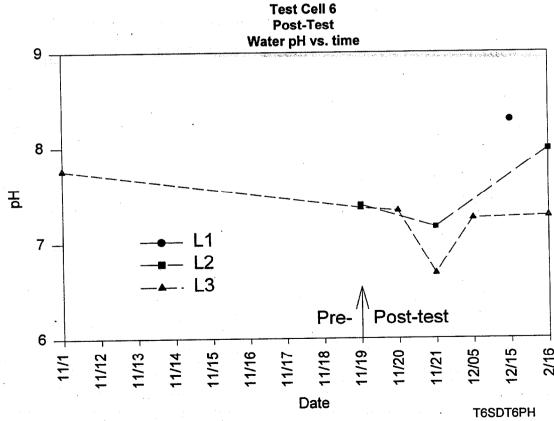


Fig. C.34. Test cell 6 - post-treatment water pH levels.

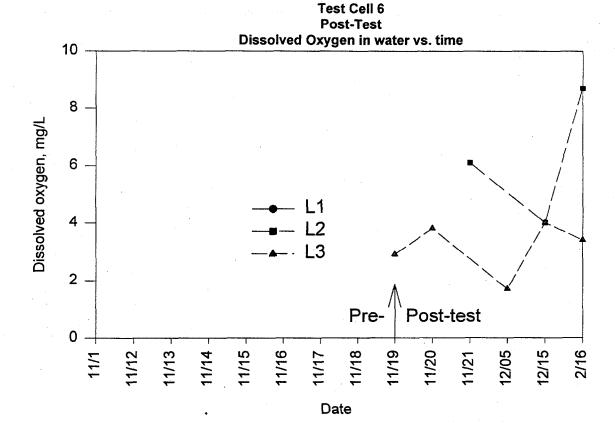


Fig. C.35. Test cell 6 - post-treatment water DO levels.

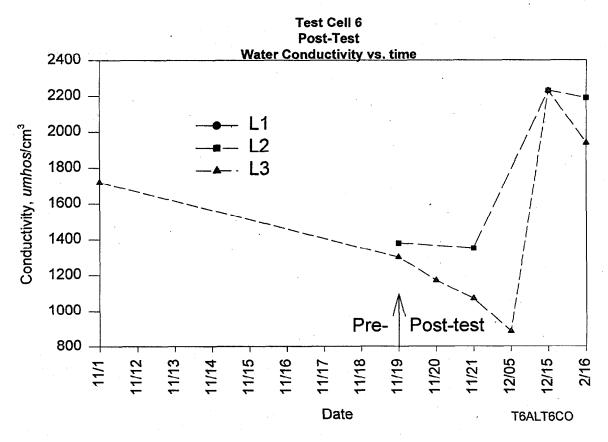


Fig. C.36. Test cell 6 - post-treatment water conductivity levels.

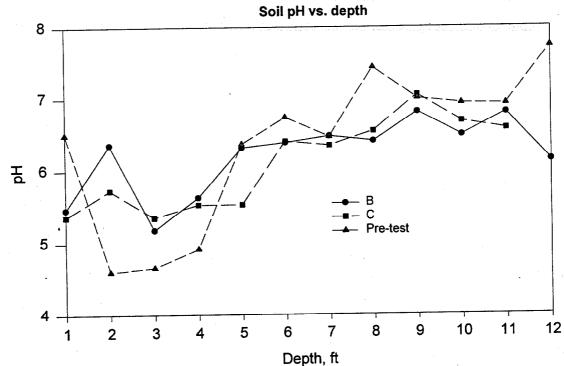


Fig. C.37. Test cell 7 - post-treatment soil pH.

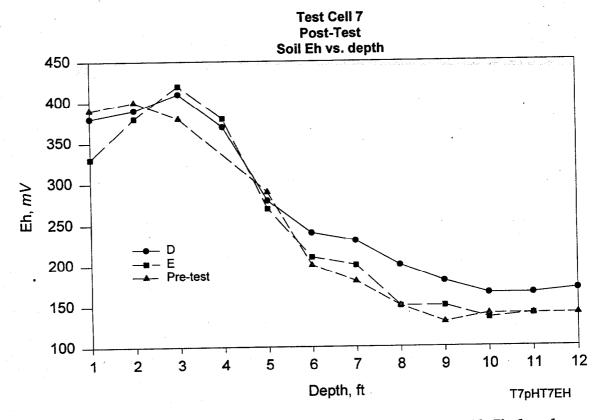
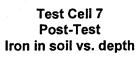


Fig. C.38. Test cell 7 - post-treatment soil Eh levels.



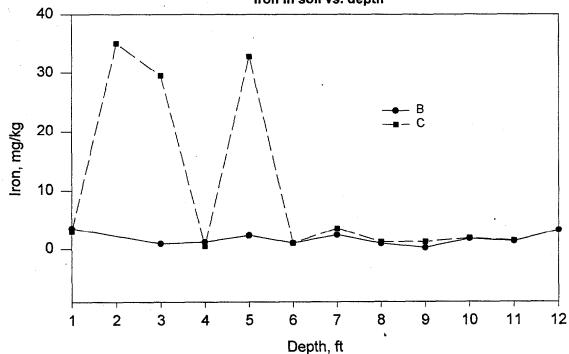


Fig. C.39. Test cell 7 - post-treatment soil iron levels.

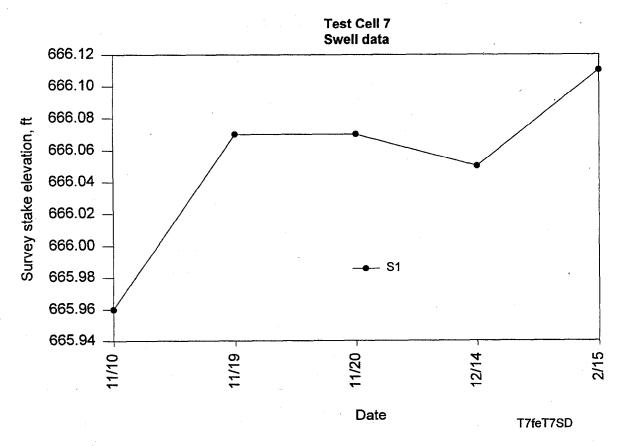
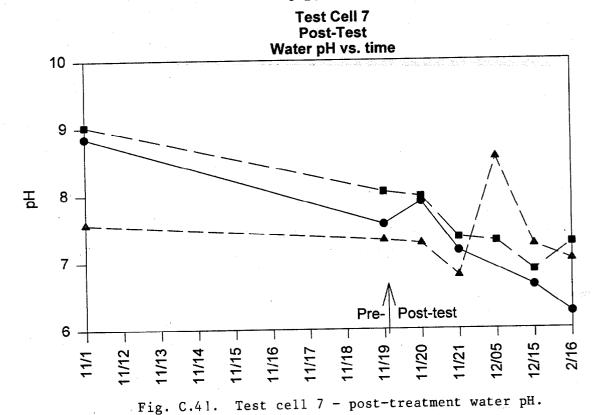


Fig. C.40. Test cell 7 - post-treatment swell data.



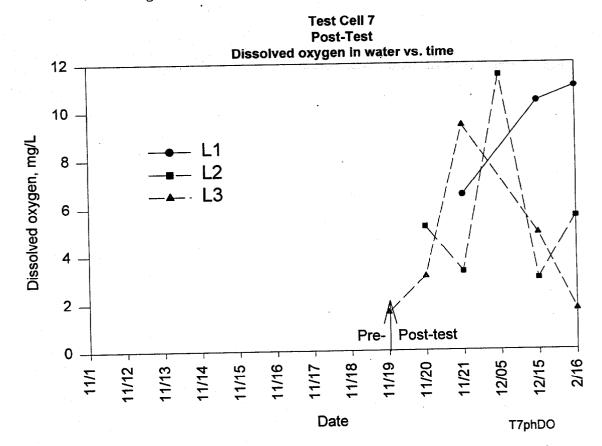


Fig. C.42. Test cell 7 - post-treatment water DO levels.

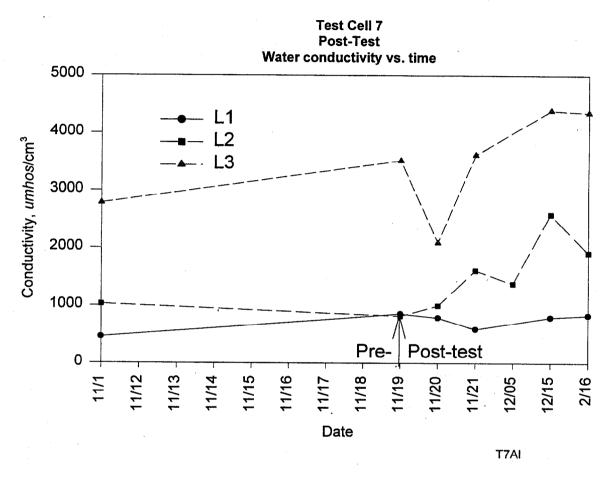


Fig. C.43. Test cell 7 - post-treatment water conductivity levels.

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